

From simple phenols to potent chain-breaking antioxidants by transposition of benzo[1,4]oxathiines to benzo[*b*]thiophenes

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Dedicated to Professor Lorenzo Testaferri in the occasion of his 75th birthday

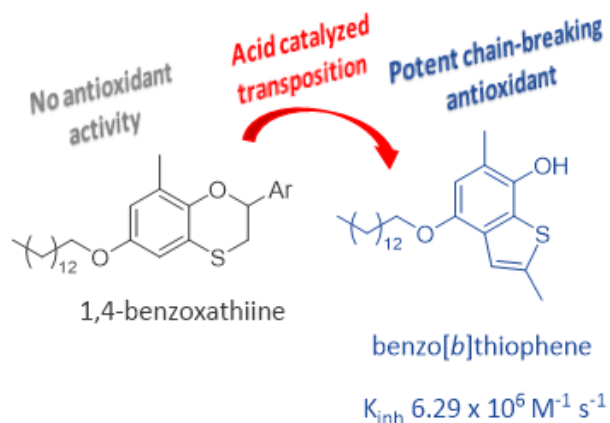
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Abstract

Simple phenols, including food stock recycled derivatives, were used for the synthesis of 1,4-benzoxathiine intermediates, with limited or no antioxidant activity. Depending upon their substitution pattern, these compounds, through an acid promoted transposition, can be converted into *o*-hydroxy-dihydrobenzo[*b*]thiophenes or *o*-hydroxy-benzo[*b*]thiophenes as a new class of potent chain-breaking antioxidants, showing, in the reaction with alkyl peroxy radicals, kinetic rate constants (k_{inh}) comparable to that of α -tocopherol, the more potent natural lipophilic phenolic antioxidant.



Keywords: Antioxidant, transposition, sulfur heterocycle, benzo[*b*]thiophenes, phenols

Introduction

Reactive Oxygen Species (ROS) and other free radicals are natural by-products of the human metabolism playing an important role in cell signalling, homeostasis and biochemical transformations.¹⁻³ On the other hand, they can also be formed through a variety of exogenous events such as exposure to UV light and ionizing radiation, smoking, air pollution and inflammation. Indeed, an anomalous high concentration of ROS in tissues is strictly related to oxidative stress, in turn cause and effect of several diseases and aging itself.⁴⁻¹¹

Our endogenous antioxidant defense systems have the function to control ROS level in order to conserve their biological benefits avoiding risky high concentrations and preventing oxidative stress and related diseases such as neurodegeneration, cardiovascular injury, mutation of healthy cells into cancerous ones, etcetera.⁴⁻¹¹

On the other hand, many substances, taken with diet, are able to control oxidative processes acting as exogenous antioxidants. In particular, natural and synthetic phenols have the ability to prevent auto-oxidation acting as chain breaking antioxidant a process that is crucial for the protection of cell membranes and LDL.

Phenols (ArOH) are known to reduce the rates of oxidation of organic materials by transferring an H atom (H[•]) from their OH groups to the more common chain-carrying species *i.e.* peroxy radicals (ROO[•]).¹²⁻¹⁴ Polyphenols as flavonoids¹⁵⁻¹⁷ (vitamin P) and tocopherols^{18,19} (vitamin E) are the more important examples of such natural antioxidants.

Recently, we focused our attention on some sulfur containing phenols, namely, o-hydroxy-dihydrobenzo[*b*]thiophenes²⁰ and benzo[*b*]thiophenes²¹ as efficient multi-defense antioxidants.

In particular, we have settled out new acid-catalysed transpositions that allow, in few steps with reasonably good overall yields, to transform easily accessible benzo[1,4]oxathiines²⁰⁻²⁸ into 7-hydroxy-dihydrobenzo[*b*]thiophenes or 7-hydroxy-benzo[*b*]thiophenes (Figure 1). These benzofused phenolic systems showed antioxidant activities similar to that of α -tocopherol (α -TOH), the main component of vitamin E and the more potent lipophilic antioxidant known in nature, showing in the reaction with ROO[•] kinetic constants (k_{inh}) as high as $3.2 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$.

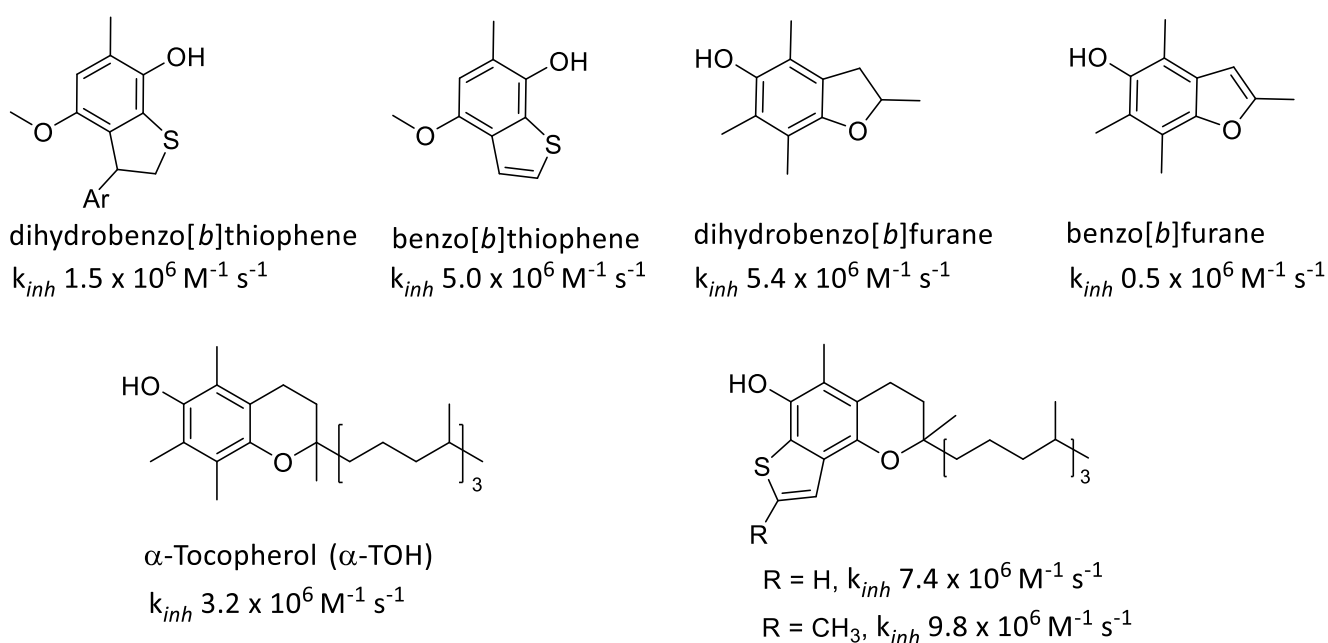
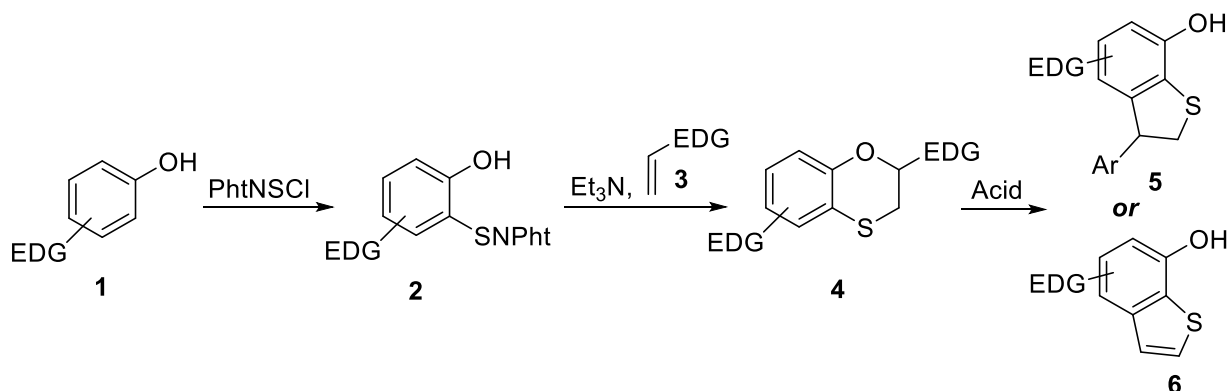


Figure 1. Structure and kinetic constants (k_{inh}) of model phenolic compounds discussed in this study.

Moving from a 6-hydroxychromane (like in α -TOH) to a 5-hydroxydihydrobenzo[*b*]furane the k_{inh} slightly increases since the five-membered ring is almost coplanar with the aromatic ring, hence one of the lone pairs of the endocyclic O atom lays parallel to the aromatic π orbitals allowing an optimal stabilization of the phenoxyl radical (ArO^{\bullet}).²⁹ On the other hand 5-hydroxybenzo[*b*]furane, despite its complete planarity, showed a ten time reduced hydrogen transfer ability due to the aromatization that, involving the lone pair of endocyclic O atom, reduces its electron donating ability and ArO^{\bullet} stability.²⁹ We demonstrated that, considering the corresponding sulfur heterocycles, moving from 7-hydroxydihydrobenzo[*b*]thiophenes to 7-hydroxybenzo[*b*]thiophenes there is an overall increasing (up to three times) of the k_{inh} .²¹ The (weak) intramolecular H-bond stabilization of the $ArOH$, and the electron donating effect stabilization of sulfur moieties on ArO^{\bullet} do not allow to rationalize this unexpected result of k_{inh} increasing with aromatization. We have suggested that the extra stabilization of the phenoxyl radical in the benzothiophene compound is due to the contribution of a chalcogen-bond effect,³⁰⁻³⁵ namely the interaction between the high electron density around the oxygen centered radical of ArO^{\bullet} and the sulfur σ -hole of the thiophene that ensures an overall increasing of the H^{\bullet} transfer attitude.

We applied this observation preparing a couple of super vitamin-E like phenolic antioxidants bearing a benzo[*b*]thiophene residue (Figure 1) perfectly suitable for studying the role of chain breaking activity in biological environments.²¹

In this paper, we report the scope and limitation of the transformation of different phenols (**1**), with limited antioxidant activity, through benzo[*b*]oxathiines **4** (typically without any pendant OH, hence without any antioxidant activity), into *o*-hydroxy benzothiophene systems **5**, **6** showing a remarkable chain breaking antioxidant ability (Scheme 1).



Scheme 1. The synthetic transformations considered in this study.

