

Supplementary Material

Comparison of affinity ranking by target-directed dynamic combinatorial chemistry and surface plasmon resonance

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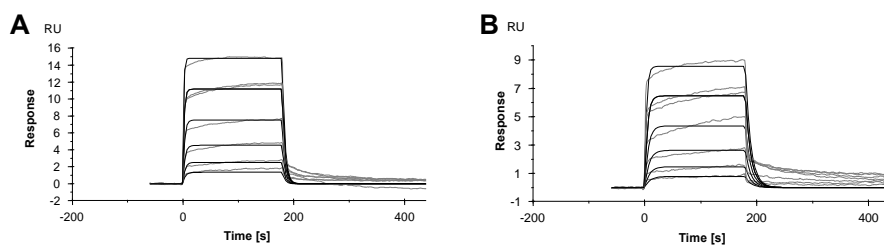
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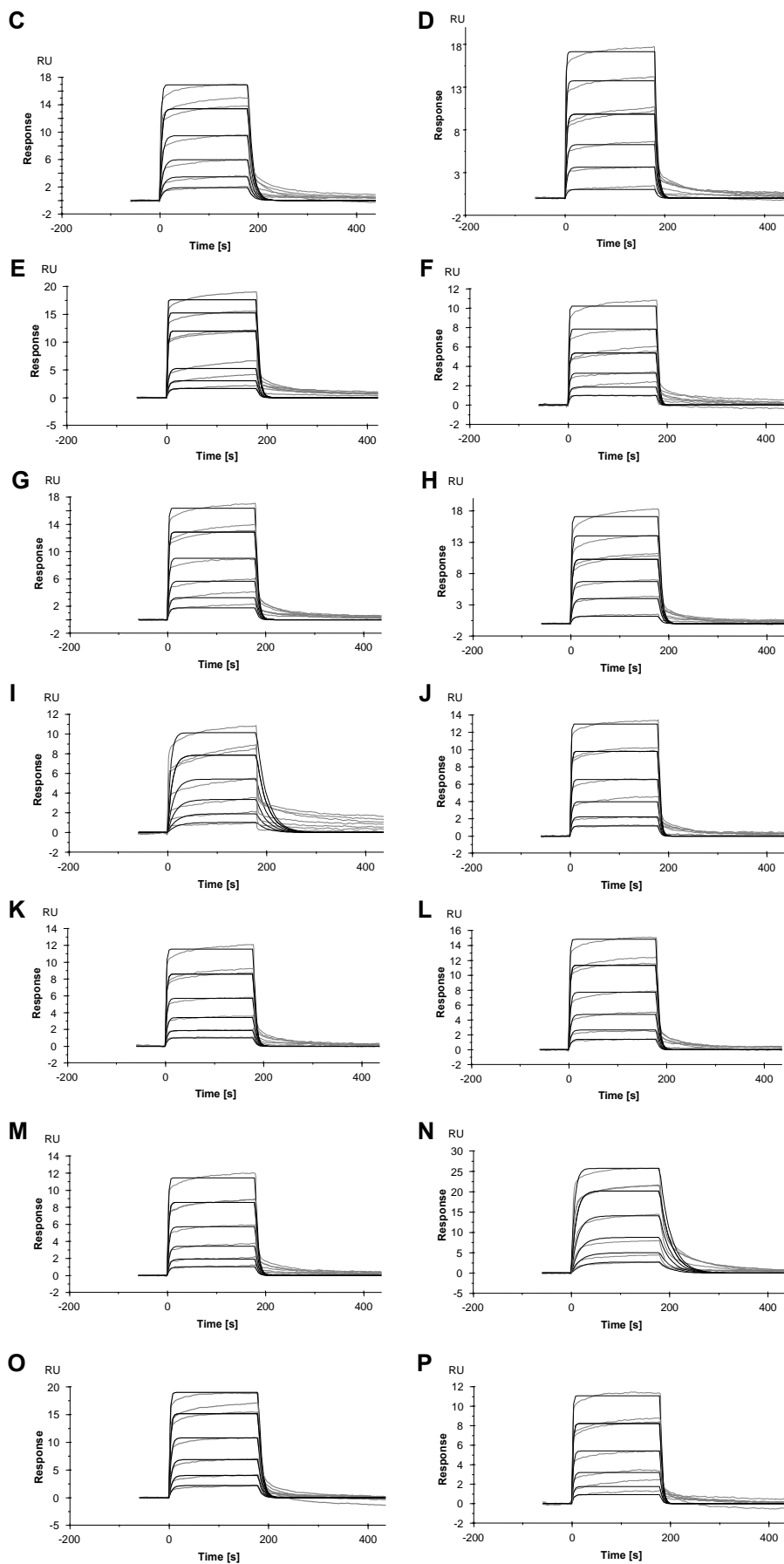
Surface Plasmon Resonance Experiments

