Supplementary Material

Formal allene insertion into amides. Reaction of propargyl magnesium bromide with morpholine amides

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General experimentation

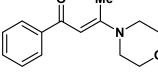
All reactions were performed in round-bottom flasks fitted with rubber septa. Reactions sensitive to air and/or moisture were performed under a positive pressure of argon. Air- and moisture-sensitive liquids were transferred by syringe. Analytical thin-layer chromatography (TLC) was performed using aluminum plates precoated with silica gel (silica gel 60 F₂₅₄, Sorbfil). TLC plates were visualized by exposure to 254 nm ultraviolet light (UV) or were stained by submersion in acidic ethanolic solution of vanillin followed by brief heating (vanillin) or submersion in aqueous potassium permanganate solution followed by extensive washing with water (KMnO₄). Flash-column chromatography was carried out on silica gel (60 Å, 230–400 mesh, Merck). All solvents for chromatography and extractions were technical grade and distilled prior use.

All reagents were obtained from commercial suppliers and were used without further purification. Et₂O and THF were stored over sodium benzophenone ketyl and were distilled directly prior use.

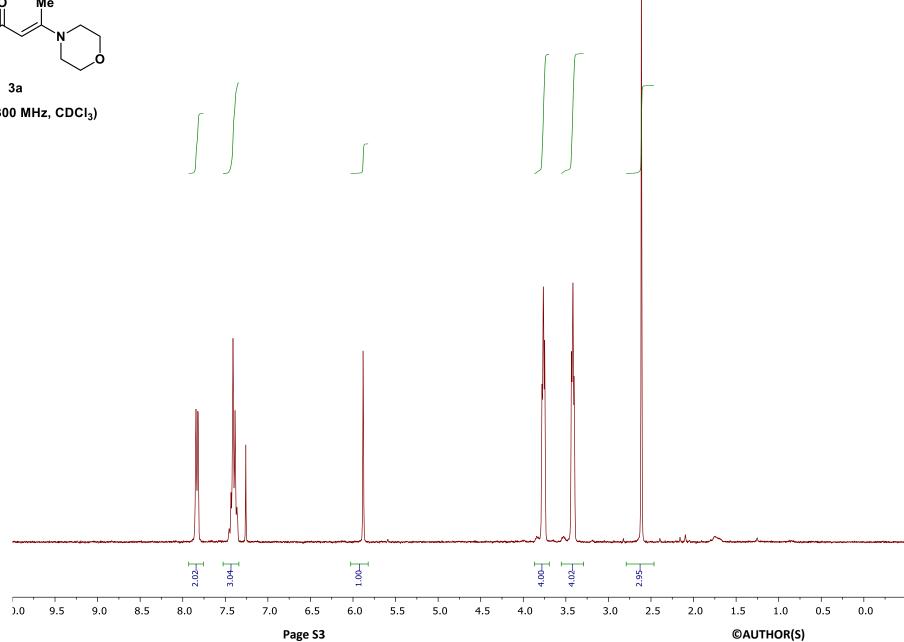
Nuclear magnetic resonance spectra were recorded using Bruker Furier 300, Bruker Avance 800 instruments at indicated temperature. Data are represented as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet and/or multiple resonances), coupling constant (J) in Hertz, integration. Proton chemical shifts are expressed in parts per million (ppm, δ scale) and are referenced to residual protium in the NMR solvents (CHCl₃, δ 7.26 ppm). Carbon chemical shifts are expressed in parts per million (ppm, δ scale) and are referenced to the carbon resonances of the NMR solvents (CDCl₃, δ 77.16 ppm).

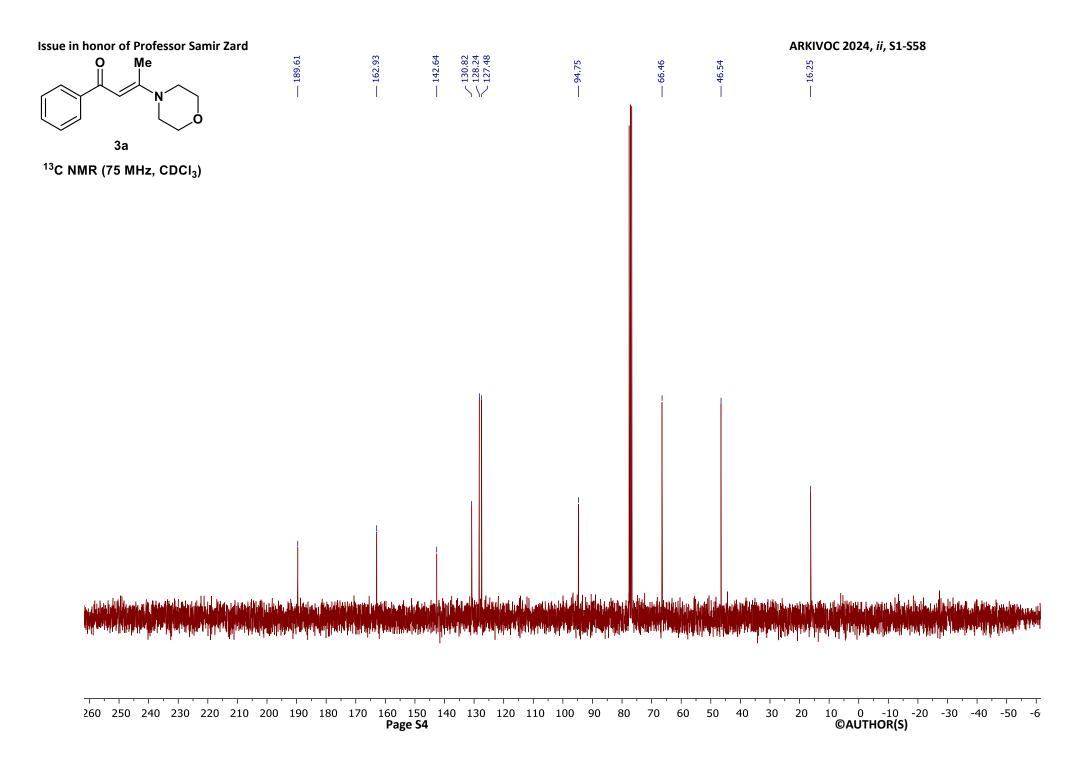
High-resolution mass spectra were recorded on a Bruker micrOTOF-Q II mass spectrometer using electrospray ionization (ESI-TOF). Melting points were determined on Kofler melting point apparatus and are uncorrected.

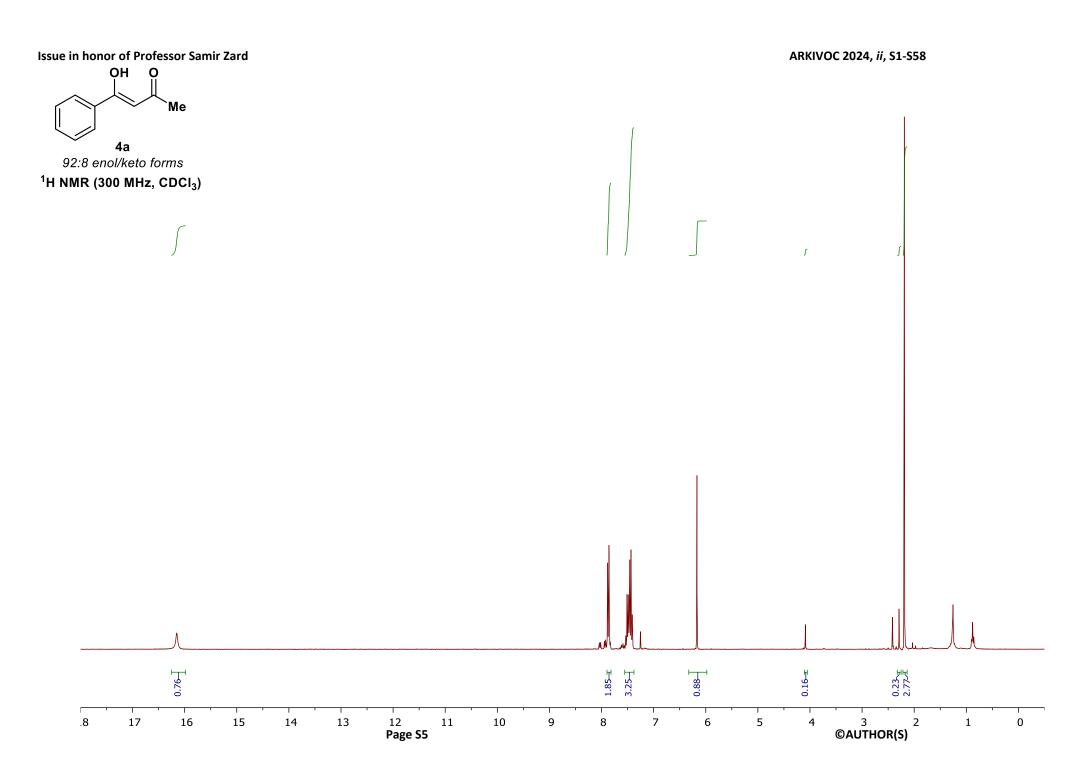
Copies of NMR spectra

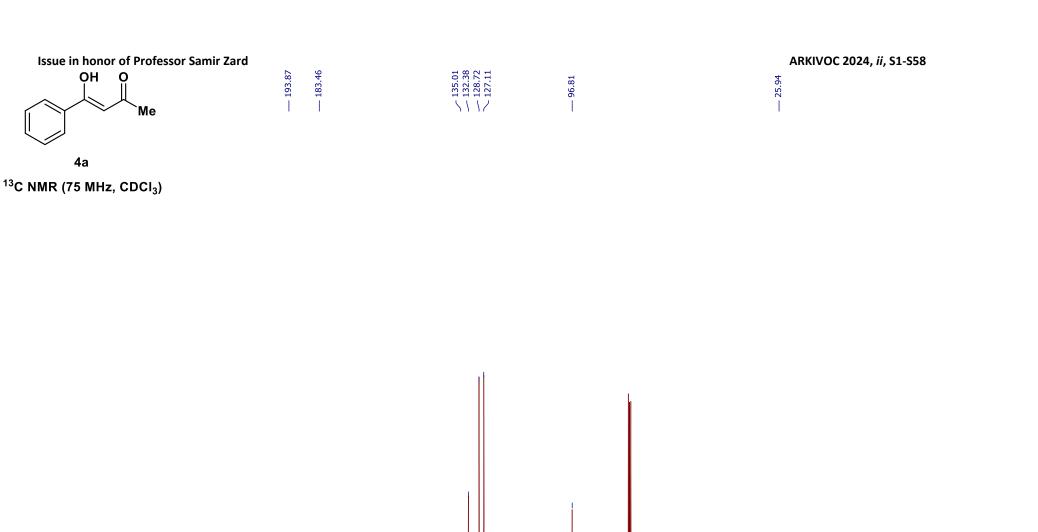


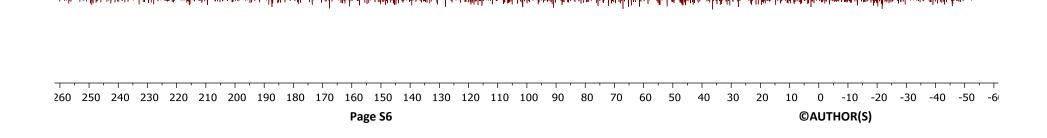
¹H NMR (300 MHz, CDCl₃)