Results and Discussion

Synthesis of *N*-thiophthalimide derivatives **2**

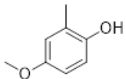
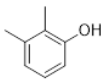
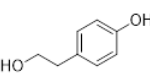
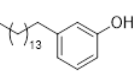
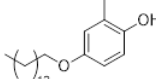
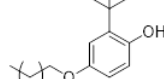
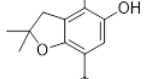
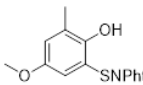
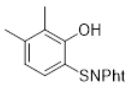
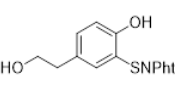
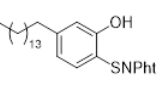
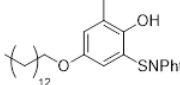
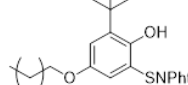
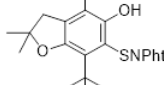
Our research group has developed the synthesis of benzo[1,4]oxathiine derivatives that exploits the reactivity of *ortho*-hydroxy-*N*-thiophthalimides **2** in turn obtained by reaction of phthalimidesulfonyl chloride (PhtNSCl, Pht = Phthaloyl) with phenols containing at least one additional electron donor group (EDG).²²⁻²⁸ *o*-Hydroxy-*N*-thiophthalimides **2** are the result of an electrophilic aromatic substitution (S_eAr) that occurs without any catalyst under mild conditions. The sulfenylation with PhtNSCl proved to be highly *ortho*-hydroxy-

regioselective and, typically, affords a mono-sulfenylated derivative. In fact, after the first substitution, the thiophthalimide group deactivates the aromatic ring toward further S_EAr . Phenols **1a-g** used as substrates and *N*-thiophthalimides **2a-g** prepared are depicted in table 1. Phenol **1a** was synthesized through a Wolff-Kishner reduction starting from 2-hydroxy-5-methoxybenzaldehyde as previously reported.²⁰ Derivative **1b** is commercially available and tyrosol **1c** is a natural phenolic antioxidant present in a variety of natural sources. Saturated cardanol **1d** is obtained from anacardic acid, the main component of cashew nutshell liquid (CNSL), a by-product of cashew nut processing. Derivatives **1e** and **1f** were prepared by alkylation of commercially available 2-methyl- and 2-*tert*-butyl-hydroquinone (see experimental). The bromide used for alkylation was chosen to obtain derivatives **1e** and **1f** bearing a long saturated tail like in Tocopherols.

The dihydrobenzofuranol **1g** was prepared in one-step starting from 2-*t*-butyl-5-methyl hydroquinone, 2-methyl-2-propen-1-ol and *p*-toluenesulfonic acid (PTSA). The steric hindrance of *tert*-butyl group allowed the complete control of the regiochemistry giving **1g** as the unique phenol isolated in 74% (see experimental).

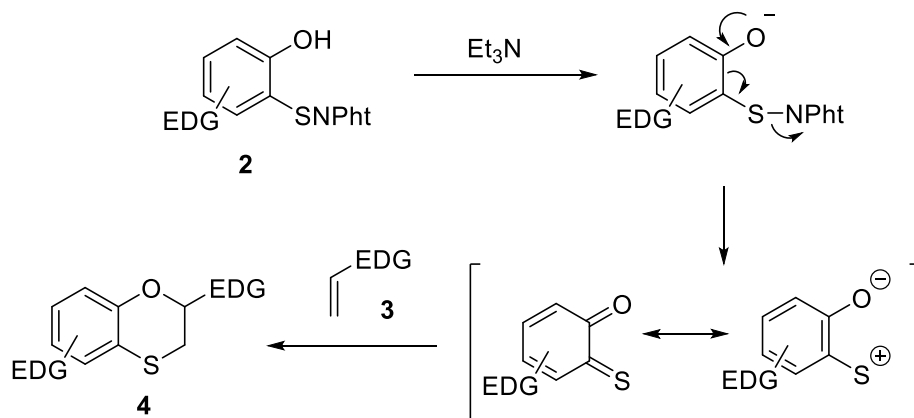
The synthesis of *N*-thiophthalimide derivatives **2** (Table 1), was carried out following our standard procedure. To a 0.1M solution of each phenol in chloroform a 0.1M solution of phthalimidesulfonyl chloride in chloroform was added dropwise keeping the reaction mixture at 0°C. Then the mixture was kept under stirring at room temperature until complete consumption of the starting phenol. The expected *o*-hydroxy-*N*-thiophthalimide derivatives **2a-g** were obtained after a trivial aqueous work-up and solvent evaporation and typically used without further purification (Table 1).

Table 1. Synthesis of *N*-thiophthalimide derivatives **2**

Entry	1	2	3	4	5	6	7
Phenol	 1a	 1b	 1c	 1d	 1e	 1f	 1g
Product (yield)	 2a	 2b (63%)	 2c (16%)	 2d (≥99%)	 2e (≥99%)	 2f (≥99%)	 2g (≥99%)

Synthesis of benzo[1,4]oxathiine cycloadducts **4**

In the last three decades²⁰⁻²⁸ we demonstrated that the reaction of *o*-hydroxy-*N*-thiophthalimide derivatives **2a-g** with a base, such as triethylamine, allows the formation of the corresponding *o*-thioquinones as the result of ArOH deprotonation and 1-4 elimination at sulfur of the phthalimide anion from the phenate ion (scheme 2). *o*-Thioquinones are transient intermediates, very efficient as electron-poor dienes in inverse electron demand hetero Diels-Alder reactions with a plethora of electron-rich alkenes (**3**, Figure 2) acting as dienophiles to afford of benzo[1,4]oxathiine cycloadducts **4** under mild conditions (Scheme 2).



Scheme 2. Access to benzo[1,4]oxathiines **4** from *o*-hydroxy-*N*-thiophthalimides **2** via *o*-thioquinones.

Noteworthy, the high polarization of the hetero-diene^{36,37} ensures, in all cases, the formation of a single regioisomeric cycloadduct in which the oxygen atom of the thioquinone is bounded to the more electron-poor carbon atom of the dienophile (Scheme 2). Electron-rich dienophiles **3** used for this study are depicted in Figure 2. All dienophiles used are commercially available, apart from 4-methoxy- α -methylstyrene (**3c**), which was prepared through a Wittig olefination from 4-methoxy-acetophenone and methyltriphenylphosphonium bromide as previously reported.²⁰

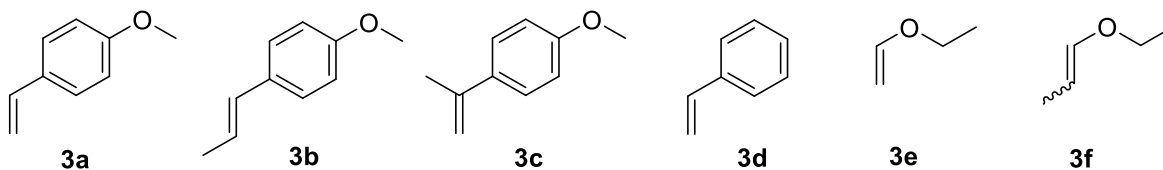


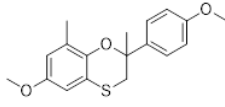
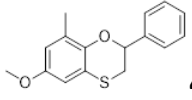
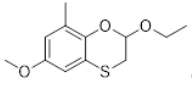
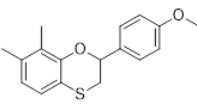
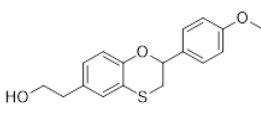
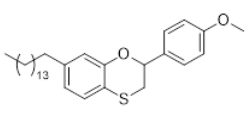
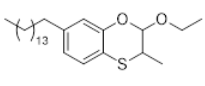
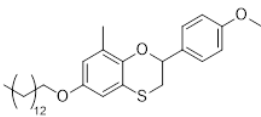
Figure 2. Electron-rich alkenes used as dienophiles in this study.

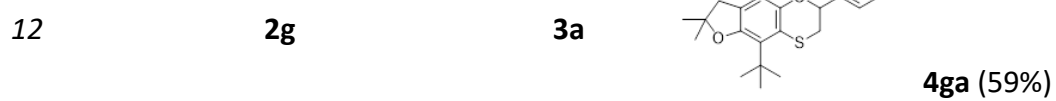
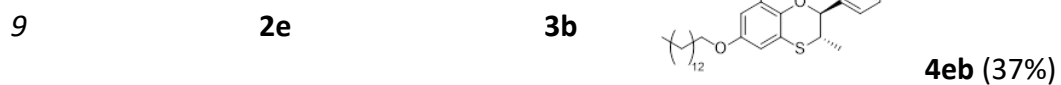
The cycloaddition reactions were carried out as following: to a (roughly) 0.1 M solution of the *o*-hydroxyl-*N*-thiophthalimide derivative **2** in chloroform, an excess (2-5 equiv) of the dienophile **3** was added followed by freshly distilled triethylamine (1 equiv). The mixture was kept under magnetic stirring at 60 °C until complete consumption of starting *o*-hydroxy-*N*-thiophthalimide. The crude obtained after an aqueous work-up was purified by column chromatography on silica gel to afford the desired benzo[1,4]oxathiine **4** as depicted in Table 2.

The use of an excess of dienophile was decided to speed up the cycloaddition allowing a relative fast consumption of the *o*-thioquinone. The reaction with styrene **3d** was carried out in dichloroethane at 80 °C since the lack of EDG decreases its ability as dienophile.

The cycloaddition reactions were indeed completely regiocontrolled allowing the isolation of a single regioisomeric cycloadduct where the oxygen atom of the thioquinones binds to the more electron-poor carbon atom of the dienophile. Additionally, the cycloaddition occurs with retention of the alkene geometry, as expected for a concerted [4+2] process. Thus, reacting alkene **3f**, commercially available as a 72:28 *trans*:*cis* mixture, allowed the isolation of cycloadduct **4df** as 72:28 *trans*:*cis* mixture of diastereoisomers (Table 2, entry 7), while reactions carried out with *trans*-anethole **3b** gave *trans*-benzo[*b*][1,4]oxathiines **4eb** and **4fb** as single diastereoisomers (Table 2, entries 9, 11).