According to an established procedure,^{S1} SPR experiments were conducted using a Biacore T200 system (GE Healthcare). In brief, dilution series with two-fold increasing concentrations were delivered over a streptavidin (SA) chip with immobilized FimH_{FL-B}. The reference cell was capped with biotin-poly(ethyleneglycol)amine. Starting from stock solutions of compounds (50 mM in DMSO), dilution series were prepared in buffer (HBS-EP; GE Healthcare). Compounds were injected for 180 s at a flow rate of 30 μ L/min, followed by an 800 s dissociation phase. The sensorgrams were referenced and blank subtracted and fitted according to a 1:1 binding model. Complete data are given in Table S1 and the sensorgrams are depicted in Figure S1.

Table S1. Results from SPR measurements.

| | k_{on} [1/Ms] | Compound | K_D [nM] | $t_{1/2}$ [s] |
|-----------|-------------------|----------|------------|---------------|
| 3a | $6.18 \cdot 10^5$ | 0.22 | 359 | 3.13 |
| 3c | $2.93 \cdot 10^5$ | 0.11 | 358 | 6.60 |
| 3d | $4.31 \cdot 10^5$ | 0.12 | 267 | 6.03 |
| 3e | $3.92 \cdot 10^5$ | 0.19 | 492 | 3.59 |
| 3g | $4.41 \cdot 10^5$ | 0.20 | 461 | 3.41 |
| 3h | $4.41 \cdot 10^5$ | 0.28 | 642 | 2.45 |
| 3j | $4.61 \cdot 10^5$ | 0.21 | 462 | 3.26 |
| 3k | $4.23 \cdot 10^5$ | 0.18 | 427 | 3.84 |
| 3l | $9.52 \cdot 10^5$ | 0.05 | 508 | 14.3 |
| 3m | $5.86 \cdot 10^5$ | 0.28 | 484 | 2.44 |
| 3n | $5.62 \cdot 10^5$ | 0.22 | 390 | 3.16 |
| 3o | $4.87 \cdot 10^5$ | 0.21 | 440 | 3.24 |
| 3p | $5.35 \cdot 10^5$ | 0.20 | 377 | 3.44 |
| 3q | $1.56 \cdot 10^5$ | 0.04 | 286 | 15.6 |
| 3r | $4.49 \cdot 10^5$ | 0.15 | 337 | 4.57 |
| 3s | $4.96 \cdot 10^5$ | 0.27 | 536 | 2.60 |
| 3t | $3.09 \cdot 10^5$ | 0.12 | 376 | 5.96 |





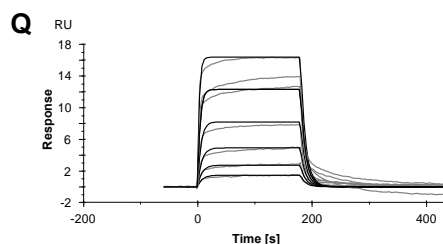


Figure S1. Multi-cycle kinetics of FimH_{FL-B} with two-fold increasing concentrations of A) **3a** (24-750nM); B) **3c** (24-750 nM); C) **3d** (24-750 nM); D) **3e** (24-1500 nM, without 47 nM); E) **3g** (39-2500 nM); F) **3h** (47-1500 nM); G) **3j** (39-1250 nM); H) **3k** (24-1500 nM, without 47 nM); I) **3l** (40-1250 nM); J) **3m** (31-1000 nM); K) **3n** (24-750 nM); L) **3o** (31-1000 nM); M) **3p** (24-750 nM); N) **3q** (24-750 nM); O) **3r** (31-1000 nM); P) **3s** (31-1000 nM); Q) **3t** (24-750 nM).