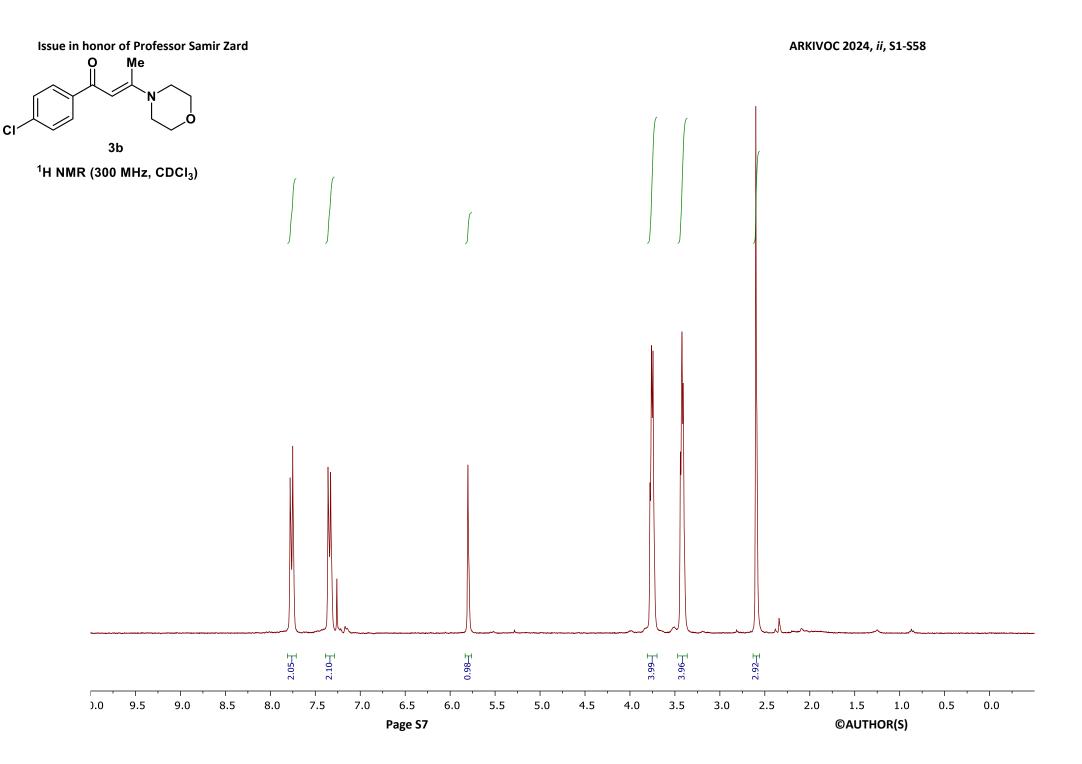


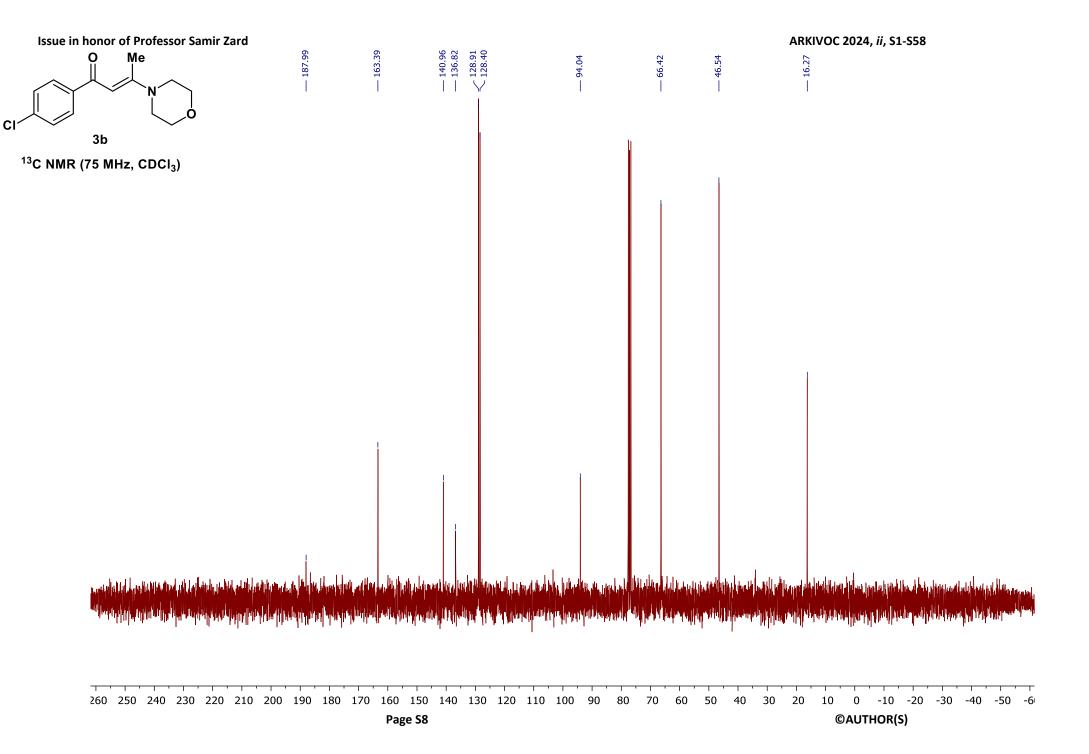


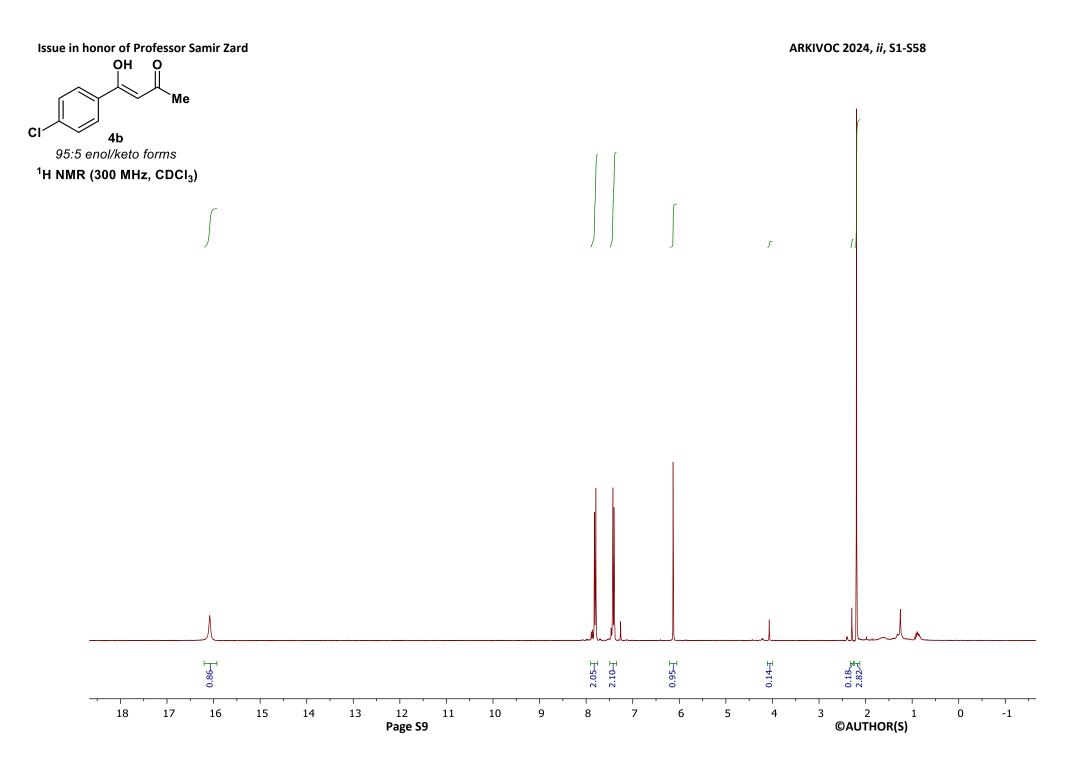


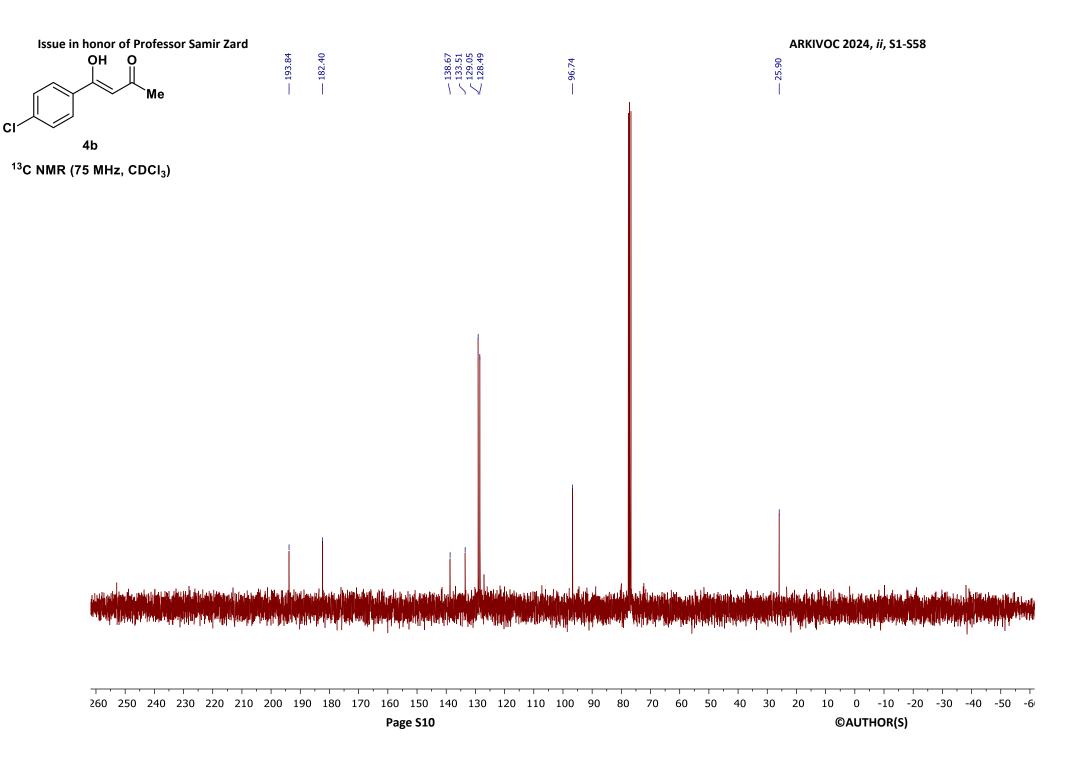


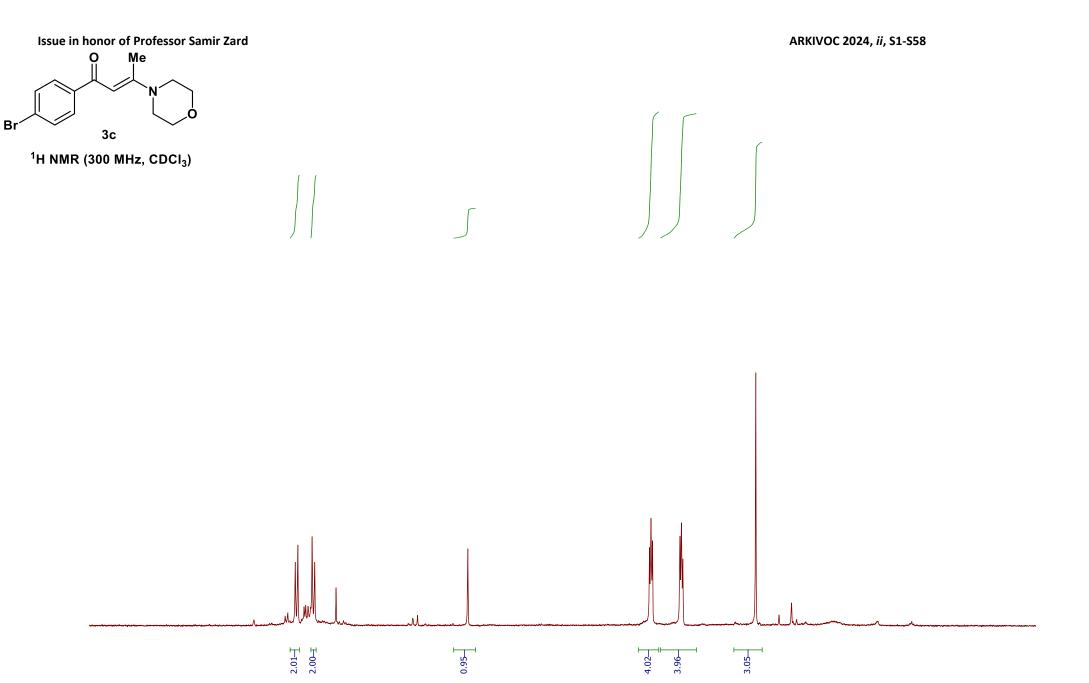












0.0

9.5

9.0

8.5

8.0

7.5

7.0

6.5

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6.0

5.5

5.0

4.5

4.0

3.5

3.0

2.5

2.0

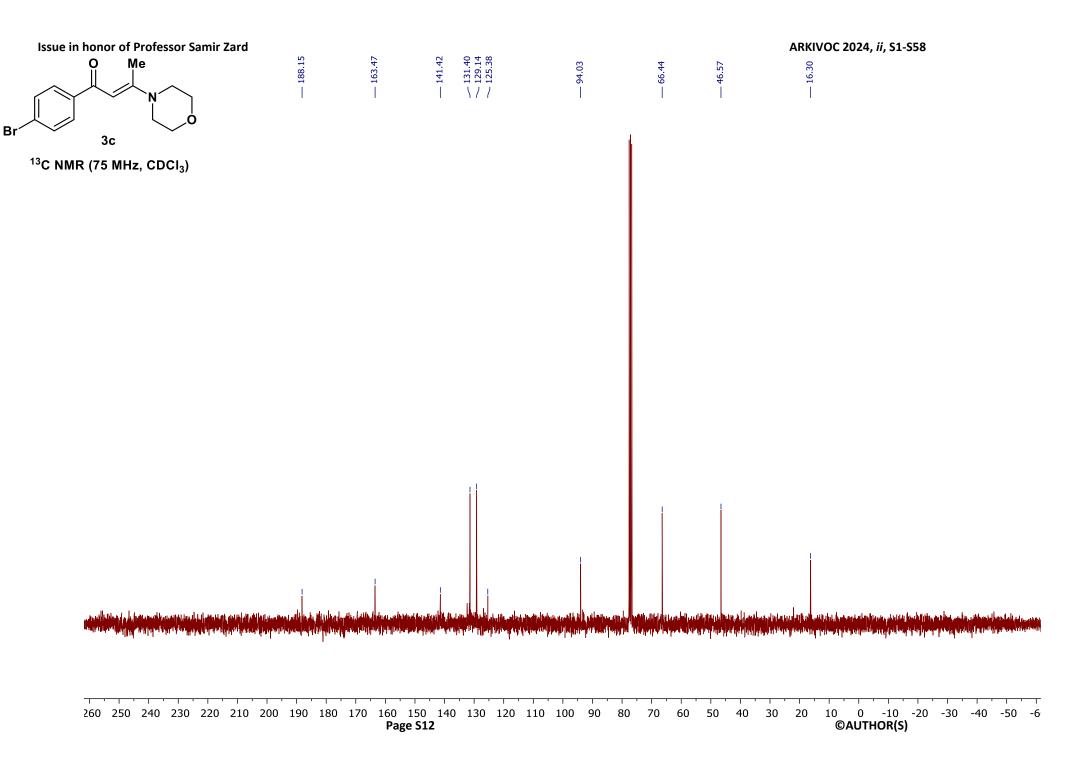