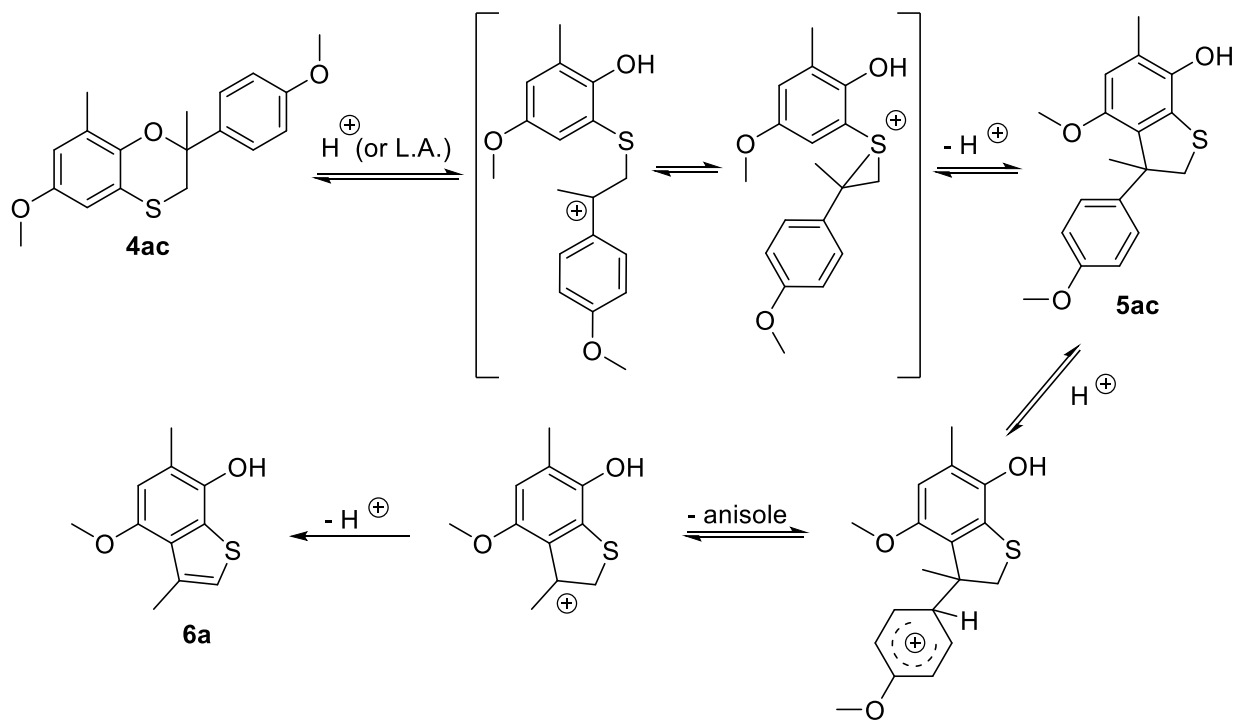
Table 2. Synthesis of benzo[1,4]oxathiine cycloadducts 4

<i>Entry</i>	<i>o</i> -hydroxy- <i>N</i> -thiophthalimide	Dienophile	<i>Product (Yield)</i>
1	2a	3c	 4ac¹
2	2a	3d	 4ad (30%)
3	2a	3e	 4ae (64%)
4	2b	3a	 4ba (71%)
5	2c	3a	 4ca (57%)
6	2d	3a	 4da (62%)
7	2d	3f	 4df (<i>cis-trans</i> mixture)
8	2e	3a	 4ea (36%)



Acid mediated transposition of benzo[1,4]oxathiines **4** to dihydrobenzo[*b*]thiophenes **5** and benzo[*b*]thiophenes **6**

Having in hand a variety of different substituted benzoxathiine cycloadducts **4** (Table 2) we had the opportunity to study and rationalize the limitation and scope of their transformation into 7-hydroxy-dihydrobenzo[*b*]thiophenes **5**,²⁰ and/or 7-hydroxybenzo[*b*]thiophenes **6**,²¹ using the better condition for both processes, *i.e.* chloroform as solvent and BF₃·OEt₂ or triflic acid (TfOH) as acid promoters. The mechanism we propose is depicted in Scheme 3 using derivative **4ac** as model substrate.



Scheme 3. Acid promoted transformation of benzoxathiines **4** into dihydrobenzo[*b*]thiophene **5** and benzo[*b*]thiophenes **6**.

Thus, benzoxathiine **4ac**, under acid conditions gives ring opening of the benzo-fused heterocyclic ring with formation of a benzylic carbocation which, through an internal electrophilic alkylation (*i*-S_EAr) gives rise to ring closure and formation of dihydrobenzo[*b*]thiophene **5ac**. This first part of the process is directly related with the well-known chromane-indene acid catalysed transposition despite occurring under milder conditions.³⁸⁻⁴⁴ In fact, using benzoxathiine as starting materials, the intermediate carbocation is stabilized by the anchimeric assistance of the sulfur atom and formation of a thiiranium ion.²⁰ Once derivative **5ac** is formed and under harsh acid conditions, we postulated the possibility of a retro-alkylation (*r*-S_EAr), *via* protonation of aromatic ring and elimination, in the case of **5ac**, of anisole. The new cyclic carbocation formed easily deprotonate to give the aromatic benzo[*b*]thiophene **6a**²¹ (Scheme 3).

Keeping this mechanism in mind, we studied the behaviour of derivatives **4** as reported in Table 3. Benzoxathiine **4ac** reacts smoothly with BF₃·Et₂O (3.0 equiv) giving **5ac** in 66% yield. Indeed, the benzylic carbocation formed from **4ac** is stabilized by the 4'-methoxy group on the aryl ring and by the methyl group on the benzylic carbon. Actually, the formation of high stabilized tertiary carbocation plays a crucial role in both processes since **4ac** is the only derivative that allowed the isolation of the corresponding benzo[*b*]thiophene **6a** (16%) just under BF₃·Et₂O catalysis (Table 3, entry 1). As a matter of fact, reacting **4ac** with 1 equiv. of TfOH,

6a becomes the only product observed in the crude mixture, and isolated in 46% yield (Table 3, entry 2), indicating an easy and fast transformation of **5ac** into **6a** facilitated by the stability of the cyclic tertiary carbocation formed after anisole elimination (Scheme 3). The critical role of carbocation stability in both processes (*i*-S_EAr and *r*-S_EAr) depicted in Scheme 3 is boosted by the result obtained with derivative **4ab**. In fact, in the absence of a 4'-methoxy group and alkyl substituents on benzylic carbon, the reaction does not take place with 3 equiv of BF₃·Et₂O at 60 °C being the starting material stable under these conditions for more than 12h. Repeating the reaction under drastic acid conditions, 5 equiv of TfOH at 60 °C, for 6.5h, we were able to isolate dihydrobenzo[*b*]thiophene **5ad** albeit in low yield, 25%, while analysis of the crude pointed out a large amount of unreacted starting material without any trace of the corresponding benzo[*b*]thiophene.

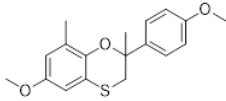
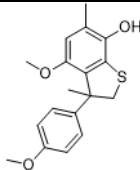
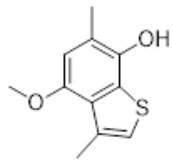
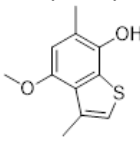
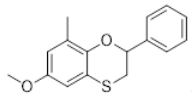
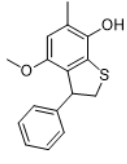
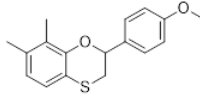
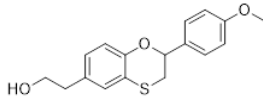
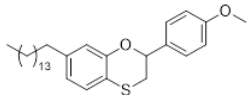
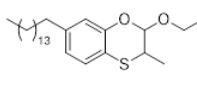
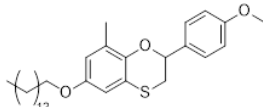
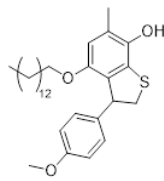
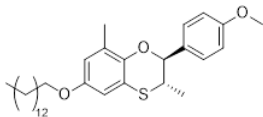
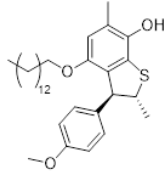
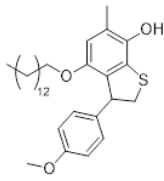
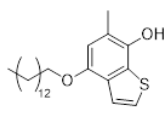
Together with the role of carbocation stability, the substitution pattern on the oxathiine benzofused aryl ring demonstrated to be crucial as well. In fact, when derivatives **4ba**, **4ca**, **4da** and **4df**, able to generate stabilized carbocations, were reacted with either BF₃·Et₂O or TfOH at 60 °C any benzothiophene was observed yet, typically, an extensive decomposition of the starting materials was experienced (Table 3, entries 4-7). In all these compounds the position in the aromatic ring *ortho* to the sulfur atom is less nucleophilic due to the lack of the alkoxy group in *ortho*' position, a structural feature that demonstrated to be mandatory for the success of the procedure. This prevented the possibility to transform stock recycled phenols, like tirosol and cardanol, into potent chain breaking antioxidants. In perfect agreement with previous observations, benzoxathiines **4ea** and **4eb** easily reacted with BF₃·Et₂O to give dihydrothiophenes **5ea** and **5eb** in 53% and 63% yield respectively (Table 3, entries 8 and 9). For these derivatives the one-pot formation of benzothiophenes **6** from **4** was poor effective. However, benzo[*b*]thiophenes **6b** and **6c** were obtained in 48% and 50% yield respectively carrying out the process in two steps by reacting the corresponding dihydrobenzo[*b*]thiophenes **5ea** and **5eb** with 0.5 equiv of TfOH (Table 3, entries 10 and 11).