Fluorescence Polarization Assay

Experiments were conducted as previously described,^{S2-3} using a non-biotinylated version of the full-length FimH protein (FimH_{FL}) and a fluorescently labeled FimH antagonist (**11**, Figure S2),^{S2} whose $K_D = 137$ nM for FimH_{FL} has been previously determined in a direct binding assay.^{S3} Stock solutions of the compounds at 50 mM in DMSO were prepared. Starting from 600 μ M, 1:2 dilution series were prepared in assay buffer (20 mM HEPES buffer, 150 mM NaCl, 50 μ g/mL BSA, pH 7.4). Measurements were done at constant concentrations of FimH_{FL} (300 nM) and fluorescently labeled antagonist (10 nM). The mixtures were incubated for 1 h in 96-well-plates (Corning, flat bottom, non-binding surface). Fluorescence polarization was measured with a Synergy™ H1 Multi-Mode microplate reader (BioTek Instruments). Equilibrium dissociation constants were determined using Prism (GraphPad Software) and the Wang equation.^{S4} The results are depicted in Table S2.

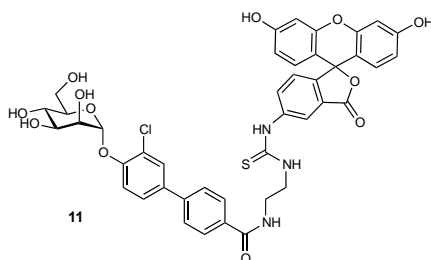


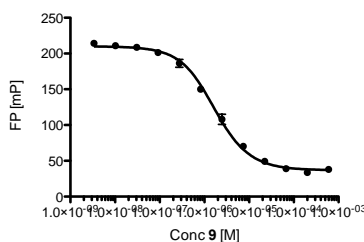
Figure S2. Fluorescently labeled FimH antagonist **11** used in the fluorescence polarization assay.^{S2,S3}

Table S2. Affinities obtained by fluorescent polarization assay.

| Compound | Curve | K_D [nM] |
|----------|-------|------------|
| 3t | | 515.0 |
| 5 | | 783.6 |
| 6 | | 1097 |
| 7 | | 3498 |
| 8 | | 2828 |
| 9 | | 1673 |

10

447.5



HPLC Traces of DCC Experiments

Purity of Target Compounds

HRMS of Target Compounds

The LC/HRMS analysis were carried out using an Agilent 1100 LC equipped with a photodiode array detector and a Micromass QTOF I equipped with a 4 GHz digital-time converter. The results are summarized in Table S9.

Table S7. Results of HRMS analysis of acylhydrazones and bioisosteres.