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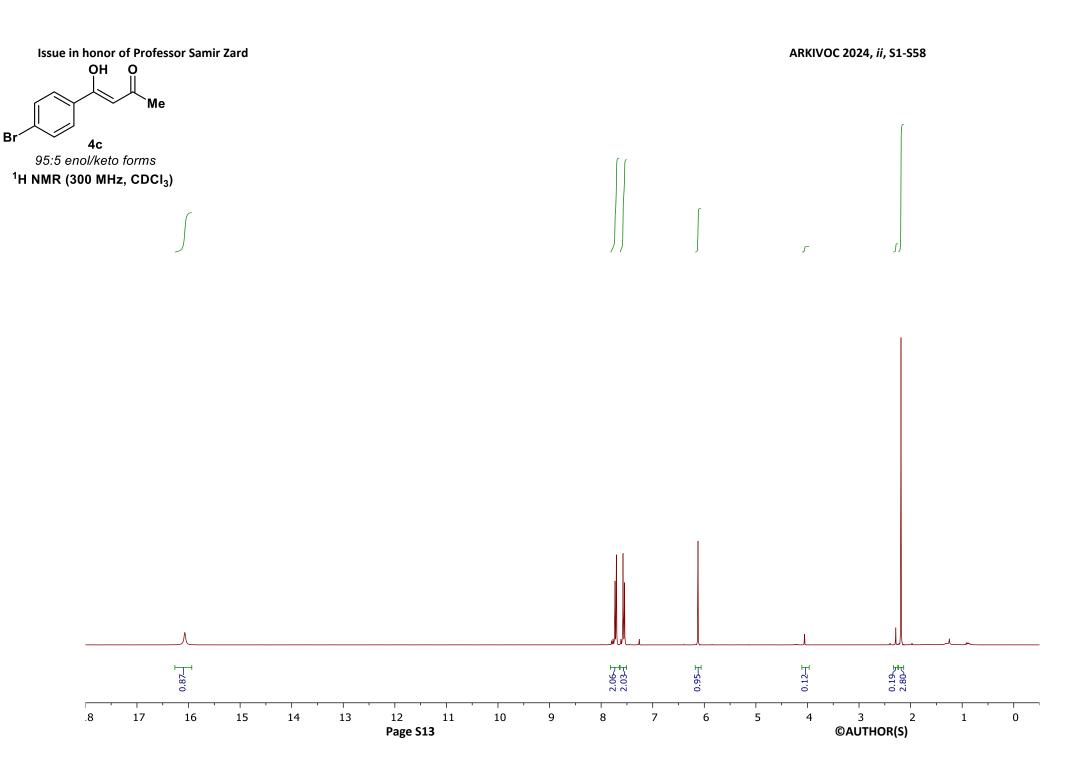
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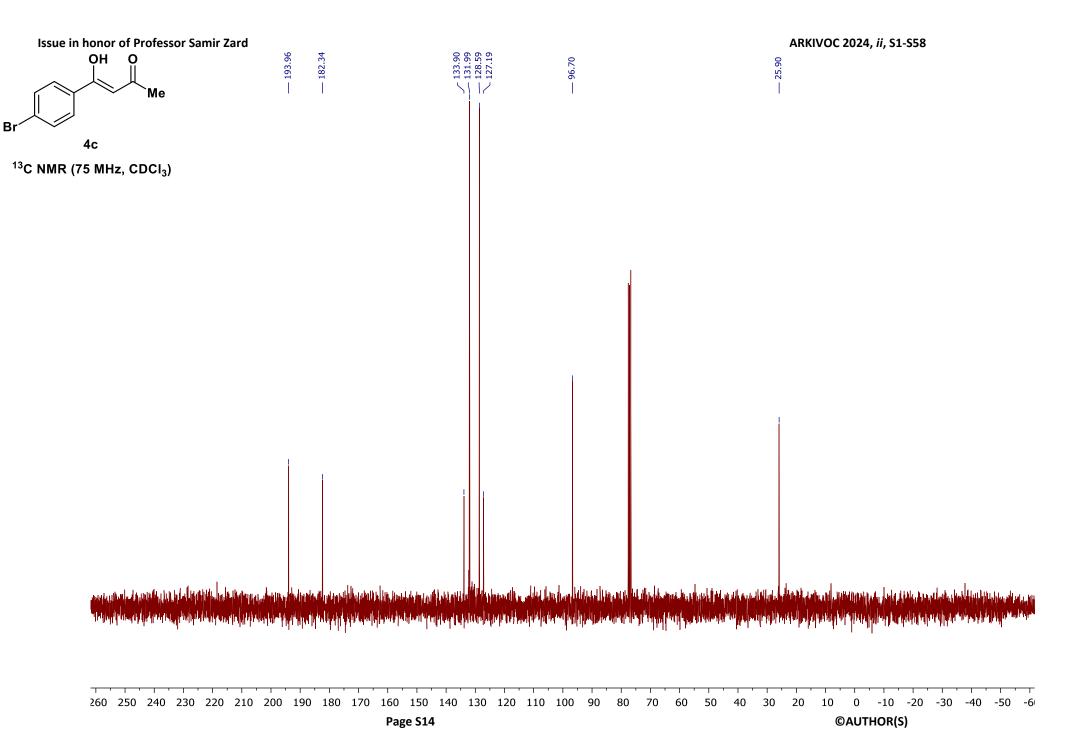
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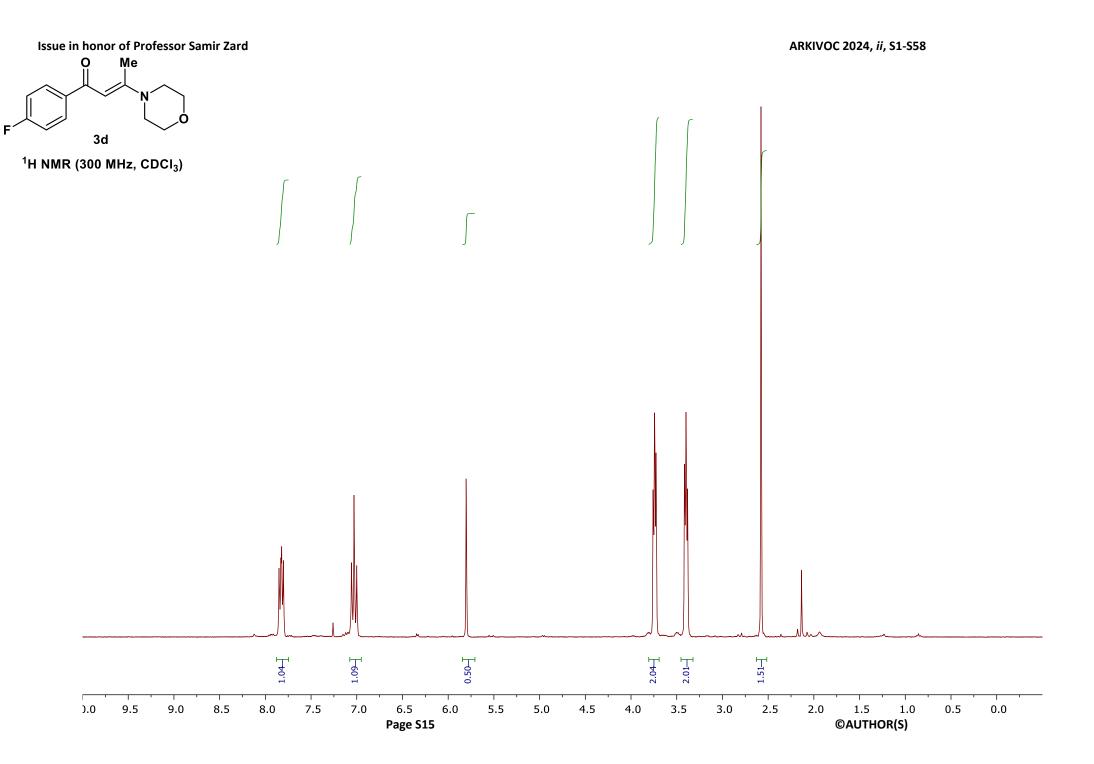
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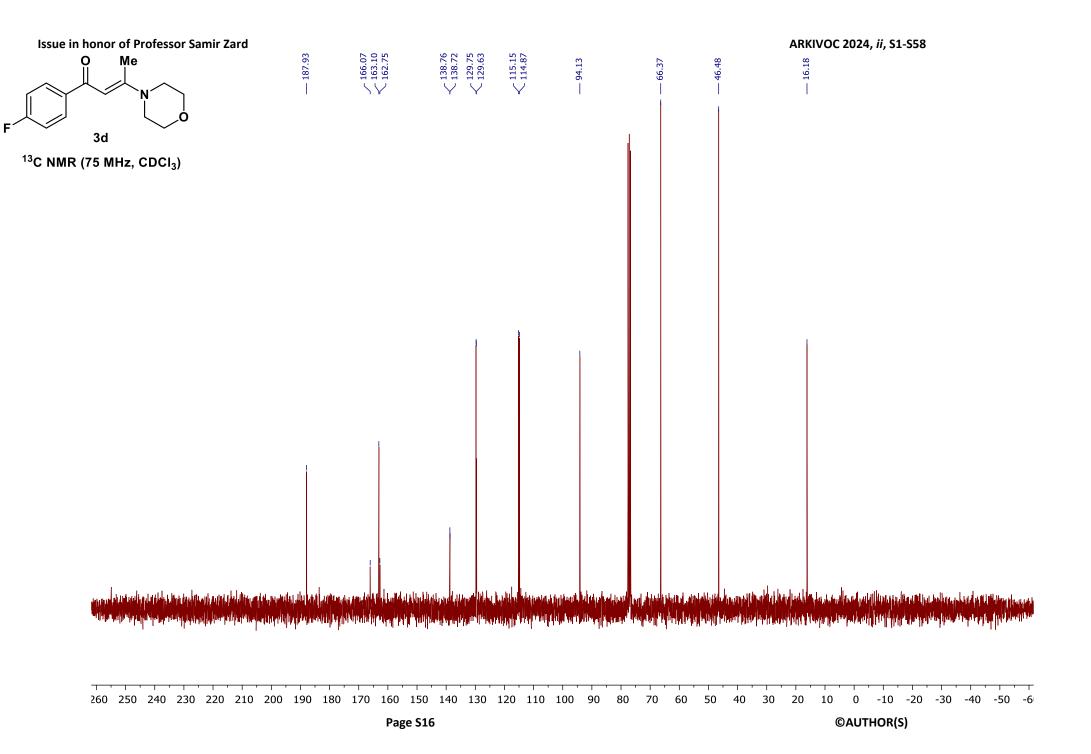
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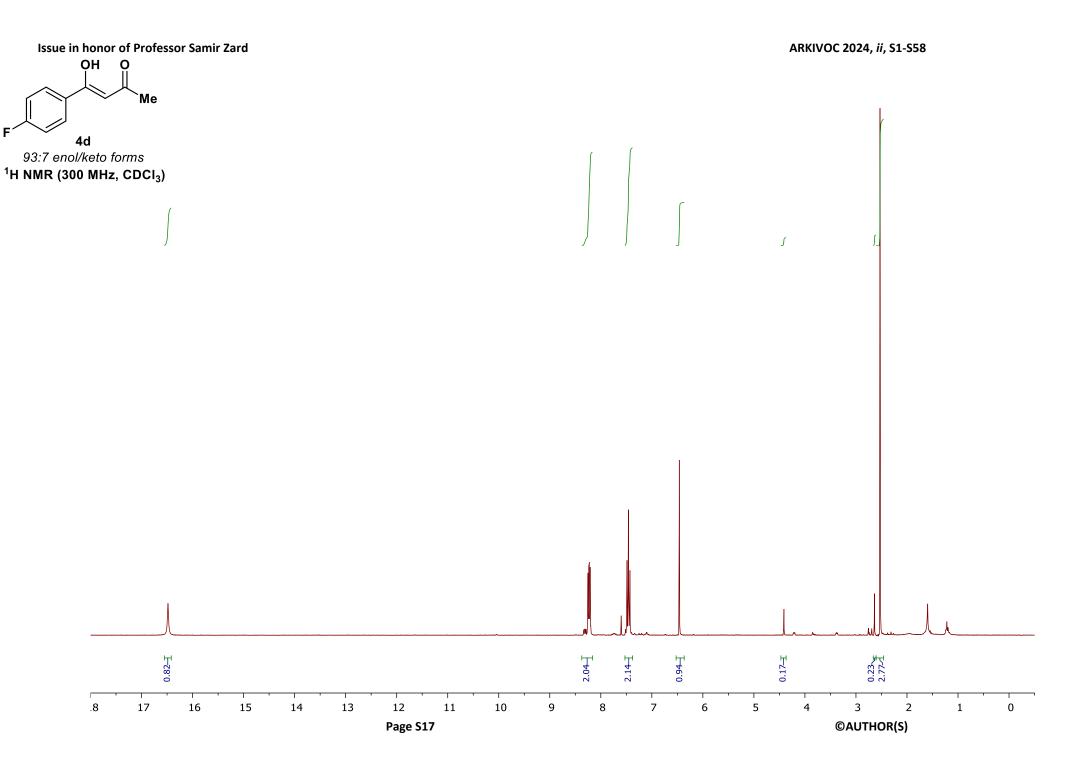