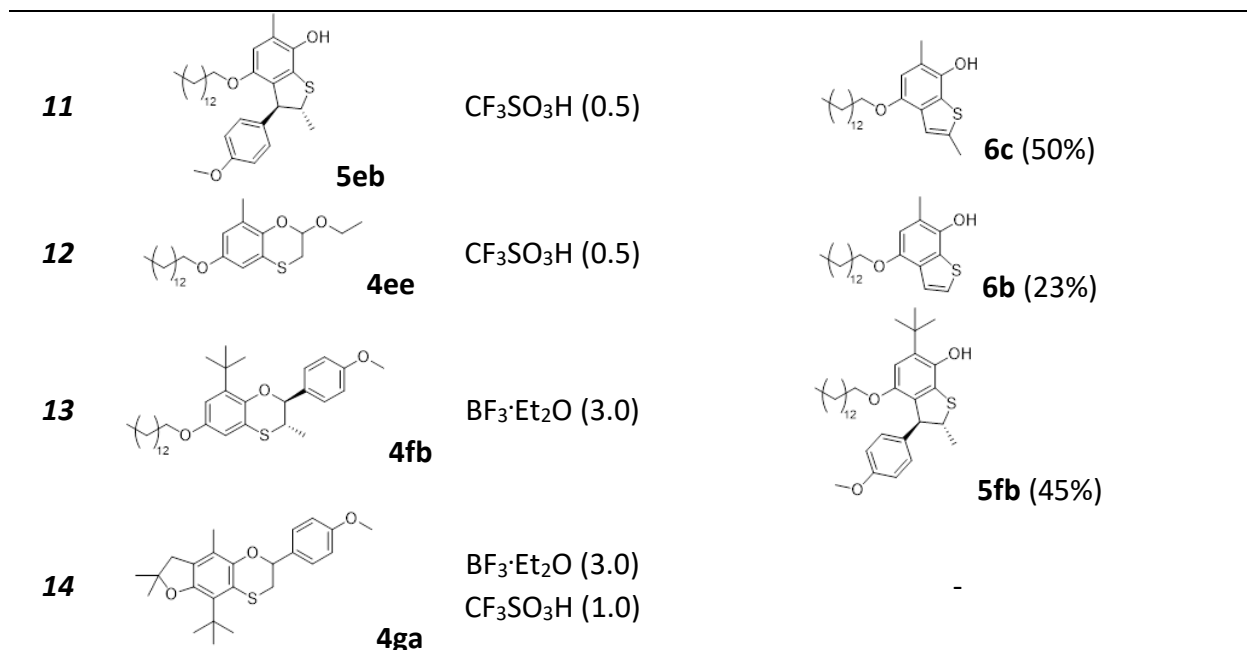
As a matter of fact, benzo[*b*]thiophene **6b** was also prepared reacting benzoxathiine **4ee** with 0.5 equiv. of TfOH. In this case, the transposition was very fast and, in just a hour, thiophene **6b** was the unique product of the reaction, while no trace of residual starting material or intermediate dihydrobenzo[*b*]thiophene were detected by TLC and ¹H NMR analysis of the crude mixture (Table 3, entry 12). Derivative **4ee** was obtained using ethyl vinyl ether as dienophile (see Table 2 entry 10). Taking in consideration the mechanism depicted in Scheme 3 we can suggest that the first carbocation, obtained by oxathiine ring opening, is stabilized by the adjacent ethoxy group, while the aromatization process of the corresponding 3-ethoxy-dihydrobenzo[*b*]thiophene foresees and easy elimination of ethanol (see scheme 3). In other words, once the structural features required for both processes are maintained, the transformation occurs independently upon the presence of a *p*-methoxyaryl group on C2 of benzoxathiine.

As a further demonstration that a fine tuning of the substitution pattern is mandatory to achieve either dihydro- and benzo[*b*]thiophenes, compound **4fb** was simply transformed into dihydrobenzo[*b*]thiophene **5fb** in 45% yield using BF₃·Et₂O (Table 3, entry 13), however, all the attempts to push the reaction to the corresponding benzo[*b*]thiophenes using TfOH as promoter, from **4fb** or **5fb** as starting materials, failed while we observed extensive de-terbutylation and decomposition.

Eventually, we tested derivative **4ga**, a benzo[1,4]oxathiine with a dihydrobenzofuranic moiety and no free nucleophilic positions on the benzofused aromatic ring (see Table 2 entry 14). Indeed, we used similar compounds for the preparation of the benzo[*b*]thiophenes containing the α-TOH skeleton²¹ (Figure 1) exploiting a TfOH promoted de-terbutylation since to liberate the required nucleophilic position *ortho* to the sulfur atom. However, in this case the dimethyl-dihydrobenzofuranic ring proved to be poor stable under acid condition and de-terbutylation occurred together with rings opening preventing the thiophene ring formation (Table 3, entry 14).

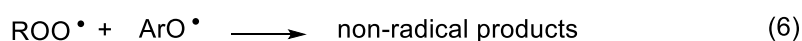
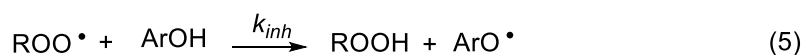
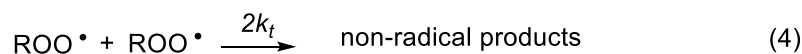
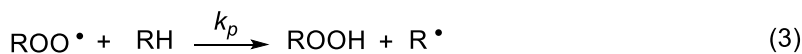
Table 3. Access to dihydrobenzo[*b*]thiophenes **5** and benzo[*b*]thiophenes **6**

Entry	Substrate	Acid (equiv)	Product (Yield)	
1	 4ac	BF ₃ ·Et ₂ O (3.0)	 5ac (66%)	 6a (16%)
2	4ac	CF ₃ SO ₃ H (1.0)	 6a (46%)	
3	 4ad	BF ₃ ·Et ₂ O (3.0) CF ₃ SO ₃ H (5.0)	<i>No products</i>  5ad (25%)	
4	 4ba	BF ₃ ·Et ₂ O (3.0)	-	
5	 4ca	BF ₃ ·Et ₂ O (3.0) CF ₃ SO ₃ H (1.0)	-	
6	 4da	BF ₃ ·Et ₂ O (3.0) CF ₃ SO ₃ H (1.0)	-	
7	 4df	BF ₃ ·Et ₂ O (3.0) CF ₃ SO ₃ H (1.0)	-	
8	 4ea	BF ₃ ·Et ₂ O (3.0)	 5ea (53%)	
9	 4eb	BF ₃ ·Et ₂ O (3.0)	 5eb (63%)	
10	 5ea	CF ₃ SO ₃ H (0.5)	 6b (48%)	



Chain-breaking antioxidant activity of dihydrobenzo[*b*]thiophenes **5** and benzo[*b*]thiophenes **6**

The antioxidant activity of the dihydrobenzo[*b*]thiophenes **5** and benzo[*b*]thiophenes **6** was assessed by studying the inhibition of the thermally initiated autoxidation of styrene (R=styrene, 50% v/v in chlorobenzene), used as reference hydrocarbon, under controlled conditions. Autoxidation of styrene is a radical chain reaction (see Equations 1-6) carried on mainly by alkylperoxyl radicals (ROO[•]) which are representative of the reactive oxygen species responsible for the oxidation of natural lipids and man-made materials under air. The initiator is represented by azobis(isobutyronitrile) (AIBN), whose decomposition at 30°C originates a constant flux of free radicals (initiation rate, *R_i*). The reaction was followed by measuring the oxygen consumption by an automatic oxygen uptake recording apparatus, build in our laboratory, based on a differential pressure transducer.⁴⁵⁻⁴⁸



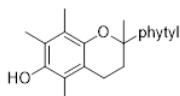
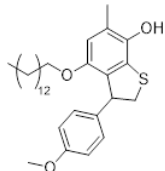
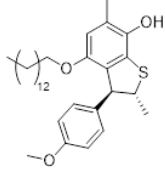
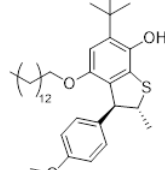
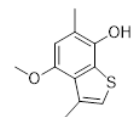
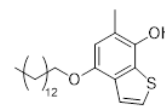
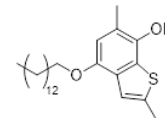
$$R_i = n[ArOH]/\tau \quad (7)$$

$$\Delta O_2 = (k_p k_{inh})[\text{styrene}] \ln(1-t/\tau) \quad (8)$$

In the absence of antioxidants, the O₂ consumption is fast and linear, whereas in the presence of an antioxidant, the O₂ consumption is retarded or completely inhibited for a period that depends on its

concentration. The number of radicals trapped by each antioxidant molecule (n) is related to the length of the inhibited period τ through Equation (7), in which the initiation rate R_i can be determined by using a reference antioxidant. The rate constant of the reaction of the antioxidant with ROO^\cdot radicals (k_{inh} , see Equation 5) was calculated by using Equation (8): A plot of $\Delta[\text{O}_2]t$ vs $\ln(1-t/\tau)$ gives a straight line of slope $k_p [\text{styrene}]/k_{inh}$ from which k_{inh} is obtained by using the known k_p value at 30°C of styrene, that is, $41 \text{ M}^{-1} \text{ s}^{-1}$.^{48,49} The results obtained by using this method are reported in Table 4 and compared with that of α -TOH.

Table 4. k_{inh} and n values of derivatives **5** and **6** compared with those of α -tocopherol

Entry	Compound	$k_{inh} \text{ M}^{-1}\text{s}^{-1}$	n
1	 α-Tocopherol	3.2e^6	2
2	 5ea	$(1.80\pm 0.4)\text{e}^6$	1.8 ± 0.1
3	 5eb	$(1.63\pm 0.06)\text{e}^6$	1.86 ± 0.06
4	 5fb	$(1.22\pm 0.02)\text{e}^6$	1.92 ± 0.14
5	 6a	$(3.90\pm 1.3)\text{e}^6$	1.83 ± 0.07
6	 6b	$(5.24\pm 0.36)\text{e}^6$	1.86 ± 0.11
7	 6c	$(6.29\pm 0.71)\text{e}^6$	1.78 ± 0.24

From Table 4 we observe that the products tested have a rate inhibition constant of the same order of magnitude of α -TOH, indicating they are all potent chain breaking antioxidants. The number of peroxy radicals

trapped by each antioxidant (n) is approximately 2 demonstrating that the mechanism of neutralization of radicals is also analogous to that of α -TOH.

In more details it is confirmed also that moving from dihydrobenzo[*b*]thiophenes **5** (Table 4 entries 2-4) to dihydrobenzo[*b*]thiophenes **6** (Table 4 entries 5-7) there is a solid increasing of k_{inh} that allowed to consider a benzo[*b*]thiophene ring as the more efficient residue to insert in *ortho* position to OH group for increasing the chain breaking antioxidant activity of a phenolic compound.

Conclusions

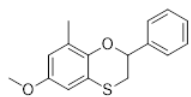
In this paper we reported a study dedicated to elucidating the scope of the acid promoted transposition of benzoxathiines into dihydrobenzo[*b*]thiophenes and benzo[*b*]thiophenes, the latter obtainable directly from oxathiines or from the dihydro- derivatives. The structural features on both C2 and benzofused aromatic ring of benzoxathiines required to allow the first and the second transposition have been clarified and exploited for the preparation of potent phenolic chain breaking antioxidants. The peculiar ability of the benzothiophene sulfur atom in promoting the H \bullet atom transfer process from the *ortho* OH group to a peroxy radical (ROO \bullet) has been confirmed. In fact, derivatives **5** and **6** showed kinetic constants (k_{inh}) in the reaction with ROO \bullet similar or superior to that of α -TOH, the more potent natural lipophilic antioxidant known.