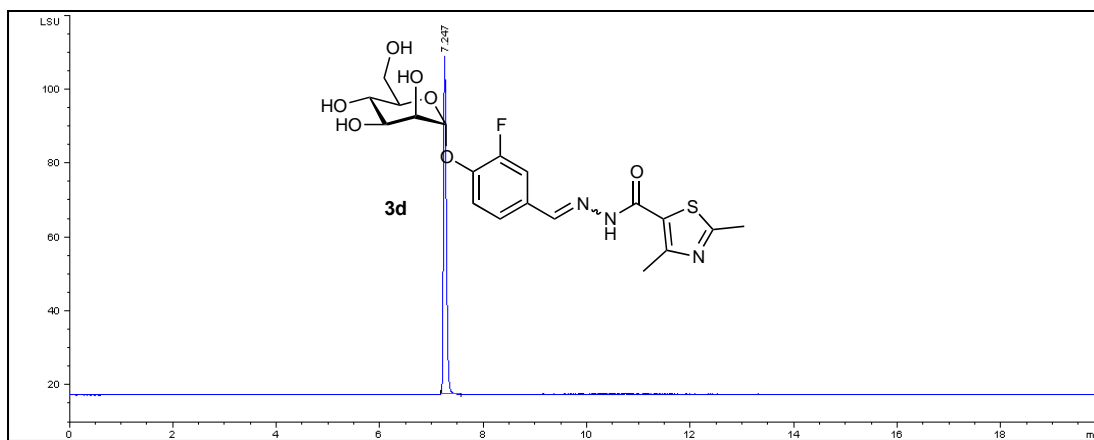
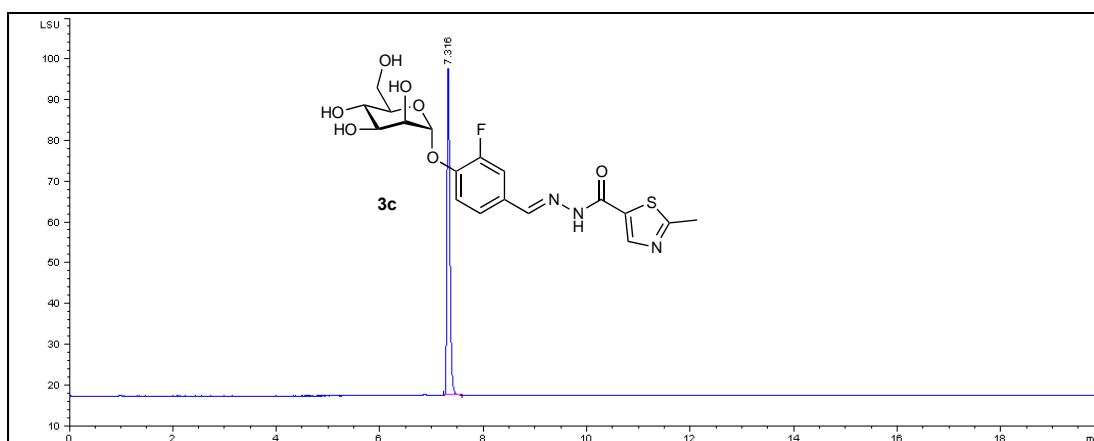
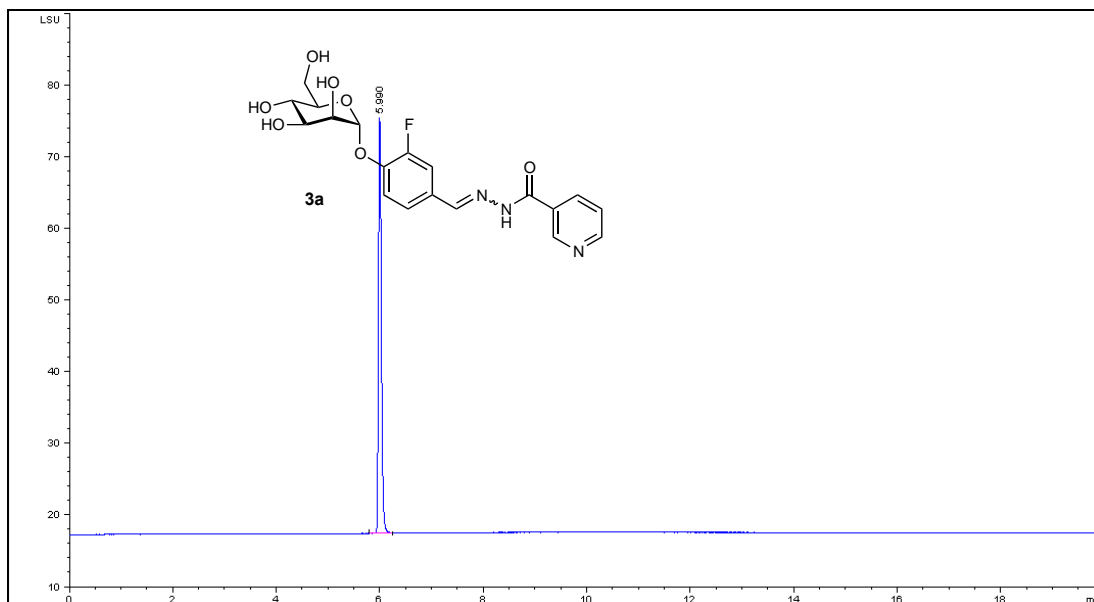
| Compound | Formula for [M+Na] ⁺ | HRMS [m/z] | |
|----------|--|------------|----------|
| | | calcd | found |
| 3a | C ₁₉ H ₂₀ FN ₃ NaO ₇ | 444.1183 | 444.1181 |
| 3c | C ₁₈ H ₂₀ FN ₃ NaO ₇ S | 464.0904 | 464.0905 |
| 3d | C ₁₉ H ₂₂ FN ₃ NaO ₇ S | 478.1060 | 478.1061 |
| 3e | C ₁₉ H ₁₉ ClFN ₃ NaO ₇ | 478.0793 | 478.0799 |
| 3g | C ₁₉ H ₂₂ FN ₃ NaO ₇ | 446.1339 | 446.1341 |
| 3h | C ₁₉ H ₂₁ FN ₂ NaO ₈ | 447.1182 | 447.1182 |
| 3j | C ₂₁ H ₂₃ FN ₂ NaO ₈ | 473.1336 | 473.1336 |
| 3k | C ₂₂ H ₂₂ FN ₃ NaO ₇ | 482.1339 | 482.1340 |
| 3l | C ₂₃ H ₂₄ FN ₃ NaO ₇ | 496.1496 | 496.1496 |