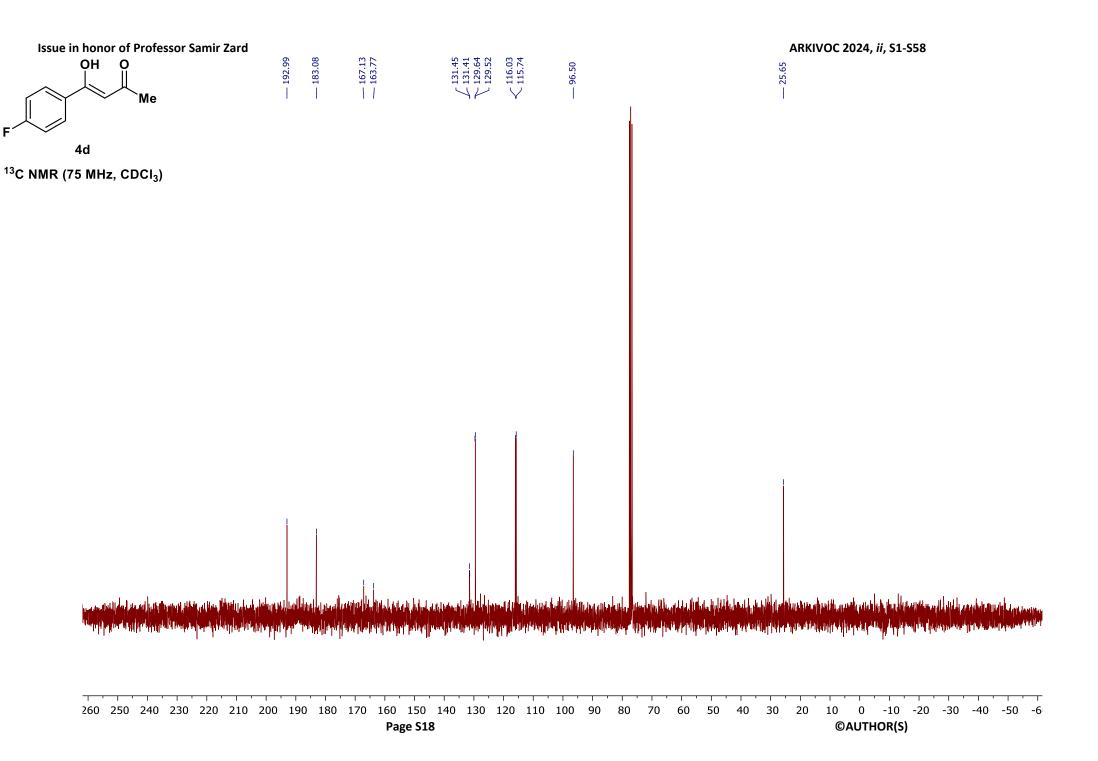


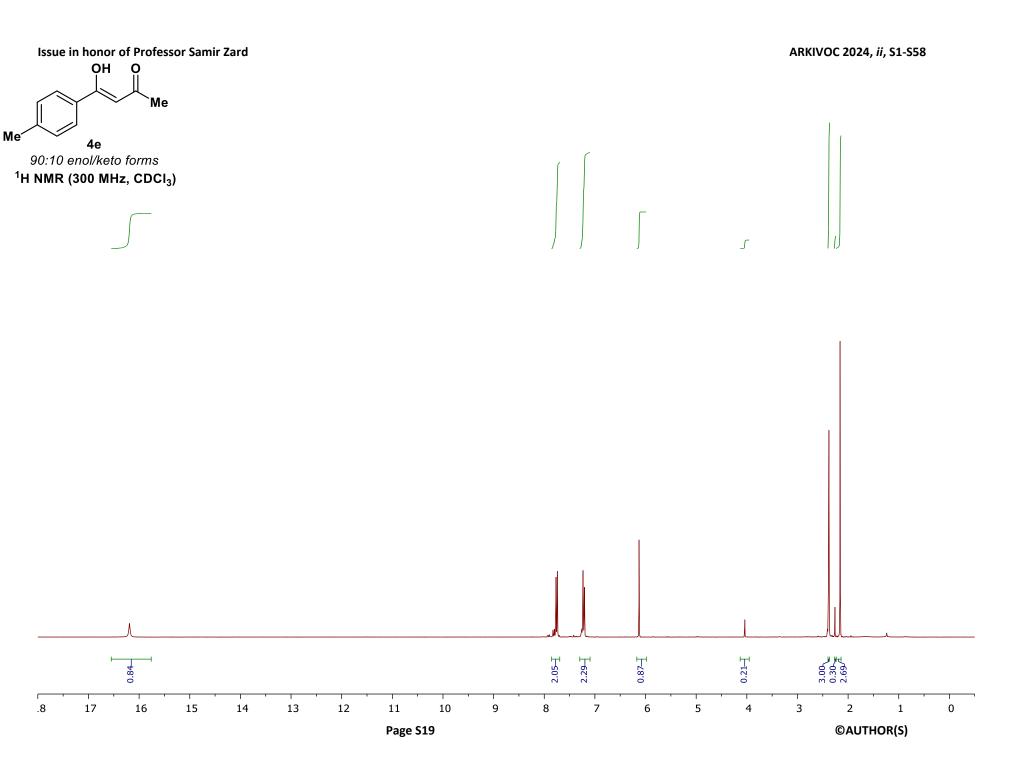


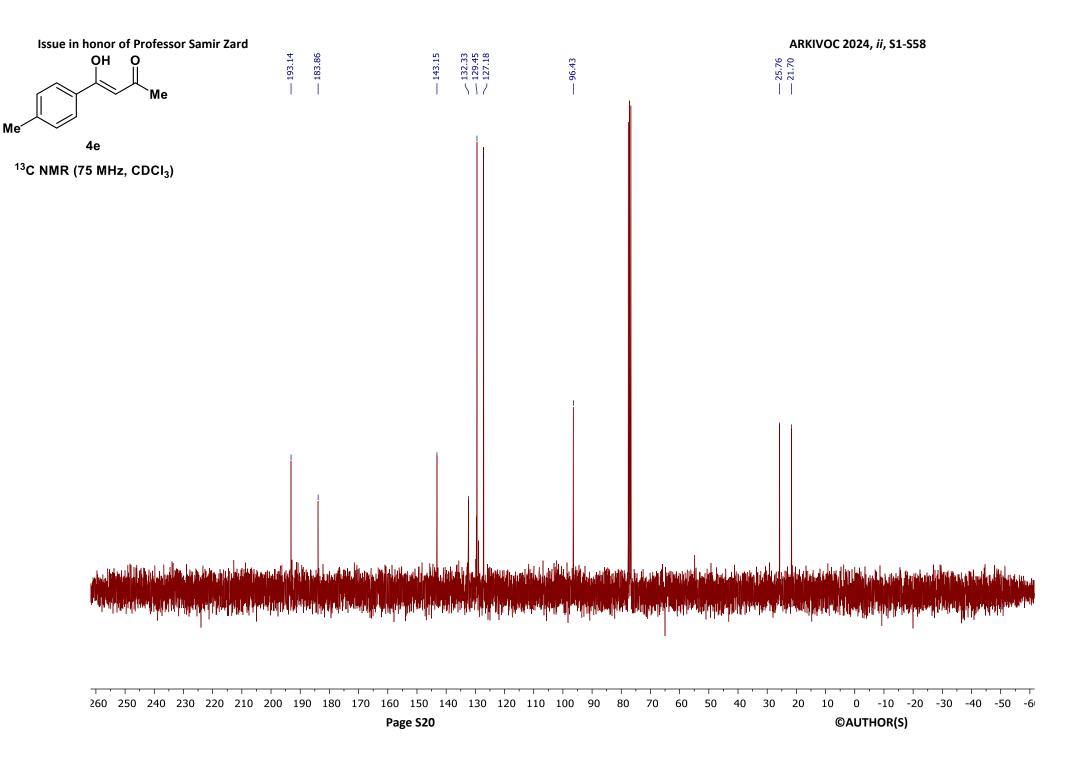


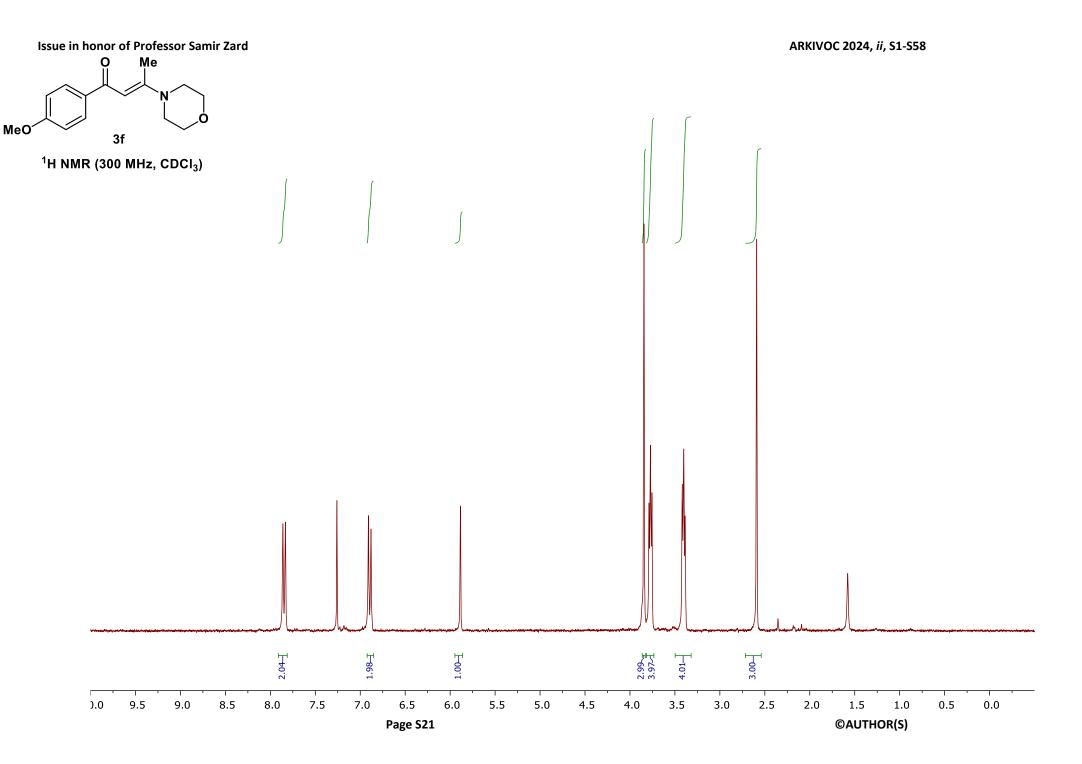








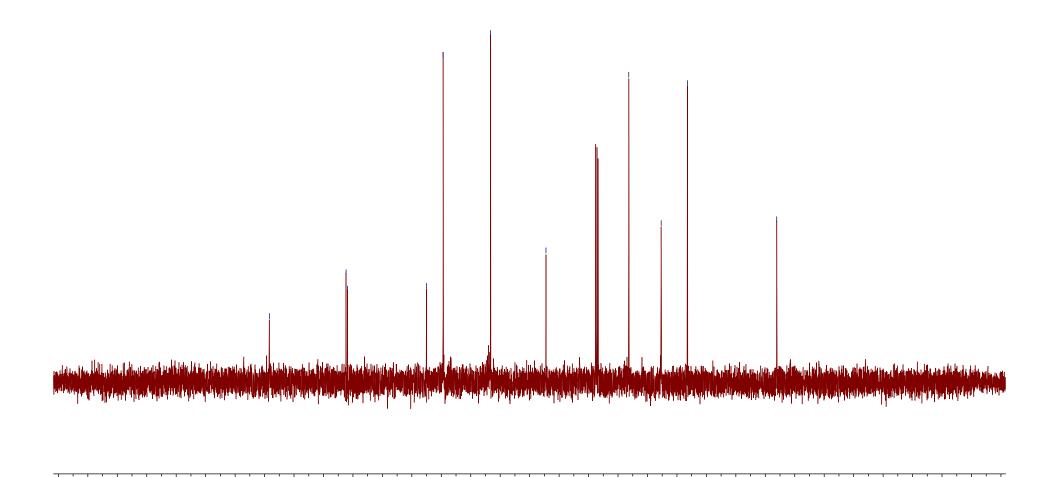






3f

13C NMR (75 MHz, CDCl₃)