We are now applying these results for the preparation of benzoxathiine derivatives to be used as a "reservoir" of potent antioxidants being unreactive towards oxidant radical species under neutral or basic conditions, but very reactive under acid conditions. The aim is to exploit these derivatives in peculiar physiologic conditions such as those cancer cells experiencing a hypoxic and low pH environment.

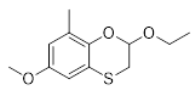
Experimental Section

General. ^1H and ^{13}C NMR spectra were recorded with Varian Mercury Plus 400, using CDCl_3 as solvent. Residual CHCl_3 at 7.26 ppm and central line of CDCl_3 at 77.00 ppm were used as the reference of ^1H and ^{13}C NMR spectra respectively. FT-IR spectra were recorded with Spectrum Two FT-IR Spectrometer. GC-MS spectra were recorded with a QMD 100 Carlo Erba. ESI-MS spectra were recorded with a JEOL MStation JMS700. Melting points were measured with Stuart SMP50 Automatic Melting Point Apparatus. All the reactions were monitored by TLC on commercially available precoated plates (silica gel 60 F 254) and the products were visualized with acidic vanillin solution. Silica gel 60 (230–400 mesh) was used for column chromatography. Dry solvents were obtained by The PureSolv Micro Solvent Purification System. Chloroform was washed with water several times and stored over calcium chloride. Pyridine and TEA were freshly distilled over KOH. Phthalimide sulfonyl chloride was prepared from the corresponding commercially available disulfide (purchased from Chemper snc) as reported elsewhere. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated.

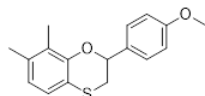
General cycloaddition procedure for the synthesis of derivatives 4. To a solution of derivatives **2** in CHCl_3 (0.1 M), was added the dienophile **3** (3 eq.) and triethylamine (1 eq.). The reaction mixture was left stirring at 60 °C until TLC analysis indicated complete consumption of the starting material, then was diluted with CH_2Cl_2 , washed once with a saturated solution of NH_4Cl . The organic layer was dried over Na_2SO_4 , filtered and evaporated *in vacuo*.



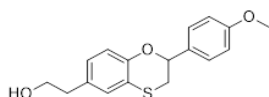
4ad. From **2a** (571 mg, 1.81 mmol). The crude was purified by silica gel column chromatography, using a mixture of petroleum ether and CH_2Cl_2 2:1 as eluent. The purified product was obtained as pale yellow solid (150 mg, 30% yield) mp 93-95 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.60 – 7.29 (m, 5H), 6.52 (s, 2H), 5.15 (bd, J 9.4 Hz, 1H), 3.75 (s, 3H), 3.25 (dd, J 12.9, 9.4 Hz, 1H), 3.11 (dd, J 12.9, 1.6 Hz, 1H), 2.22 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 153.3, 144.6, 140.7, 128.9, 128.7, 128.2, 125.8, 117.1, 114.0, 108.4, 76.2, 55.6, 32.3, 16.5. Elemental Analysis Calcd for $\text{C}_{16}\text{H}_{16}\text{O}_2\text{S}$: C 70.56, H 5.92; found C 70.64, H 5.97.



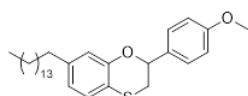
4ae. From **2a** (446 mg, 1.42 mmol). The crude was purified by silica gel column chromatography, using a mixture of petroleum ether and CH_2Cl_2 1:7 as eluent. The purified product was obtained as light brown solid (216 mg, yield 64%) mp 34-38°C. ^1H NMR (400 MHz, CDCl_3) δ 6.55 – 6.39 (m, 2H), 5.35 (dd, J 5.0, 2.3 Hz, 1H), 4.07 – 3.81 (m, 1H), 3.79 – 3.60 (s+m, 4H, $-\text{OCH}_3$), 3.14 (dd, J 12.7, 2.3 Hz, 1H), 3.00 (dd, J 12.7, 5.0 Hz, 1H), 2.19 (s, 3H), 1.25 (t, J 7.1 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 153.4, 141.7, 128.9, 117.9, 114.1, 108.4, 94.7, 64.1, 55.5, 29.6, 16.3, 15.0. Elemental Analysis Calcd for $\text{C}_{12}\text{H}_{16}\text{O}_3\text{S}$: C 59.98, H 6.71; found C 59.84, H 6.85.



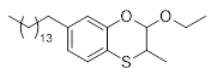
4ba. From **2a** (411 mg, 1.38 mmol). The crude was purified by silica gel column chromatography, using a mixture of petroleum ether and CH_2Cl_2 3:1 as eluent. The purified product was obtained as white solid (280 mg, yield 71%) mp 94-96 °C. ^1H NMR (400 MHz, CDCl_3) δ 7.38 – 7.35 (m, 2H), 6.98 – 6.93 (m, 2H), 6.87 (d, J 8.4 Hz, 1H), 6.71 (d, J 8.4 Hz, 1H), 5.15 (dd, J 9.6, 2.0 Hz, 1H), 3.84 (s, 3H), 3.21 (dd, J 13.2, 9.6 Hz, 1H), 3.06 (dd, J 13.2, 2.0 Hz, 1H), 2.23 (s, 3H), 2.14 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 159.5, 150.2, 134.2, 133.3, 127.3, 127.0, 126.2, 123.8, 122.7, 114.0, 76.4, 55.3, 32.0, 19.8, 11.8. Elemental Analysis Calcd for $\text{C}_{17}\text{H}_{18}\text{O}_2\text{S}$: C 71.30, H 6.34; found C 69.84, H 6.15.



4ca. From **2c** (144 mg, 0.46 mmol). The crude was purified by silica gel column chromatography, using a mixture of CH_2Cl_2 and AcOEt 20:1 as eluent. The purified product was obtained as colourless oil (79 mg, 57% yield). ^1H -NMR (400 MHz, CDCl_3) δ 2.77 (t, J 2.0 Hz, 2H), 3.04 (dd, J 6.3, 1.0 Hz, 1H), 3.27 (dd, J 6.3, 4.8 Hz, 1H), 3.78-3.86 (m, 5H), 5.1 (dd, J 4.8, 1.0 Hz, 1H), 6.85-6.89 (m, 2H), 6.92-6.96 (m, 2H), 6.97-6.99 (m, 1H), 7.31-7.34 (m, 2H). ^{13}C -NMR (100 MHz, CDCl_3) δ : 31.7, 38.2, 55.3, 63.6, 114.1, 117.2, 118.8, 126.2, 127.3, 127.4, 131.6, 132.4, 151.2, 159.7. Elemental Analysis Calcd for $\text{C}_{17}\text{H}_{18}\text{O}_3\text{S}$: C 67.52, H 6.00; found C 67.58, H 6.05.



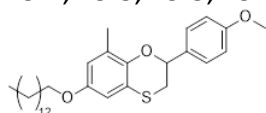
4da. From **2d** (103 mg, 0.38 mmol). The crude was purified by silica gel column chromatography, using a mixture of petroleum ether and CH_2Cl_2 6:1 as eluent. The purified product was obtained as colourless oil (110 mg, 62% yield). ^1H -NMR (400 MHz, CDCl_3) δ 0.85-0.93 (m, 3H), 1.22-1.35 (m, 26H), 2.45-2.57 (at, 2H), 3.03 (dd, J 13.1, 2.0 Hz, 1H), 3.27 (dd, J 13.1, 9.6 Hz, 1H), 5.14 (dd, J 9.6, 2.0 Hz, 1H), 6.65-6.80 (m, 2H), 6.90-6.98 (m, 2H), 7.02 (d, J 6.0 Hz, 1H), 7.30-7.40 (m, 2H). Elemental Analysis Calcd for $\text{C}_{30}\text{H}_{44}\text{O}_2\text{S}$: C 76.87, H 9.46; found C 76.90, H 9.51.



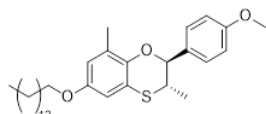
4df. From **2d** (589 mg, 1.22 mmol). The *cis* and *trans* products (*cis*-**4df** + *trans*-**4df** 60% yield) were isolated after silica-gel column chromatography using a mixture of CH₂Cl₂ and petroleum ether 1:3 as eluent.

cis-**4df** white solid, mp 41-44°C. ¹H-NMR (400 MHz, CDCl₃) δ: 0.85-0.93 (m, 3H), 1.17-1.34 (m, 29H), 1.37 (d, *J* 3.4 Hz, 3H), 2.45-2.55 (pt, 2H), 3.36 (qd, *J* 3.4, 1.0 Hz, 1H), 3.72 (dq, *J* 5.1, 3.5 Hz, 1H), 3.93 (dq, *J* 5.1, 3.5 Hz, 1H), 5.17 (d, *J* 1.0 Hz), 6.68-6.72 (m, 2H), 6.90-6.94 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ 14.1, 15.0, 15.4, 22.7, 29.2, 29.3, 29.5, 29.6 (2C), 29.7 (5C), 31.2, 31.9, 35.3, 36.2, 64.5, 97.1, 114.4, 118.2, 122.0, 126.6, 140.8, 148.5.

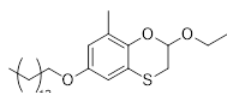
trans-**4df** white solid, mp 41-42°C. ¹H-NMR (400 MHz, CDCl₃) δ: 0.85-0.93 (m, 3H), 1.18-1.34 (m, 29H), 1.37 (d, *J* 3.4 Hz, 3H), 2.45-2.55 (pt, 2H), 3.36 (dq, *J* 2.1, 3.4 Hz, 1H), 3.72 (dq, *J* 5.0, 3.6 Hz, 1H), 3.93 (dq, *J* 5.0, 3.6 Hz, 1H), 5.17 (d, *J* 2.1 Hz), 6.68-6.72 (m, 2H), 6.92-6.96 (m, 1H). ¹³C-NMR (100 MHz, CDCl₃) δ: 14.1, 15.0, 15.4, 22.7, 29.2, 29.3, 29.5, 29.6 (2C), 29.7 (5C), 31.2, 31.9, 35.3, 36.2, 64.5, 97.1, 114.4, 118.2, 122.0, 126.6, 140.8, 148.5.