| 3m | C ₂₁ H ₂₃ FN ₂ NaO ₇ | 457.1387 | 457.1387 |
| 3n | C ₁₉ H ₂₁ FN ₂ NaO ₇ S | 463.0951 | 463.0954 |
| 3o | C ₂₀ H ₂₀ ClFN ₂ NaO ₇ | 477.0841 | 477.0841 |
| 3p | C ₁₉ H ₂₀ ClFN ₂ NaO ₇ S | 497.0561 | 497.0561 |
| 3q | C ₁₈ H ₁₈ ClFN ₂ NaO ₇ S | 483.0405 | 483.0406 |
| 3r | C ₂₄ H ₂₃ FN ₂ NaO ₇ | 493.1387 | 493.1388 |
| 3s | C ₂₁ H ₂₀ F ₄ N ₂ NaO ₇ | 511.1104 | 511.1107 |
| 3t | C ₂₂ H ₂₀ ClFN ₂ NaO ₇ S | 533.0561 | 533.0562 |
| 5 | C ₂₀ H ₂₃ FN ₂ NaO ₇ | 445.1387 | 445.1386 |
| 6 | C ₂₀ H ₂₃ FN ₂ NaO ₇ | 445.1387 | 445.1392 |
| 7 | C ₂₀ H ₂₃ FN ₂ NaO ₇ | 445.1387 | 445.1385 |
| 8 | C ₂₀ H ₂₃ FN ₂ NaO ₆ S | 461.1159 | 461.1161 |
| 9 | C ₂₀ H ₂₃ FN ₂ NaO ₆ S | 461.1159 | 461.1160 |
| 10 | C ₂₁ H ₂₂ FNNaO ₈ | 458.1227 | 458.1227 |