80 70

60

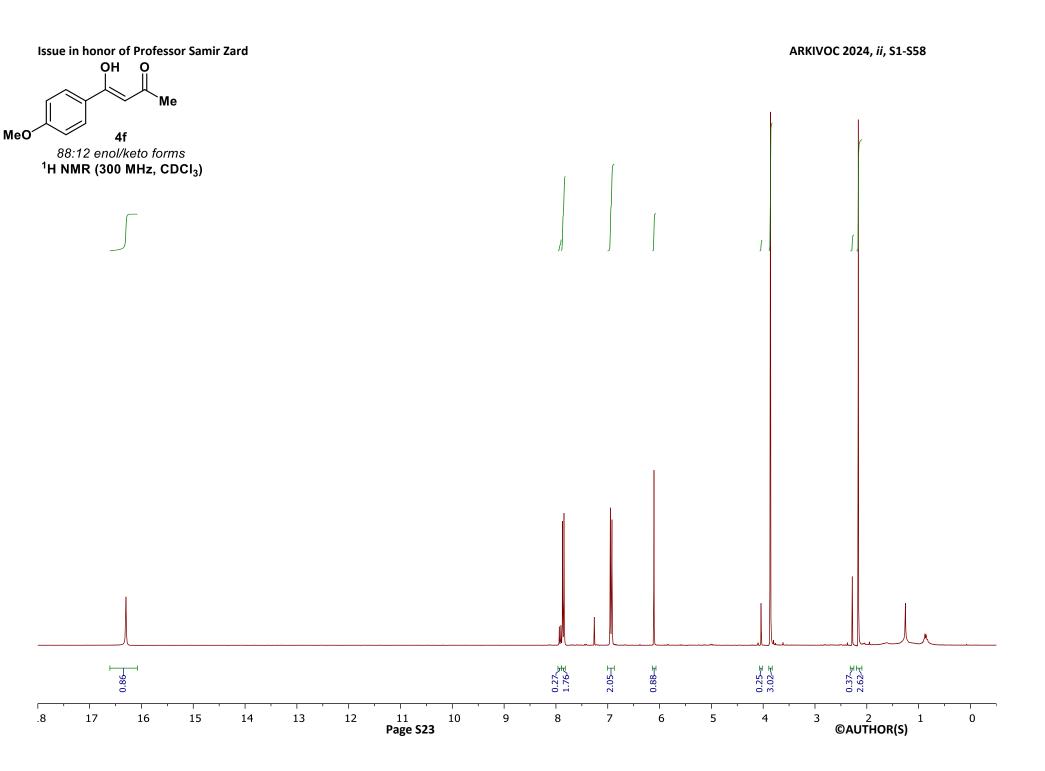
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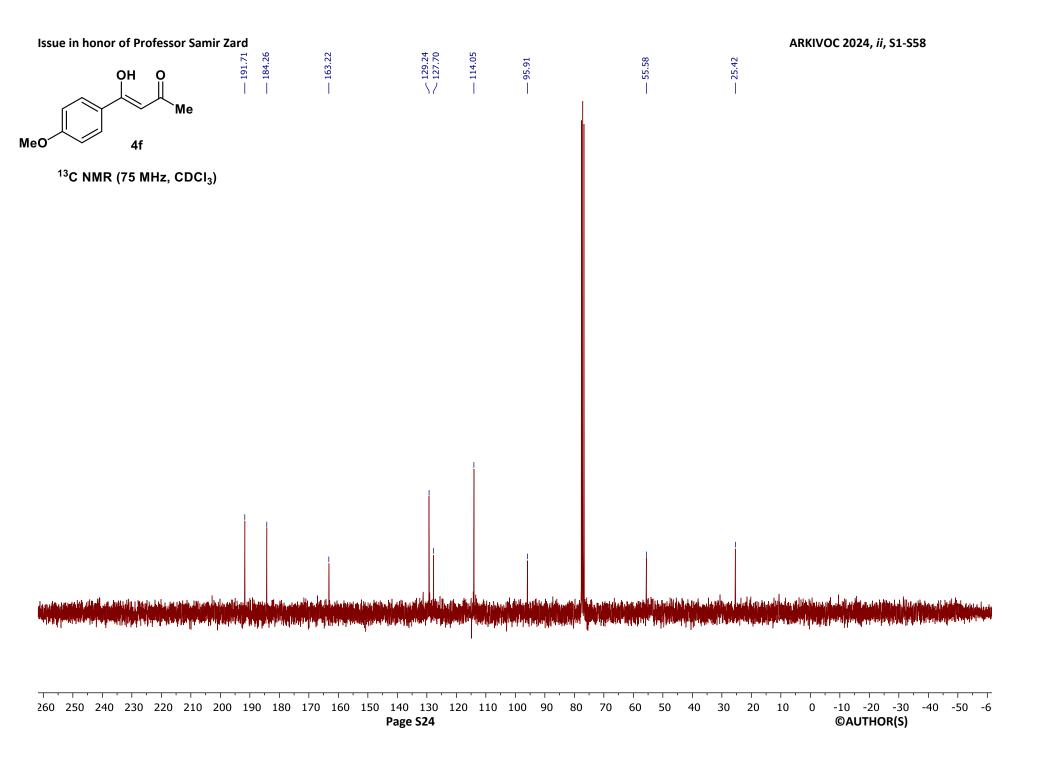
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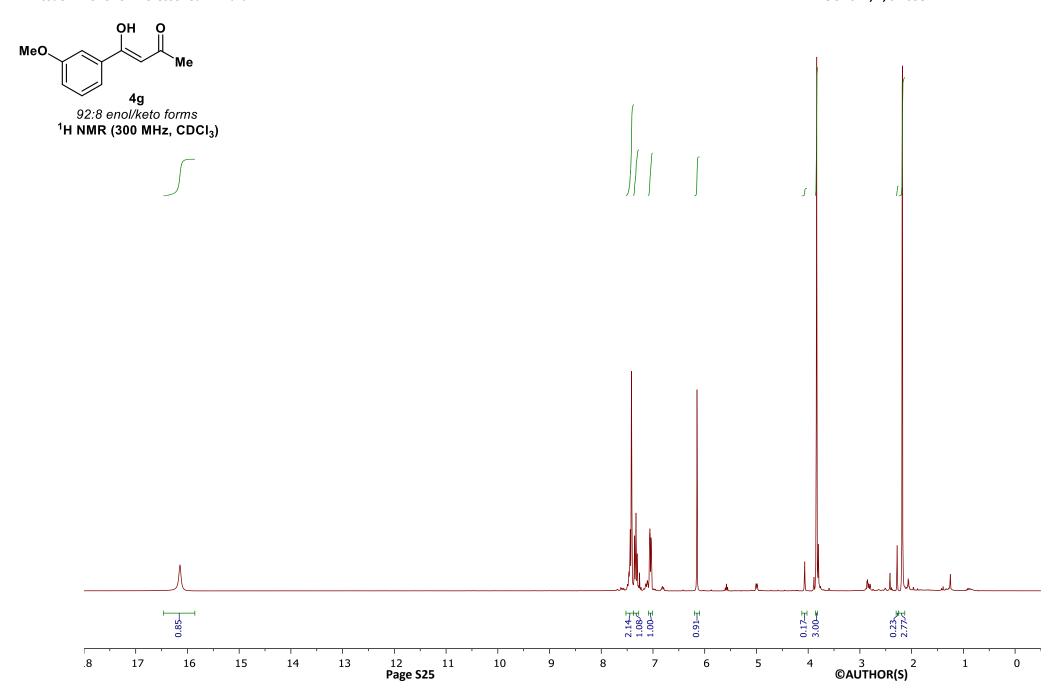
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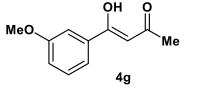




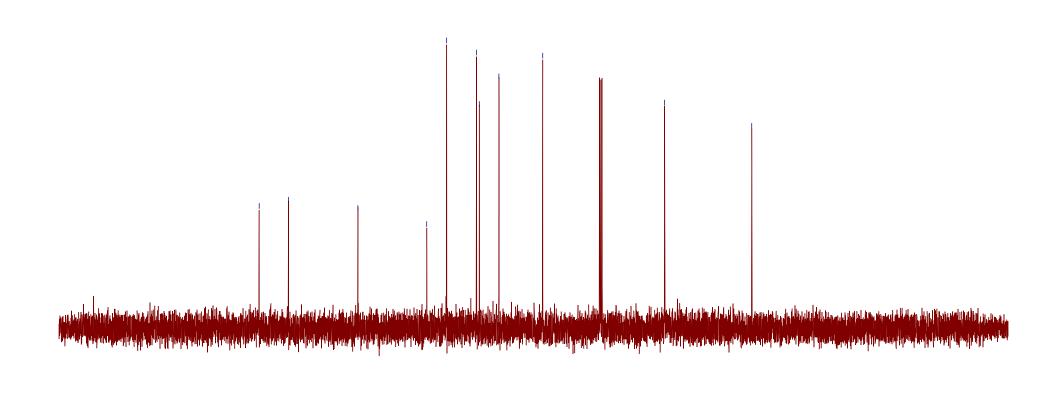


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¹³C NMR (75 MHz, CDCl₃)



80 70

60 50

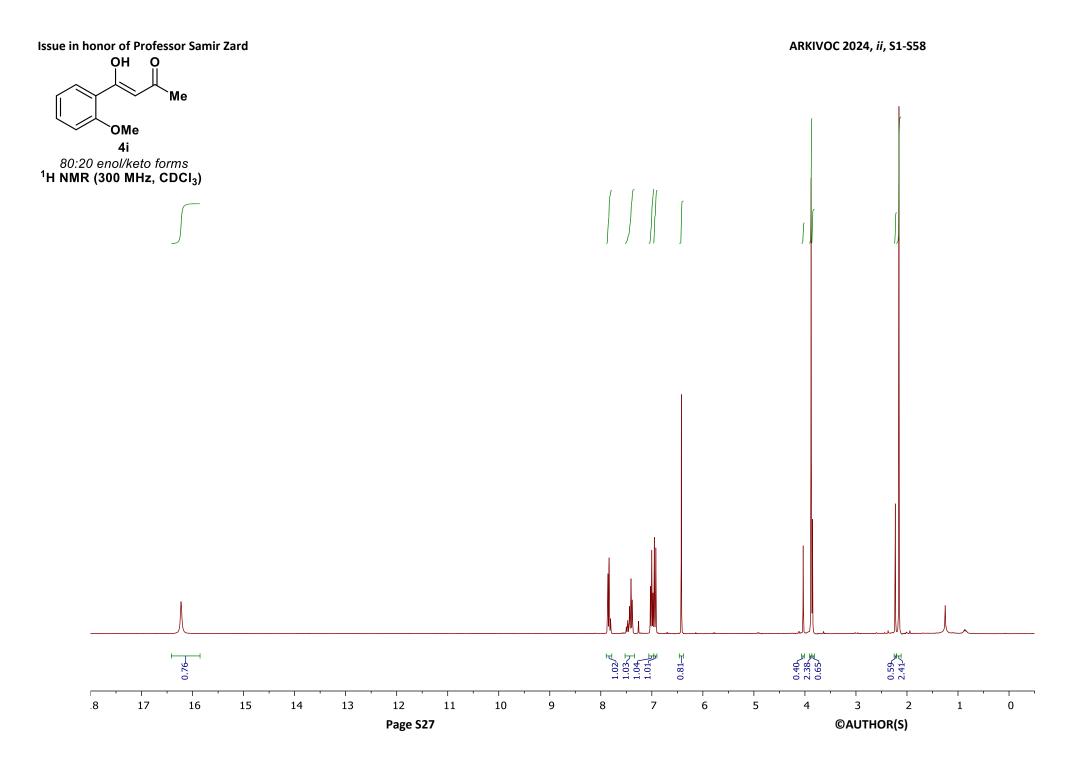
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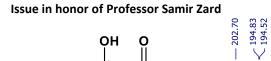
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©AUTHOR(S)

-10 -20 -30 -40 -50 -6

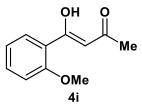




-- 181.35

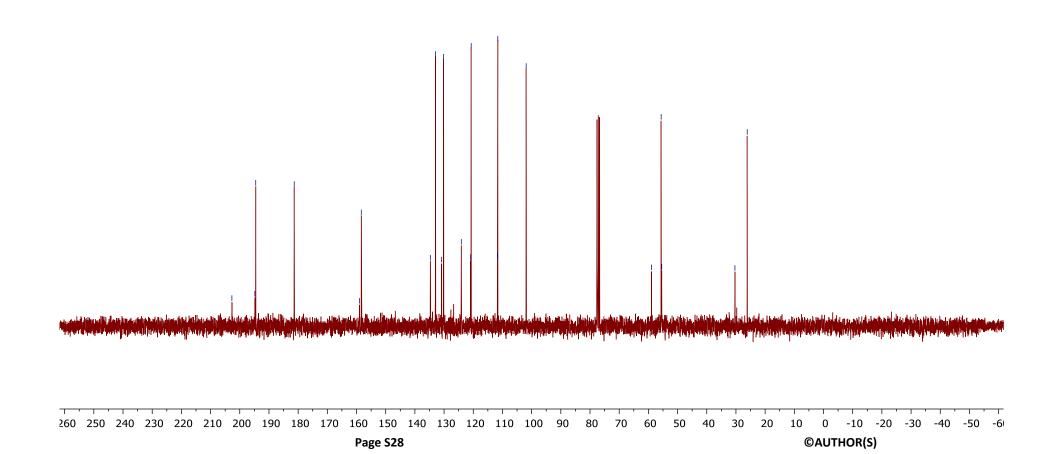
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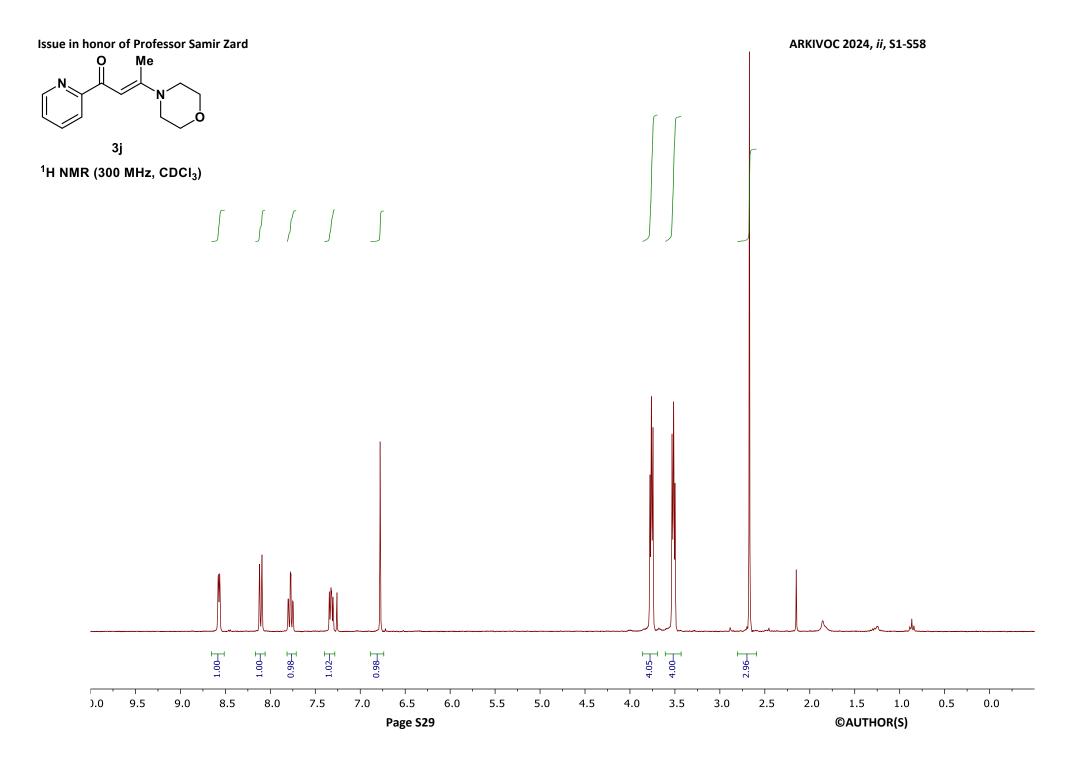
58.93 55.63 55.42 30.34



80:20 enol/keto forms

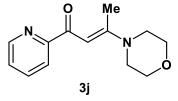
13C NMR (75 MHz, CDCI₃)



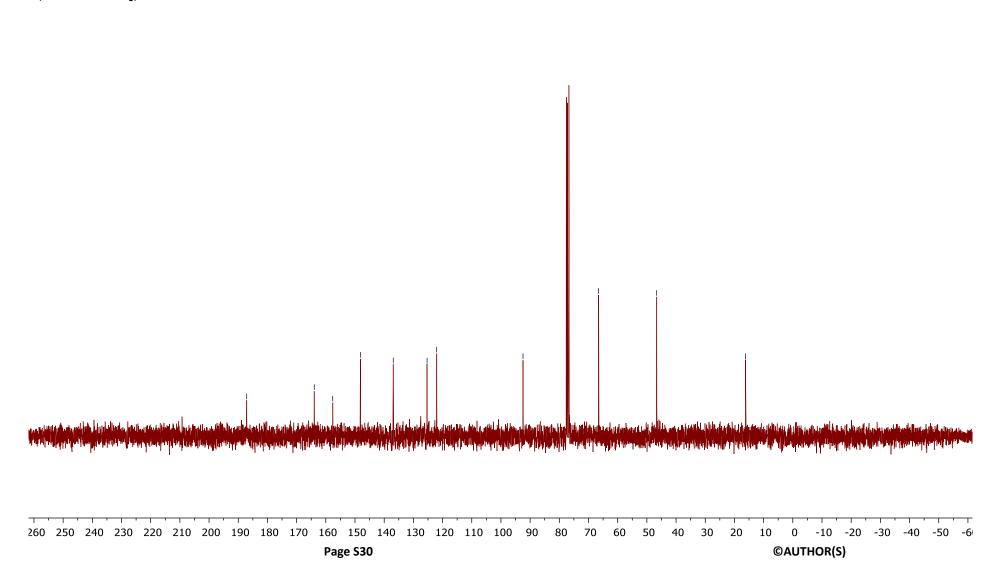




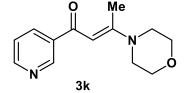
— 163.96 — 157.62 - 148.14



¹³C NMR (75 MHz, CDCl₃)