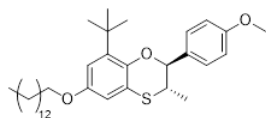
4ea. From **2e** (101 mg, 0.20 mmol). The crude product was purified by silica-gel column chromatography with a mixture of petroleum ether : CH₂Cl₂ = 3 : 1. The purified product was obtained as oil (37 mg, 36% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.35 (d, *J* 8.8 Hz, 2H), 6.94 (d, *J* 8.8 Hz, 2H), 6.49 (s, 2H), 5.08 (dd, *J* 9.6, 1.8 Hz, 1H), 3.87 (t, *J* 6.6 Hz, 2H), 3.83 (s, 3H), 3.22 (dd, *J* 12.9, 9.6 Hz, 1H), 3.05 (dd, *J* 12.9, 1.9 Hz, 1H), 2.17 (s, 3H), 1.77 – 1.70 (m, 2H), 1.44 – 1.39 (m, 2H), 1.35 – 1.26 (m, 20H), 0.88 (t, *J* 6.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 159.5, 152.8, 144.6, 133.0, 128.8, 127.0, 117.0, 114.5, 114.0, 109.2, 68.5, 55.3, 32.3, 31.9, 30.9, 29.7, 29.6, 29.4, 29.3 (4 signals for 9 non-equivalent CH₂ groups), 26.0, 22.7, 16.4, 14.1. Elemental Analysis Calcd for C₃₀H₄₄O₃S: C 74.33, H 9.15; found C 74.38, H 9.19.



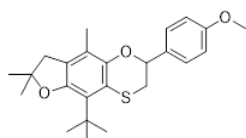
4eb. From **2e** (100 mg, 0.20 mmol). The crude product was purified by silica-gel flash chromatography with a mixture of petroleum ether : CH₂Cl₂ = 3 : 1 as eluent. The purified product was obtained as oil (37 mg 37% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, *J* 8.6 Hz, 2H), 6.94 (d, *J* 8.6 Hz, 2H), 6.48 (s, 2H), 4.61 (d, *J* 8.6 Hz, 1H), 3.87 (t, *J* 6.6 Hz, 2H), 3.84 (s, 3H), 3.46 – 3.38 (m, 1H), 2.12 (s, 3H), 1.78 – 1.71 (m, 2H), 1.45 – 1.40 (m, 2H), 1.33 – 1.28 (m, 20H), 1.09 + 1.07 (s, 3H), 0.91 – 0.88 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 159.6, 152.9, 144.7, 131.3, 128.4, 128.3, 118.9, 114.2, 113.9, 108.7, 82.5, 68.5, 55.2, 38.9, 31.9, 29.7, 29.6, 29.4 (3 signals for 9 non-equivalent CH₂ groups), 26.0, 22.7, 17.6, 16.5, 14.1. Elemental Analysis Calcd for C₃₁H₄₆O₃S: C 74.65, H 9.30; found C 74.70, H 9.34.



4ee. From **2e** (150 mg, 0.30 mmol). The crude product was purified by silica-gel column chromatography with a mixture of petroleum ether and CH₂Cl₂ 3 : 1 as eluent. The purified product was obtained as a pale yellow oil (37 mg, 53% yield). ¹H NMR (400 MHz, CDCl₃) δ 6.49 – 6.45 (m, 2H), 5.34 (dd, *J* 5.0, 2.2 Hz, 1H), 3.97 – 3.89 (m, 1H), 3.45 (t, *J* 6.6 Hz, 2H), 3.75 – 3.68 (m, 1H), 3.14 (dd, *J* 12.7, 2.2 Hz, 1H), 3.00 (dd, *J* 12.7, 5.0 Hz, 1H), 2.18 (s, 3H), 1.76 – 1.69 (m, 2H), 1.44 – 1.40 (m, 2H), 1.34 – 1.24 (m, 23H), 0.88 (t, *J* 6.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 153.0, 141.6, 128.8, 120.7, 117.8, 114.8, 109.2, 94.8, 68.4, 64.1, 31.9, 29.6, 29.4, 29.3 (3 signals for 9 non-equivalent CH₂ groups), 26.0, 22.7, 16.3, 15.0, 14.1. Elemental Analysis Calcd for C₂₅H₄₂O₃S: C 71.04, H 10.02; found C 71.07, H 10.04.

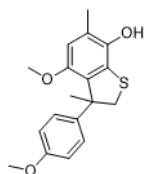


4fb. From **2f** (206 mg, 0.40 mmol). The crude product was purified by silica-gel column chromatography using a mixture of CH_2Cl_2 and petroleum ether 1:1 as eluent. The purified product was obtained as yellow oil (70 mg, 37%). ^1H NMR (400 MHz, CDCl_3) δ 7.28 (d, J 8.8 Hz, 2H), 6.93 (d, J 8.8 Hz, 2H), 6.60 (d, J 2.9 Hz, 1H), 6.50 (d, J 2.9 Hz, 1H), 4.40 (d, J 9.1 Hz, 1H), 3.87 (t, J 6.6 Hz, 2H), 3.84 (s, 3H), 3.56 (dd, J 9.1, 6.6 Hz, 1H), 1.77 – 1.73 (m, 2H), 1.46 – 1.42 (m, 2H), 1.36 – 1.27 (m, 22H), 1.24 (s, 9H), 0.88 (t, J 6.6 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 159.6, 153.2, 145.9, 141.0, 131.0, 128.3, 120.8, 113.9, 111.2, 107.7, 82.4, 68.4, 55.2, 39.8, 35.0, 31.9, 30.9, 29.7, 29.6, 29.4 (2 signals for 9 non-equivalent CH_2 groups), 26.1, 22.7, 17.9, 14.1. MS m/z (I_{rel} , %): 419 (31.29); 148 (100.00). Elemental Analysis Calcd for $\text{C}_{34}\text{H}_{52}\text{O}_3\text{S}$: C 75.51, H 9.69; found C 75.57, H 9.74.

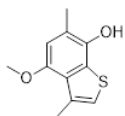


4ga. From **2g** (140 mg, 0.34 mmol). The crude was purified by silica-gel column chromatography, using a mixture of petroleum ether and CH_2Cl_2 2:1 as eluent. The purified product was obtained as an incolor oil (80 mg, 59% yield). ^1H NMR (400 MHz, CDCl_3) δ 7.36 (d, J 8.7 Hz, 2H), 6.93 (d, J 8.7 Hz, 2H), 5.19 (dd, J 9.2, 3.6 Hz, 1H), 3.83 (s, 3H), 3.08 (dd, J 12.9, 3.6 Hz, 1H), 2.99 (dd, J 12.9, 9.2 Hz, 1H), 2.84 (s, 2H), 2.05 (s, 3H), 1.58 (s, 9H), 1.45 (s, 3H), 1.44 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 159.4, 151.3, 145.8, 133.8, 127.2, 126.1, 125.9, 122.8, 119.0, 113.9, 84.2, 77.9, 55.3, 41.9, 37.7, 34.6, 31.7, 28.3, 13.4. Elemental Analysis Calcd for $\text{C}_{24}\text{H}_{30}\text{O}_3\text{S}$: C 72.33, H 7.59; found C 72.37, H 9.63.

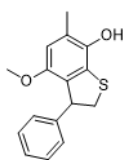
General procedure for transposition. A solution of **4** or **5** and acid ($\text{BF}_3 \cdot \text{OEt}_2$ or $\text{CF}_3\text{SO}_3\text{H}$) in dry CHCl_3 (0.03 M) was left under magnetic stirring at r.t. or 60°C until complete consumption of the starting material as monitored by TLC. The solution was then diluted with CH_2Cl_2 (30 mL) and washed with a saturated solution of NaHCO_3 (2x30 mL) and H_2O (2x30 mL). The organic layer was dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude was purified by silica gel column chromatography to yield the desired dihydrobenzo[*b*]thiophene **5** or benzothiophene **6**.



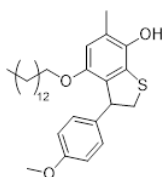
5ac. The reaction carried out following the general procedure from **4ac** (40 mg, 0.13 mmol) using $\text{BF}_3\text{Et}_2\text{O}$ (3 equiv.) at 60°C for 30 min. The crude was purified by silica gel column chromatography (CH_2Cl_2 as eluent) to yield the desired product **5ac** as light orange solid mp $111\text{--}113^\circ\text{C}$ (27 mg, yield 66%) and 16% benzo[*b*]thiophene **6a**. ^1H NMR (400 MHz, CDCl_3) δ 7.19 – 7.17 (m, 2H), 6.82 – 6.80 (m, 2H), 6.41 (s, 1H), 3.78 (s, 3H), 3.50 – 3.40 (m, 5H), 2.25 (s, 3H), 1.85 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 157.9, 150.5, 142.2, 138.8, 132.7, 129.3, 127.0, 124.2, 113.3, 111.7, 56.0, 55.8, 55.2, 50.0, 24.9, 15.9. FT-IR (CCl_4 0.05M, cm^{-1}) ν 3615, 3572, 3000, 2934, 2834, 1717, 1610, 1583, 1511, 1484, 1464, 1400, 1248, 1182. Elemental Analysis Calcd for $\text{C}_{18}\text{H}_{20}\text{O}_3\text{S}$: C 68.33, H 6.37; found C 68.47, H 6.51.



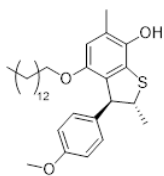
6a. The reaction carried out following the general procedure from **4ac** (51 mg, 0.16 mmol) using $\text{CF}_3\text{SO}_3\text{H}$ (1 equiv.) at 60°C for 20 min. The crude was purified by silica gel column chromatograph (petroleum ether: CH_2Cl_2 1:1 as eluent) and the second, on the mixed fraction containing the desired product, using a mixture of petroleum ether and CH_2Cl_2 1:1. The purified product was obtained as brown oil (15 mg, yield 46%). IR (CCl_4 0.05M, cm^{-1}) ν 3614, 3590, 2925, 2845, 1655, 1614, 1529, 1485, 1464, 1173. ^1H NMR (400 MHz, CDCl_3) δ 6.80 (d, J 1.2 Hz, 1H), 6.51 (s, 1H), 4.46 (bs, 1H), 3.85 (s, 3H), 2.56 (d, J 1.2 Hz, 3H), 2.35 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 150.9, 141.7, 134.1, 131.2, 129.2, 119.2, 117.6, 108.3, 55.9, 17.3, 15.6. Elemental Analysis Calcd for $\text{C}_{11}\text{H}_{12}\text{O}_2\text{S}$: C 63.44, H 5.81; found C 63.49, H 5.85.