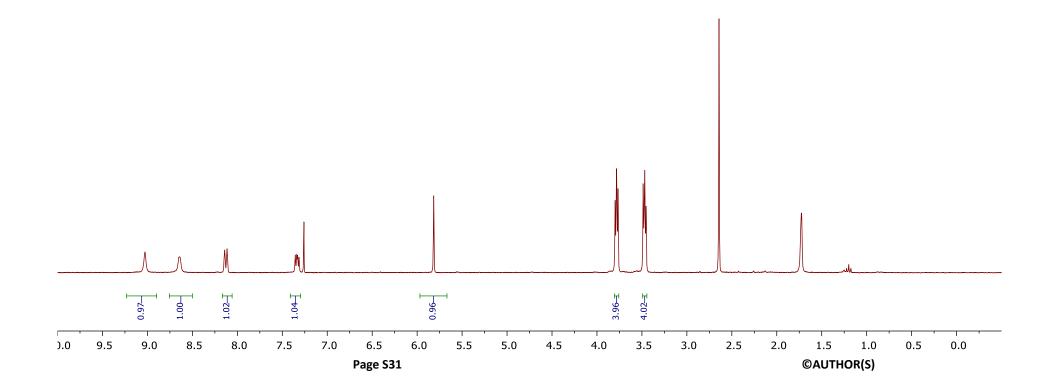


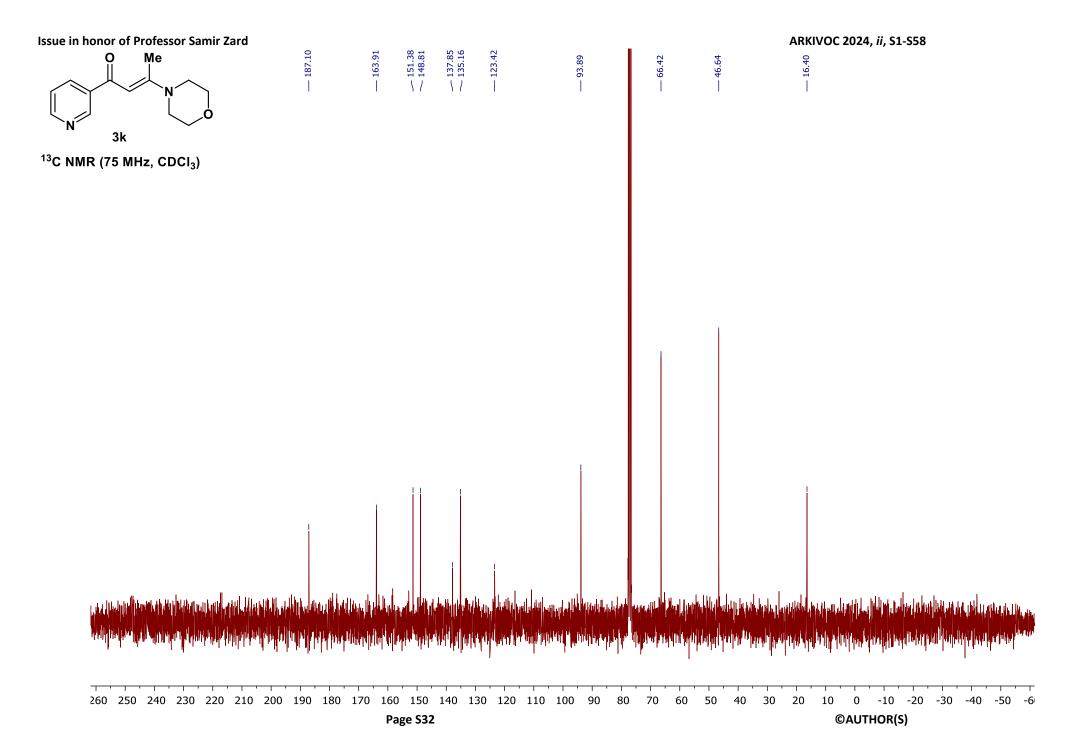
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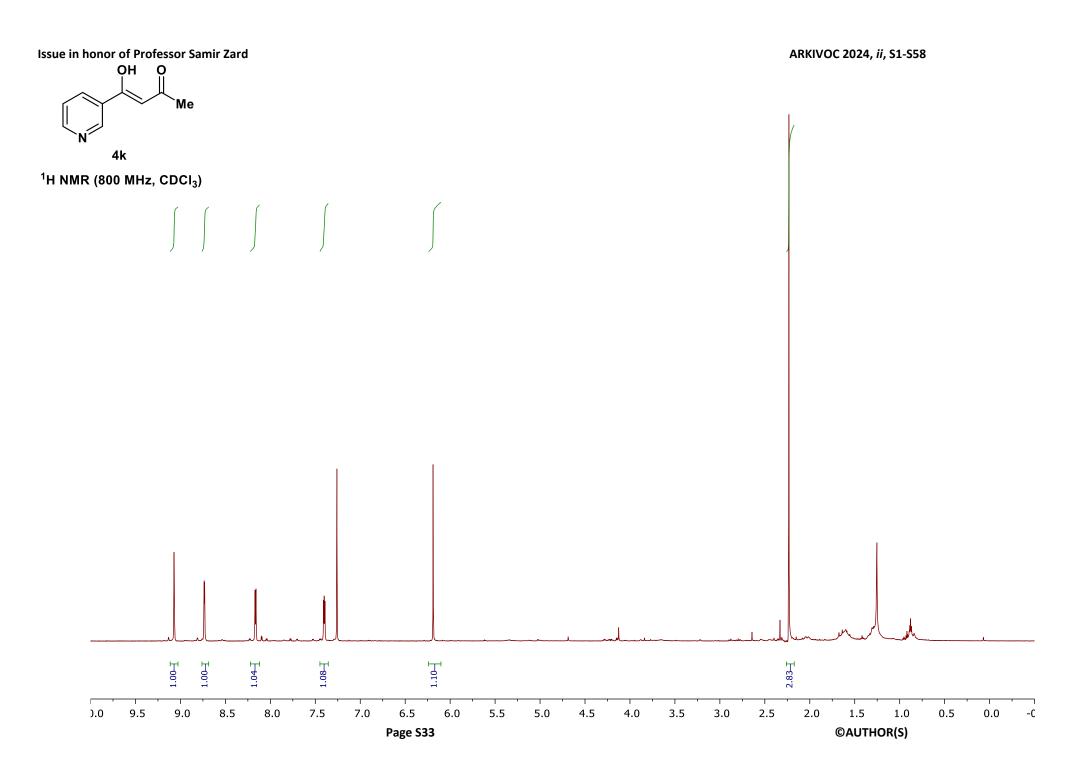


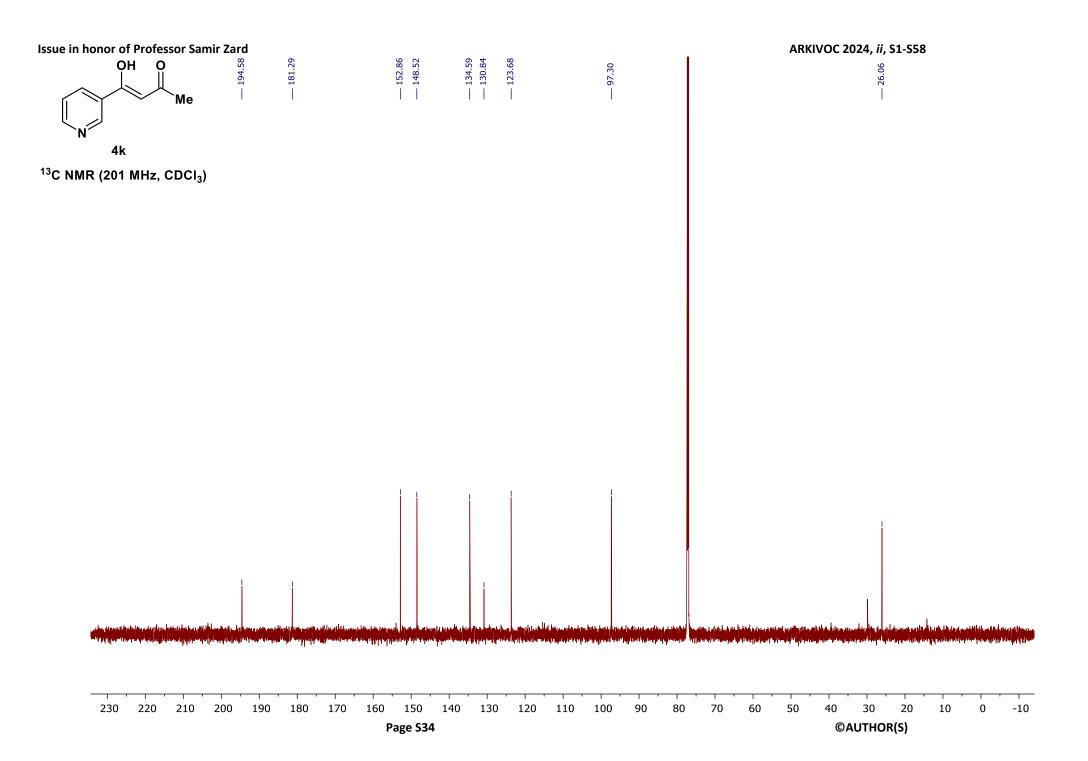
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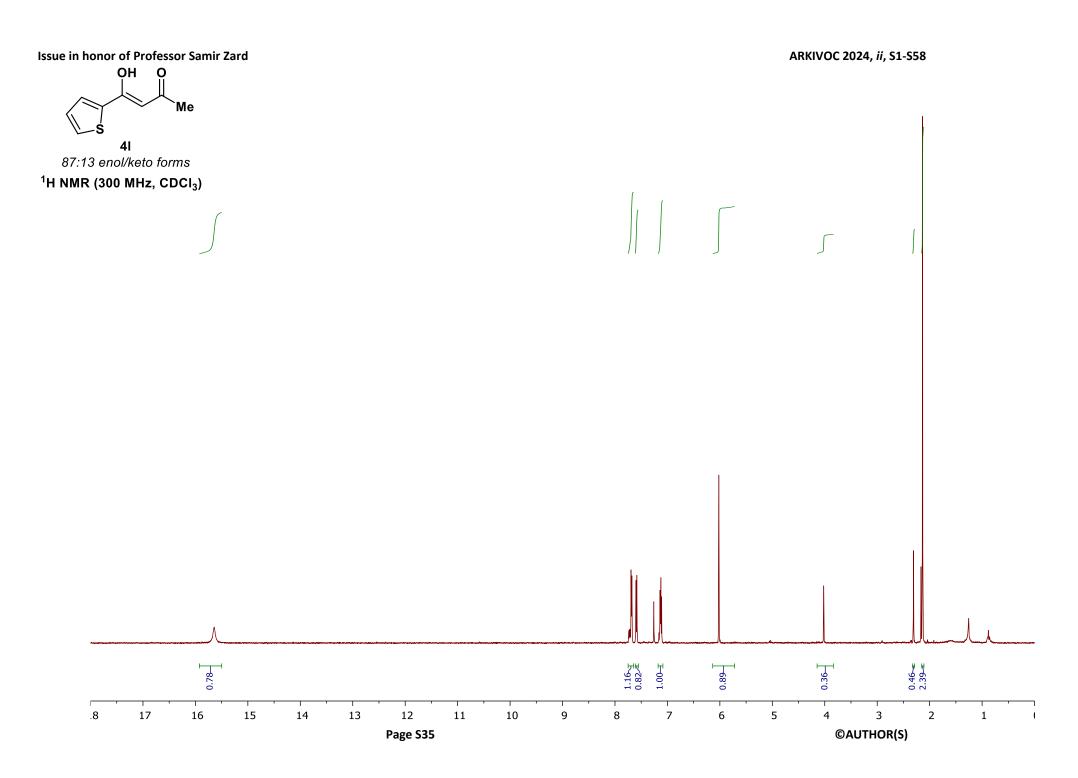


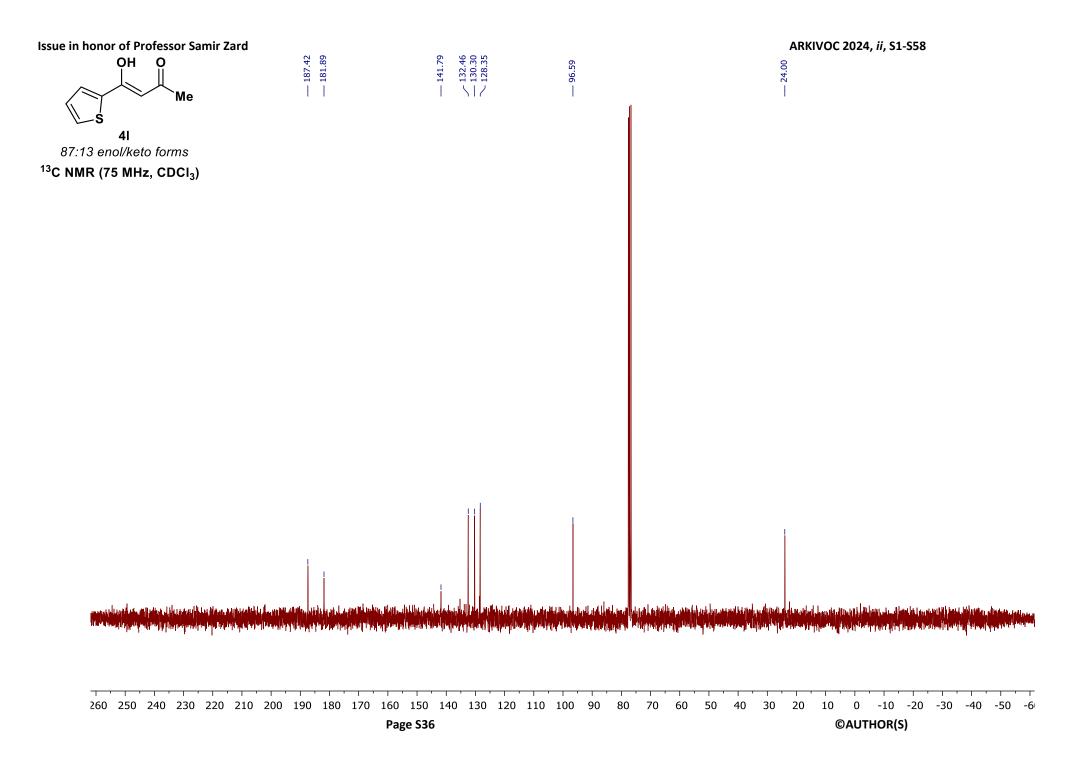


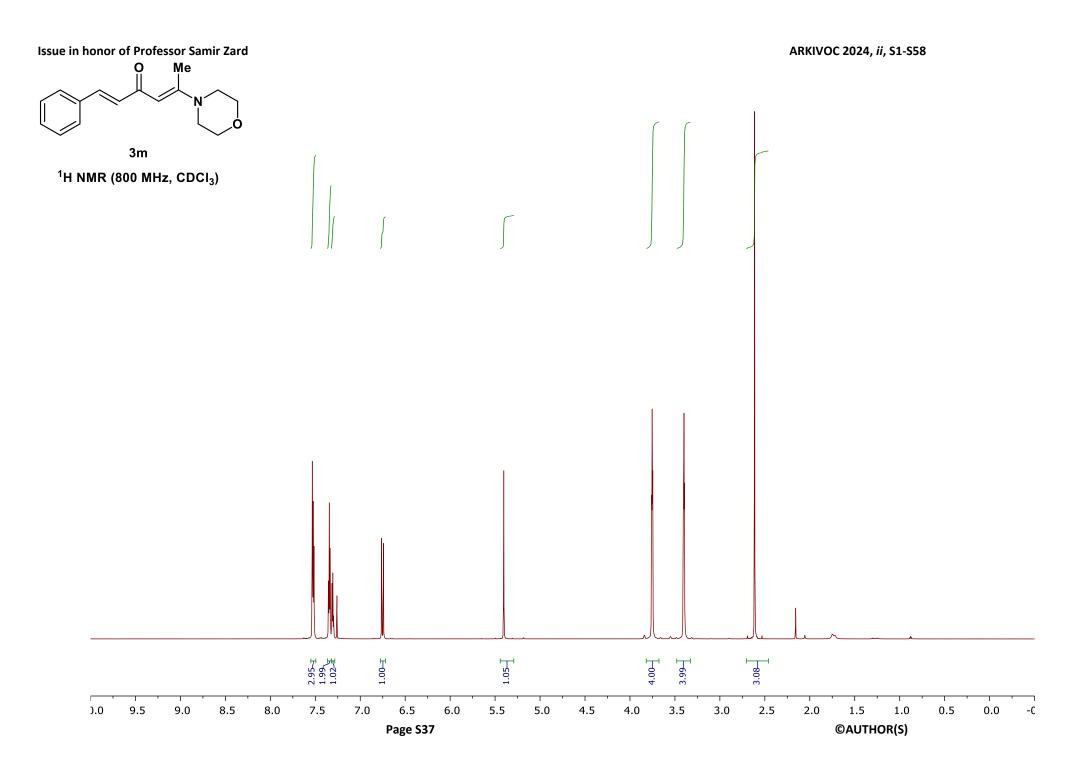


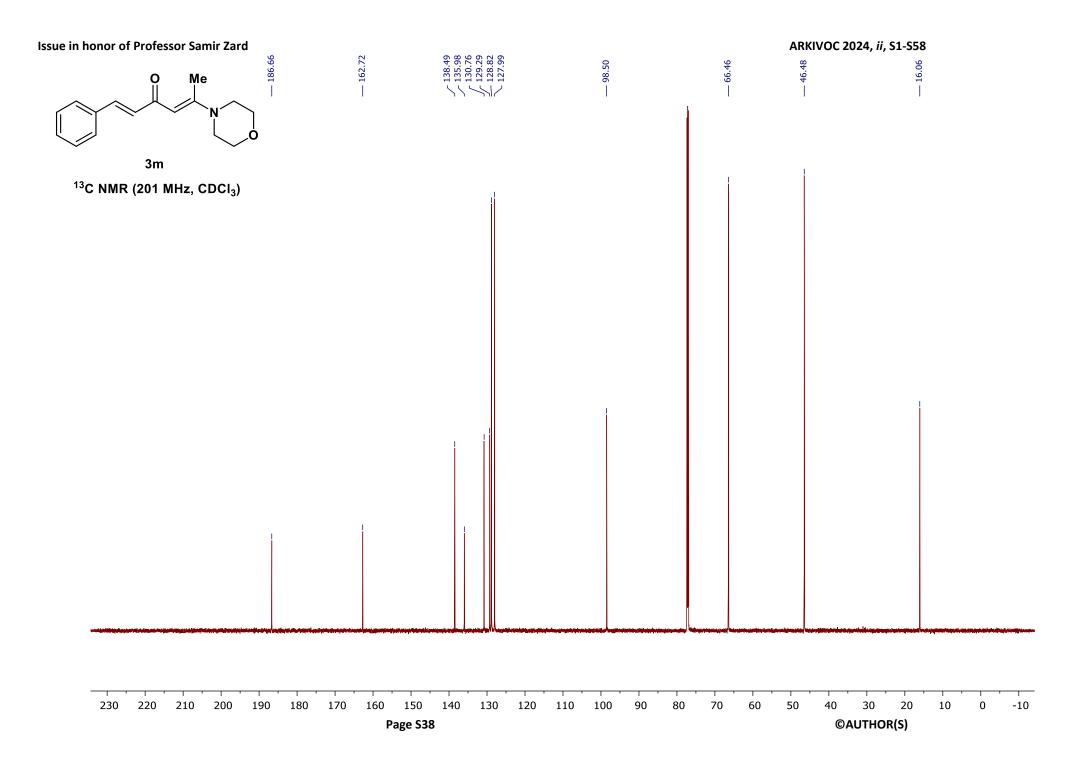


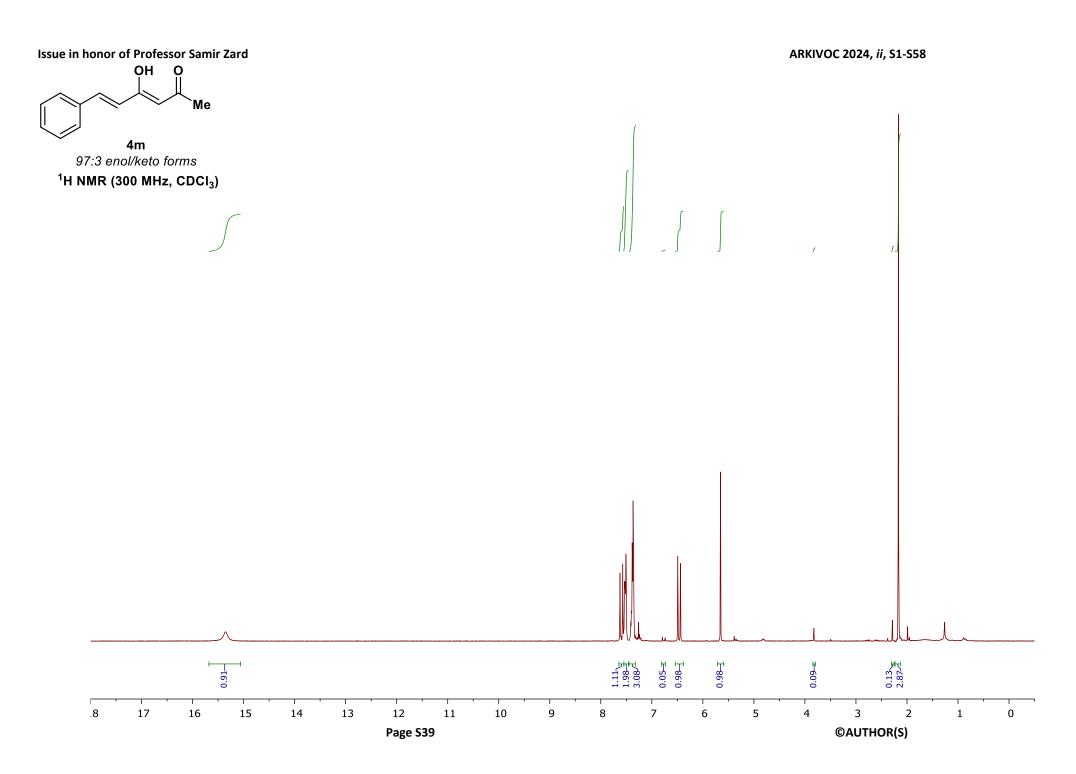


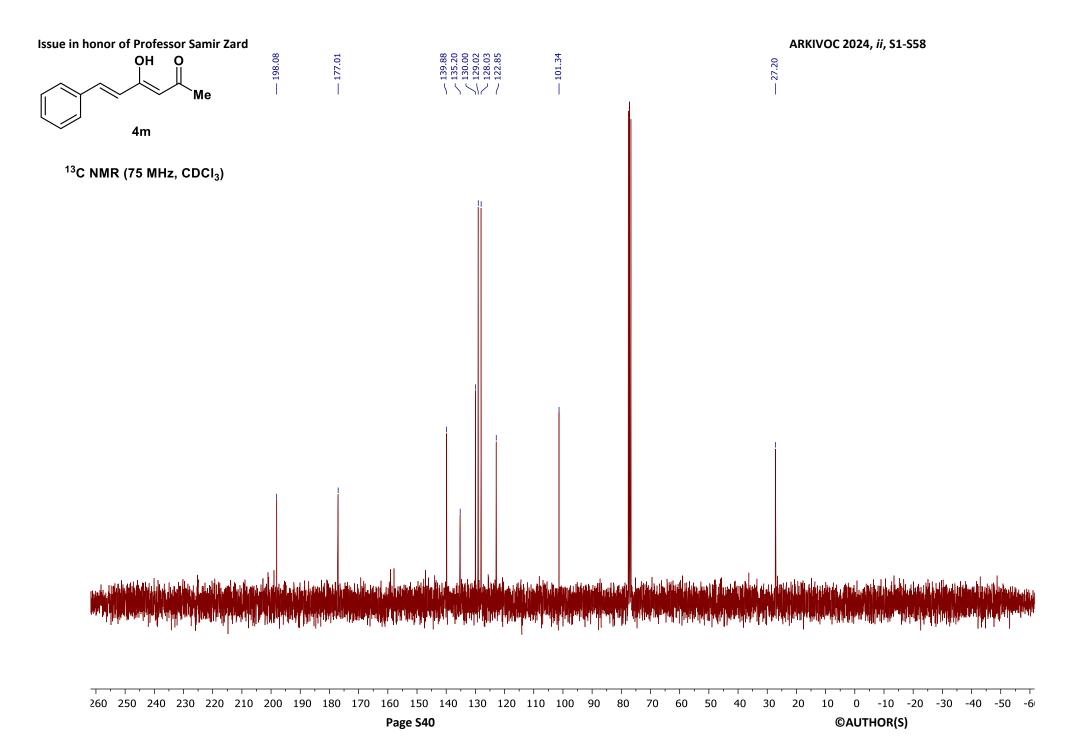


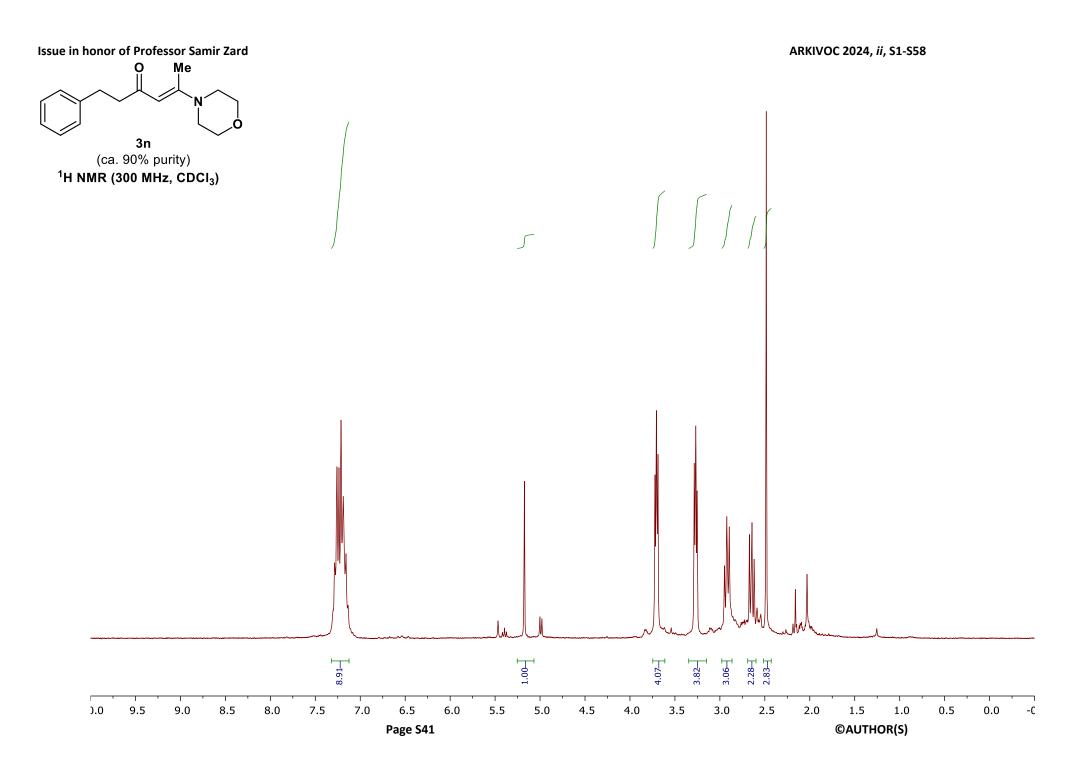


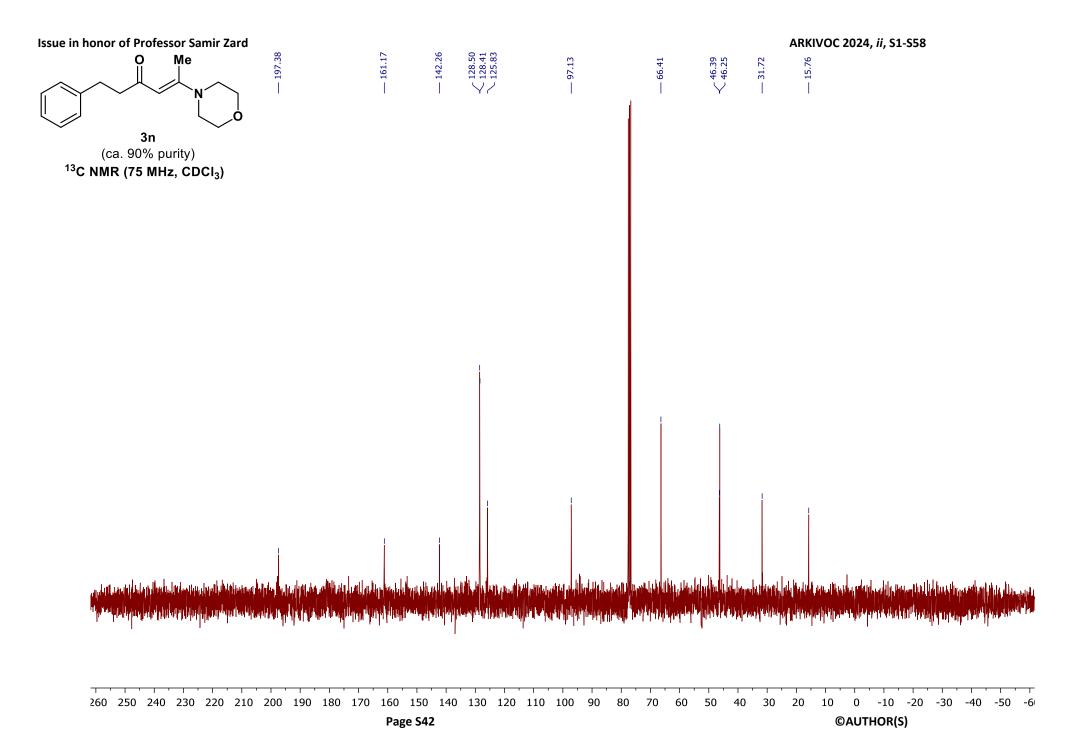


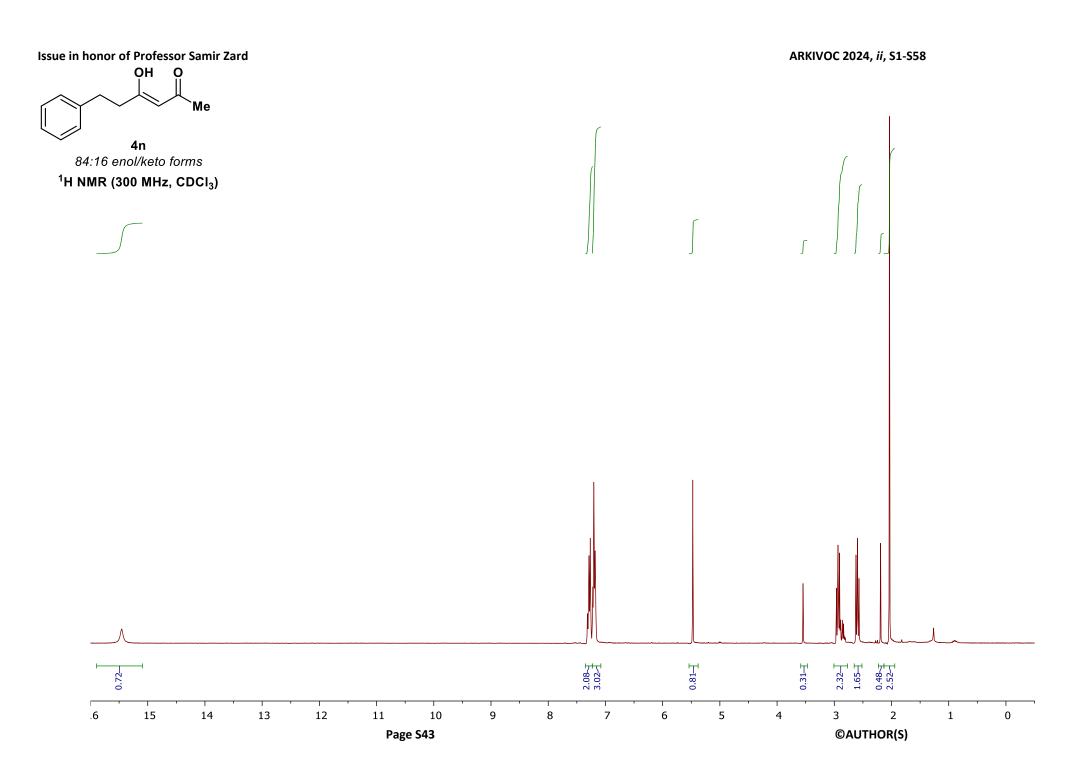


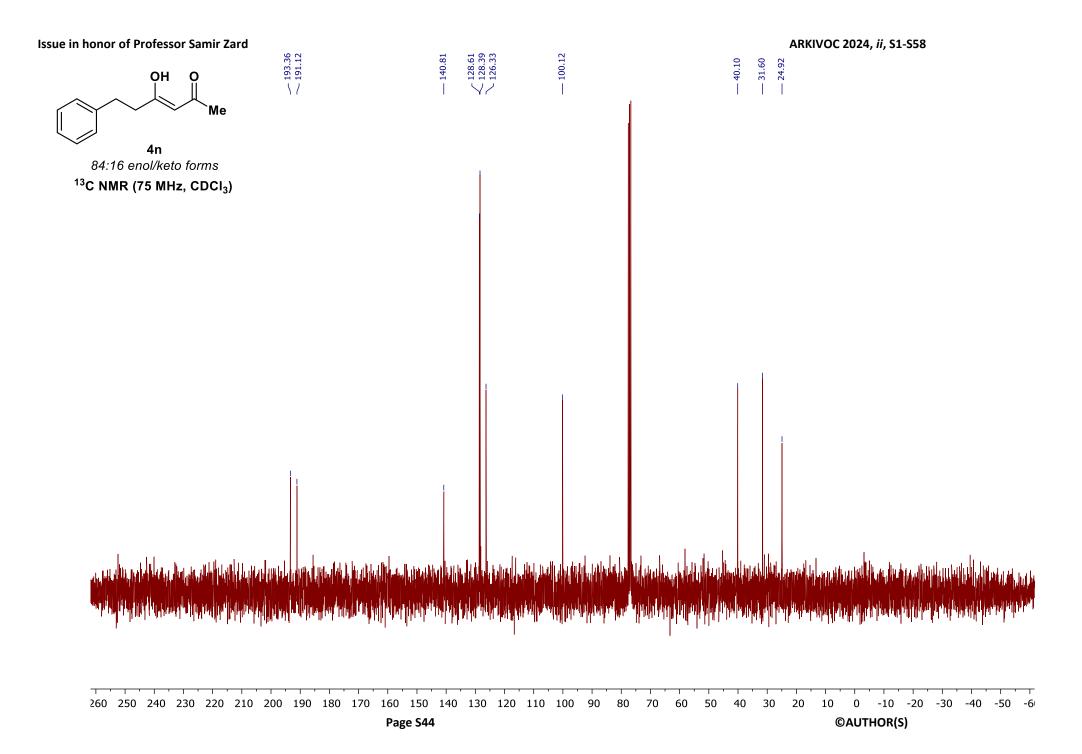