5ad. The reaction carried out following the general procedure from **4ad** (25 mg, 0.09 mmol) using $\text{CF}_3\text{SO}_3\text{H}$ (5 equiv.) at 60°C for 7h. The crude was purified by silica gel column chromatography (petroleum ether: CH_2Cl_2 1:1 as eluent) to yield the desired product as a brown oil (6 mg, yield 25%). ^1H NMR (400 MHz, CDCl_3) δ 7.28 – 7.05 (m, 5H), 6.31 (s, 1H), 4.74 (dd, J 8.5, 2.1 Hz, 1H), 4.10 (s, 1H), 3.97 – 3.78 (m, 1H), 3.50 (s, 3H), 3.22 (dd, J 11.1, 2.1 Hz, 1H), 2.18 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 150.2, 142.7, 142.2, 129.6, 128.6, 128.3, 127.1, 126.7, 124.5, 110.6, 55.9, 50.7, 42.9, 16.0. IR (CCl_4 0.025M, cm^{-1}) ν 3614, 3576, 3063, 3028, 3001, 2932, 2854, 2833, 1602, 1593, 1490, 1464, 1453, 1405, 1186, 1034. Elemental Analysis Calcd for $\text{C}_{16}\text{H}_{16}\text{O}_2\text{S}$: C 70.56, H 5.92; found C 70.61, H 5.97.

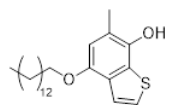


5ea. The reaction carried out following the general procedure from **4ea** (30 mg, 0.06 mmol) using $\text{BF}_3\text{Et}_2\text{O}$ (3 equiv.) at 60°C for 3h. The crude product was purified by silica-gel column chromatography (petroleum ether: CH_2Cl_2 1:2 as eluent) obtaining the purified product **5ea** as a beige oil (15 mg, yield 53%). ^1H NMR (400 MHz, CDCl_3) δ 7.16 (d, J 8.6 Hz, 2H), 6.78 (d, J 8.6 Hz, 2H), 6.34 (s, 1H), 4.76 (dd, J 8.5, 2.8 Hz, 1H), 4.18 (s, 1H), 3.92 (dd, J 11.1, 8.5 Hz, 1H), 3.81 – 3.75 (m, 1H), 3.76 (s, 3H), 3.66 – 3.60 (m, 1H), 3.26 (dd, J 11.1, 2.8 Hz, 1H), 2.23 (s, 3H), 1.52 – 1.45 (m, 2H), 1.30 – 1.17 (m, 22H), 0.88 (t, J 6.62 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 158.3, 149.6, 142.0, 135.4, 130.0, 129.2, 128.1, 124.3, 113.6, 111.5, 68.5, 55.2, 50.3, 42.7, 31.9, 29.71, 29.70, 29.68, 29.66, 29.63, 29.55, 29.4, 29.3, 29.2, 25.8, 22.7, 16.0, 14.1. IR (CDCl_3 , 0.05M, cm^{-1}) ν 3603, 2927, 2855, 1609, 1511, 1487, 1468, 1407, 1245, 1179. Elemental Analysis Calcd for $\text{C}_{30}\text{H}_{44}\text{O}_3\text{S}$: C 74.33, H 9.15; found C 74.37, H 9.18.

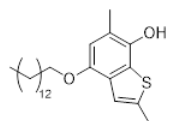


5eb. The reaction carried out following the general procedure from **4eb** (35 mg, 0.07 mmol) using $\text{BF}_3\cdot\text{Et}_2\text{O}$ (3 equiv.) at 60°C for 1h. The crude product was purified by silica-gel column chromatography (petroleum ether: CH_2Cl_2 1:2 as eluent) obtaining the purified product **5eb** as light orange oil (22 mg, yield

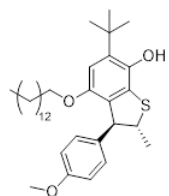
63%). ^1H NMR (400 MHz, CDCl_3) δ 7.12 (d, J 8.7 Hz, 2H), 6.77 (d, J 8.7 Hz, 2H), 6.35 (s, 1H), 4.32 (d, J 3.1 Hz, 1H), 4.16 (bs, 1H), 3.80 – 3.70 (m, 2H), 3.76 (s, 3H), 3.63 – 3.57 (m, 1H), 2.23 (s, 3H), 1.53 + 1.52 (s, 3H), 1.48 – 1.41 (m, 2H), 1.31 – 1.12 (m, 22H), 0.89 (t, J 6.6 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 158.3, 150.1, 142.1, 135.1, 128.8, 128.3, 128.1, 124.2, 113.6, 111.7, 68.6, 58.9, 55.2, 54.8, 31.9, 29.71, 29.70, 29.68, 29.66, 29.65, 29.55, 29.36, 29.34, 29.23, 25.8, 23.7, 22.7, 16.0, 14. IR (CDCl_3 , 0.05M, cm^{-1}) ν 3571, 2928, 2856, 1610, 1511, 1488, 1468, 1408, 1304, 1245, 1178, 1108, 1036. MS (ESI): m/z 497.3 $[\text{M-H}]^-$. Elemental Analysis Calcd for $\text{C}_{31}\text{H}_{46}\text{O}_3\text{S}$: C 74.65, H 9.30; found C 74.68, H 9.35.



6b. The reaction carried out following the general procedure from **4ee** (20 mg, 0.047 mmol) using $\text{CF}_3\text{SO}_3\text{H}$ (0.5 equiv.) at room temperature for 30 min, was gradually heated to 30 °C (3h) and then to 60 °C (1h). The crude was purified by silica gel column chromatography, using a mixture of petroleum ether and CH_2Cl_2 1:2 as eluent. The purified product was obtained as a light brown oil (4 mg, yield 23%). IR (CDCl_3 , 0.05M, cm^{-1}) ν 3602, 2928, 2856, 1466, 1409, 1262, 1099. MS (ESI): m/z 375.2 $[\text{M-H}]^-$. ^1H NMR (400 MHz, CDCl_3) δ 7.44 (d, J 5.8 Hz, 1H), 7.25 (d, J 5.8 Hz, 1H), 6.54 (s, 1H), 4.54 (bs, 1H), 4.04 (t, J 6.5 Hz, 2H), 2.36 (s, 3H), 1.87 – 1.80 (m, 2H), 1.52 – 1.46 (m, 2H), 1.26 (bs, 20H), 0.90 – 0.86 (m, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 148.6, 141.7, 135.0, 134.7, 123.8, 121.4, 117.8, 109.0, 68.9, 31.9, 29.7, 29.6, 29.4 (3 signals for 9 non-equivalent CH_2 groups), 26.2, 22.7, 15.7, 14.1. Elemental Analysis Calcd for $\text{C}_{23}\text{H}_{36}\text{O}_2\text{S}$: C 73.35, H 9.64; found C 73.38, H 9.69.



6c. The reaction carried out following the general procedure from **5eb** (18 mg, 0.036 mmol) using $\text{CF}_3\text{SO}_3\text{H}$ (0.5 equiv.) at 60°C for 2h. The crude product was purified by silica-gel column chromatography (petroleum ether: CH_2Cl_2 1:1 as eluent) obtaining a light brown oil (7 mg, yield 50%). ^1H NMR (400 MHz, CDCl_3) δ 7.08 (s, 1H), 6.50 (s, 1H), 4.45 (bs, 1H), 4.00 (t, J 6.5 Hz, 2H), 2.56 (s, 3H), 2.33 (s, 3H), 1.85 – 1.78 (m, 2H), 1.52 – 1.45 (m, 2H), 1.35 – 1.26 (m, 20H), 0.88 (t, J 6.7 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 147.8, 141.3, 138.4, 131.1, 129.2, 119.0, 117.0, 109.2, 68.9, 31.9, 29.7, 29.6, 29.5, 29.4 (4 signals for 9 non-equivalent CH_2 groups), 26.2, 22.7, 16.1, 15.7, 14.1. IR (CDCl_3 , 0.05M, cm^{-1}) ν 3602, 2928, 2856, 1469, 1277, 1229, 1139, 1039. MS (ESI): m/z 389.3 $[\text{M-H}]^-$. Elemental Analysis Calcd for $\text{C}_{24}\text{H}_{38}\text{O}_2\text{S}$: C 73.80, H 9.81; found C 73.84, H 9.87.



5fb. The reaction carried out following the general procedure from **4fb** (34 mg, 0.063 mmol, 0.001 M) using $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (3 equiv.) at 60°C for 22h after was added more $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (3 equiv.) and the reaction was left for additional 8h at 60°C. The crude was purified by silica-gel column chromatography using before a mixture of petroleum ether and CH_2Cl_2 8:1 as eluent to 1:1 to obtain the desired product as an orange solid (15 mg, yield 45%). ^1H NMR (400 MHz, CDCl_3) δ 7.13 (d, J 8.6 Hz, 2H), 6.79 (d, J 8.6 Hz, 2H), 6.51 (s, 1H), 4.32 (d, J 3.7 Hz, 1H), 4.06 (s, 1H), 3.80 – 3.75 (m, 2H), 3.76 (s, 3H), 3.63 – 3.58 (m, 1H), 1.53 + 1.51 (s, 3H), 1.47- 1.43 (m, 2H), 1.40 (s, 9H), 1.32 – 1.14 (m, 22H), 0.88 (t, J 6.6 Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 158.3, 149.7, 142.5, 136.9, 135.0, 129.7, 128.6, 128.3, 113.6, 109.0, 68.7, 59.2, 55.2, 55.1, 34.9, 31.9, 29.7, 29.6, 29.4 (2 signals for

9 non-equivalent CH₂), 25.8, 23.2, 22.7, 14.1. Elemental Analysis Calcd for C₃₄H₅₂O₃S: C 75.51, H 9.69; found C 75.55, H 9.71.

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Supplementary Material

Synthetic procedures and NMR spectra are available as supplementary material of this article.