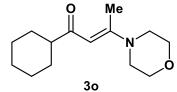






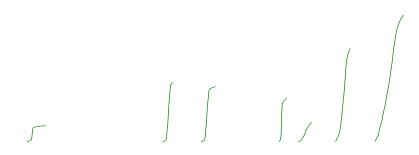


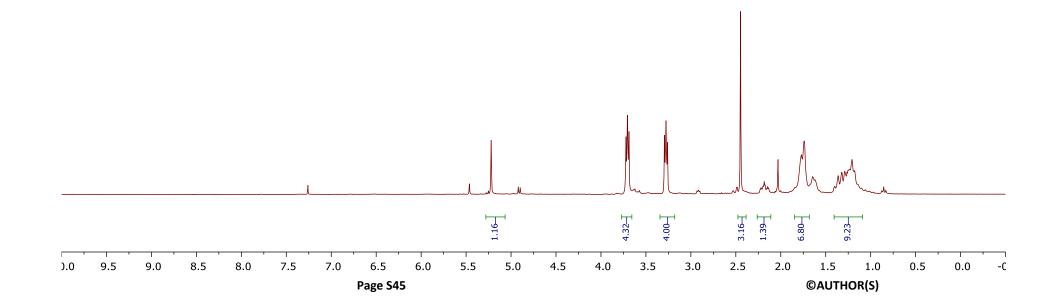
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(ca. 85% purity)

¹H NMR (300 MHz, CDCI₃)

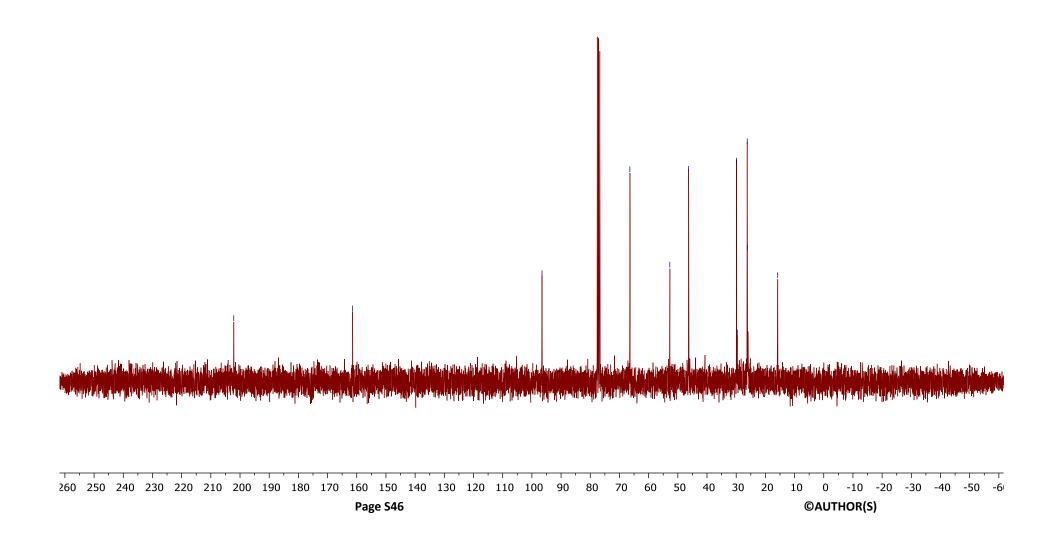


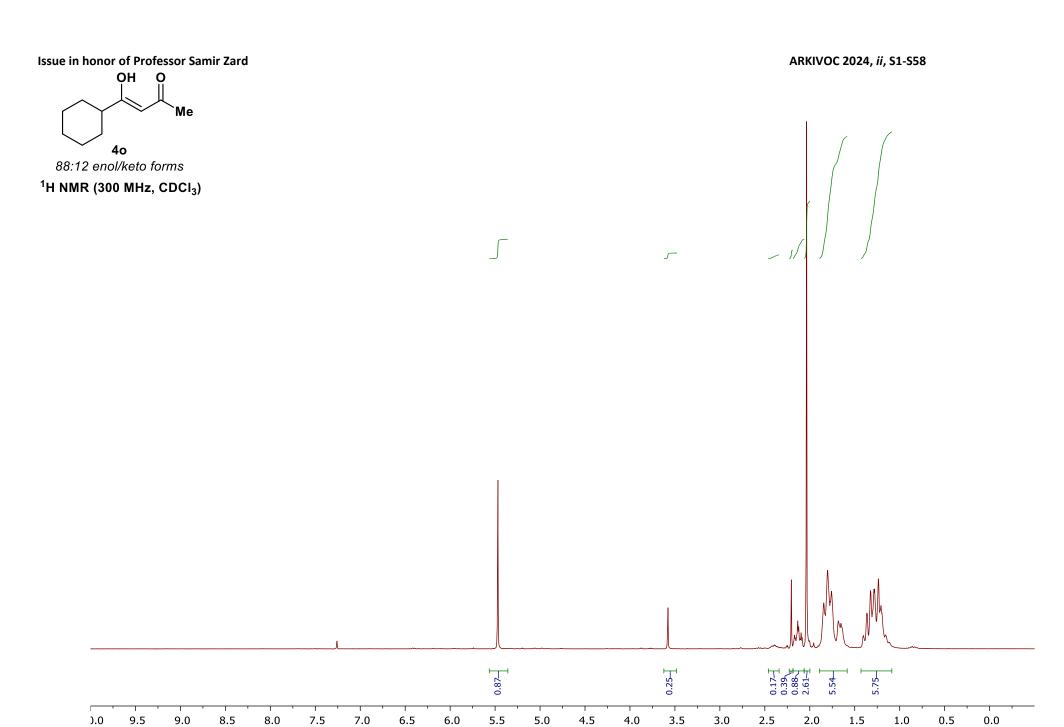




(ca. 85% purity)

13C NMR (75 MHz, CDCl₃)





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