References

1. Stubbe, J.; van Der Donk, W. A. *Chem. Rev.* **1998**, *98*, 705.
<https://doi.org/10.1021/cr9400875>
2. Hensley, K.; Robinson, K. A.; Gabbita, S. P.; Salsman, S.; Floyd, R. A. *Free Radic. Biol. Med.* **2000**, *28*, 1456.
[https://doi.org/10.1016/S0891-5849\(00\)00252-5](https://doi.org/10.1016/S0891-5849(00)00252-5)
3. Stubbe, J. J. *Chem. Commun.* **2003**, 2511.
<https://doi.org/10.1039/b307617m>
4. Halliwell, B.; Gutteridge, J. M. C. *Antioxidants in Nutrition, Health, and Disease*, Oxford University Press Inc., New York **1996**.
5. Halliwell, B.; *Encyclopedia of life sciences*, 2001, Nature Publishing Group, 1-7.
6. Vroegop, S. M.; Decker, D. E.; Buxser, S. E. *Free Radic. Biol. Med.* **1995**, *18*, 141.
7. Breen, A.P.; Murphy, J. A. *Free Radic. Biol. Med.* **1995**, *18*, 1033.
[https://doi.org/10.1016/0891-5849\(94\)00209-3](https://doi.org/10.1016/0891-5849(94)00209-3)
8. Pace, G. W.; Leaf, C. D. *Free Radic. Biol. Med.* **1995**, *19*, 523.
[https://doi.org/10.1016/0891-5849\(95\)00047-2](https://doi.org/10.1016/0891-5849(95)00047-2)
9. Aruoma, O. I. *Free Radic. Biol. Med.* **1996**, *20*, 675.
[https://doi.org/10.1016/0891-5849\(95\)02110-8](https://doi.org/10.1016/0891-5849(95)02110-8)
10. Schwarz, K. B. *Free Radic. Biol. Med.* **1996**, *21*, 641.
[https://doi.org/10.1016/0891-5849\(96\)00131-1](https://doi.org/10.1016/0891-5849(96)00131-1)
11. Viglianisi, C.; Menichetti, S.; Morelli, P.; Baschieri, A.; Amorati, A. *Heteroatom Chemistry* **2018**, e21466.
<https://doi.org/10.1002/hc.21466>
<https://doi.org/10.1002/hc.21466>
12. Ingold, K. U. *Chem. Rev.* **1961**, *61*, 563.
<https://doi.org/10.1021/cr60214a002>
13. Finkel, T.; Holbrook N. J., *Nature*, **2000**, *408*, 239.
<https://doi.org/10.1038/35041687>
14. Lucarini, M.; Pedulli, G. F. *Chem. Soc. Rev.* **2010**, *39*, 2106.
<https://doi.org/10.1039/b901838g>

- 15 Middleton Jr, E.; Kandaswarni, C. in *The Flavonoids Advances in Research Since 1986*, (Ed. J. B. Harborne), Chapman & Hall, London, **1994**, pp. 619–645.
- 16 Bohm, B. A. *Introduction to Flavonoids*, Harwood Academic Publishers, Amsterdam, **1998** and references cited therein.
- 17 Dixon, R. A.; Ferreira, D. *Phytochemistry* **2002**, *60*, 205.
[https://doi.org/10.1016/S0031-9422\(02\)00116-4](https://doi.org/10.1016/S0031-9422(02)00116-4)
- 18 Burton, G. W.; Ingold, K. U. *Acc. Chem. Res.* **1986**, *19*, 194.
<https://doi.org/10.1021/ar00127a001>
- 19 Jiang, Q. *Free Rad. Biol. Med.* **2014**, *72*, 76.
<https://doi.org/10.1016/j.freeradbiomed.2014.03.035>
- 20 Viglianisi, C.; Amorati, R.; Di Pietro, L.; Menichetti, S. *Chem. Eur. J.* **2015**, *21*, 16639.
<https://doi.org/10.1002/chem.201502650>
- 21 Menichetti, S.; Amorati, R.; Meoni, V.; Tofani, L.; Caminati, G.; Viglianisi, C. *Org. Lett.* **2016**, *18*, 5464.
<https://doi.org/10.1021/acs.orglett.6b02557>
- 22 Capozzi, G.; Falciani, C.; Menichetti, S.; Nativi, C. *J. Org. Chem.* **1997**, *62*, 2611.
<https://doi.org/10.1021/jo962014m>
- 23 Capozzi, G.; Lo Nostro, P.; Menichetti, S.; Nativi, C.; Sarri, P. *Chem. Commun.* **2001**, 551.
<https://doi.org/10.1039/b100359n>
- 24 Menichetti, S.; Aversa, M. C.; Cimino, F.; Contini, A.; Viglianisi, C.; Tomaino, A. *Org. Biomol. Chem.* **2005**, *3*, 3066.
<https://doi.org/10.1039/b507496g>
- 25 Lodovici, M.; Menichetti, S.; Viglianisi, C.; Caldini, S.; Giuliani, E. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 1957.
<https://doi.org/10.1016/j.bmcl.2005.12.078>
- 26 Amorati, R.; Cavalli, A.; Fumo, M. G.; Masetti, M.; Menichetti, S.; Pagliuca, C.; Pedulli, G. F.; Viglianisi, C. *Chem. Eur. J.* **2007**, *13*, 8223.
<https://doi.org/10.1002/chem.200700309>
- 27 Amorati, R.; Catarzi, F.; Menichetti, S.; Pedulli, G. F.; Viglianisi, C. *J. Am. Chem. Soc.* **2008**, *130*, 237.
<https://doi.org/10.1021/ja075554h>
- 28 Menichetti, S.; Amorati, R.; Bartolozzi, M. G.; Pedulli, G. F.; Salvini, A.; Viglianisi, C. *Eur. J. Org. Chem.* **2010**, 2218.
<https://doi.org/10.1002/ejoc.200901493>
- 29 Zahalka, H. A.; Robillard, B.; Hughes, L.; Luszyk, J.; Burton, G. W.; Janzen, E. G.; Kotake, Y.; Ingold, K. U. *J. Org. Chem.* **1988**, *53*, 3739.
<https://doi.org/10.1021/jo00251a014>
- 30 Politzer, P.; Murray, J. S.; Clark, T. *Phys. Chem. Chem. Phys.* **2013**, *15*, 11178.
<https://doi.org/10.1039/c3cp00054k>
- 31 Beno, B. R.; Yeung, K.-S.; Bartberger, M. D.; Pennington, L. D.; Meanwell, N. A. *J. Med. Chem.* **2015**, *58*, 4383.
<https://doi.org/10.1021/jm501853m>
- 32 Fick, R. J.; Kroner, G. M.; Nepal, B.; Magnani, R.; Horowitz, S.; Houtz, R. L.; Scheiner, S.; Trievel, R. C. *ACS Chem. Biol.* **2016**, *11*, 748.
<https://doi.org/10.1021/acscchembio.5b00852>
- 33 Turbiez, M.; Frere, P.; Allain, M.; Videlot, C.; Ackermann, J.; Roncali, J. *Chem. Eur. J.* **2005**, *11*, 3742–3752.
<https://doi.org/10.1002/chem.200401058>

- 34 Fanfrlik, J.; Prada, A.; Padelkova, Z.; Pecina, A.; Machacek, J.; Lepsik, M.; Holub, J.; Ruzicka, A.; Hnyk, D.; Hobza, P. *Angew. Chem., Int. Ed.* **2014**, *53*, 10139.
<https://doi.org/10.1002/anie.201405901>
- 35 Nziko, V. de P. N.; Scheiner, S. *J. Org. Chem.* **2015**, *80*, 2356.
<https://doi.org/10.1021/acs.joc.5b00012>
- 36 Viglianisi, C.; Marcantoni, E.; Carapacchi, V.; Menichetti, S.; Marsili, L. *Eur. J. Org. Chem.* **2014**, 6405.
<https://doi.org/10.1002/ejoc.201402894>
- 37 Viglianisi, C.; Vasa, K.; Tanini, D.; Capperucci, A.; Amorati, R.; Valgimigli, L.; Baschieri, A.; Menichetti, S. *Chem. Eur. J.* **2019**, *25*, 9108.
- 38 Anchel, M.; Blatt, A. H. *J. Am. Chem. Soc.* **1941**, *63*, 1948.
<https://doi.org/10.1021/ja01852a045>
- 39 Loudon, J. D.; Razdan, R. K. *J. Chem. Soc.* **1954**, 4299.
<https://doi.org/10.1039/jr9540004299>
- 40 Anastasis, P.; Brown, P. E. *J. Chem. Soc. Perkin Trans. 1* **1982**, 2013.
<https://doi.org/10.1039/p19820002013>
- 41 Bellamy, D. Chazan, J. B.; Ou, K. *Tetrahedron* **1983**, *39*, 2803.
[https://doi.org/10.1016/S0040-4020\(01\)82450-9](https://doi.org/10.1016/S0040-4020(01)82450-9)
- 42 Tanaka, T.; Miyaguchi, M.; Mochisuki, R. K.; Tanaka, S.; Okamoto, M.; Kitajima, Y.; Miyazaki, T. *Heterocycles* **1987**, *25*, 463.
<https://doi.org/10.3987/S-1987-01-0463>
- 43 Levy, J.-N.; Latham, C. M.; Roisin, L.; Kandziora, N.; Di Fruscia, P.; White, A. J. P.; Woodward, S.; Fuchter, M. *J. Org. Biomol. Chem.* **2012**, *10*, 512
<https://doi.org/10.1039/C1OB06780J>
- 44 Killander, D.; Sterner, O. *Eur. J. Org. Chem.* **2014**, 6507.
<https://doi.org/10.1002/ejoc.201402722>
- 45 Burton, G. W.; Doba, T.; Gabe, E. J.; Hughes, L.; Lee, F. L.; Prasad, L.; Ingold, K. U. *J. Am. Chem. Soc.* **1985**, *107*, 7053.
<https://doi.org/10.1021/ja00310a049>
- 46 Lucarini, M.; Pedulli, G. F. *Chem. Soc. Rev.*, **2010**, *39*, 2106.
<https://doi.org/10.1039/b901838g>
- 47 Nam, T.-g.; Rector, C. L.; Kim, Hye-y.; Sonnen, A. F.-P. Meyer, R.; Nau, W. M.; Atkinson, J.; Rintoul, G.; Pratt, D. A.; Porter, N. A. *J. Am. Chem. Soc.* **2007**, *129*, 10211.
<https://doi.org/10.1021/ja072371m>
- 48 Amorati, R.; Baschieri, A. Valgimigli, L. *J. Chem.* **2017**, 6369358.
<https://doi.org/10.1155/2017/6369358>
- 49 Viglianisi, C.; Bartolozzi, M. G.; Pedulli, G. F.; Amorati, R.; Menichetti, S. *Chem. Eur. J.* **2011**, *17*, 12396.
<https://doi.org/10.1002/chem.201101146>