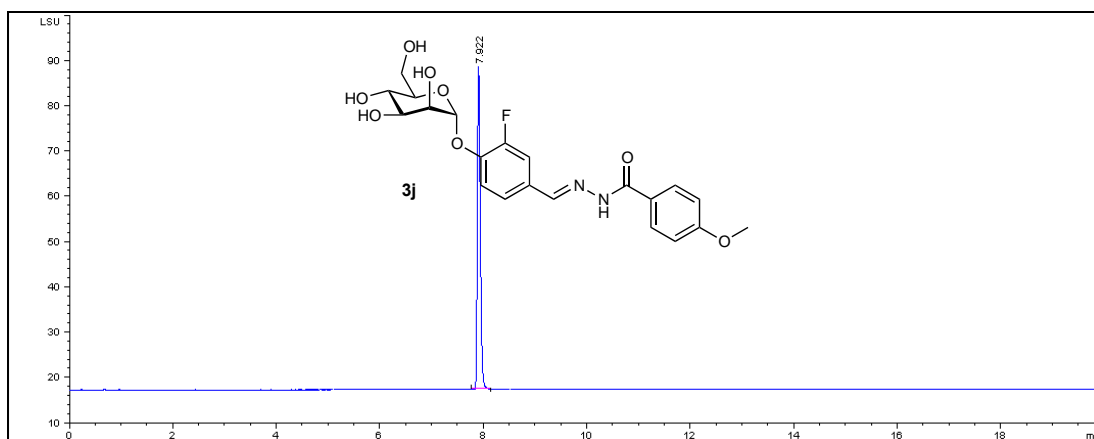
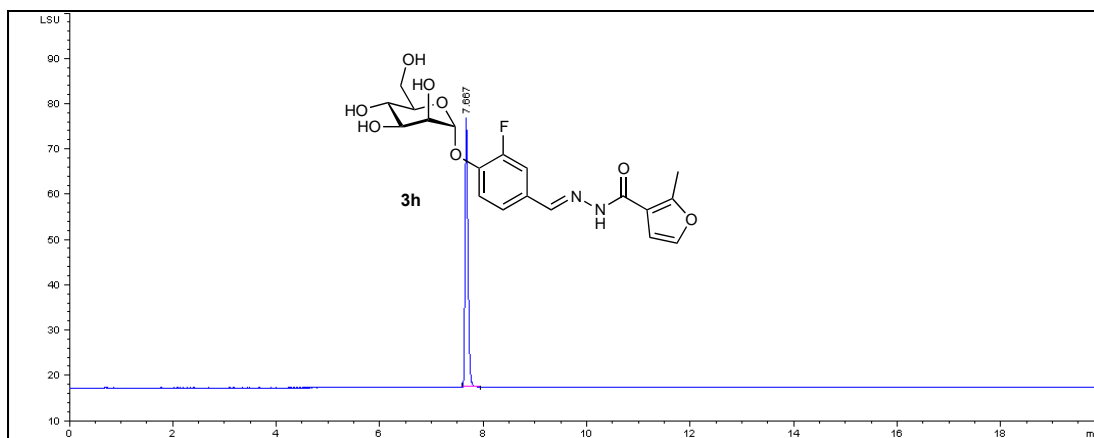
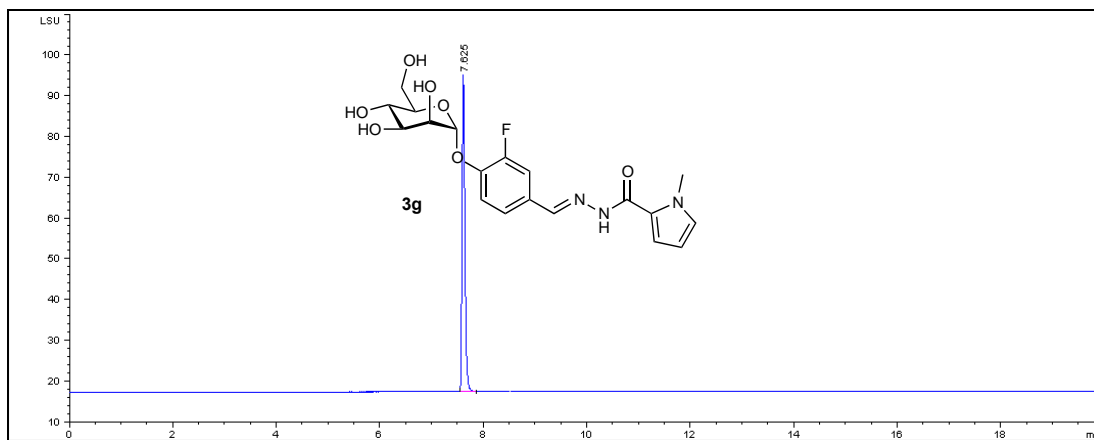
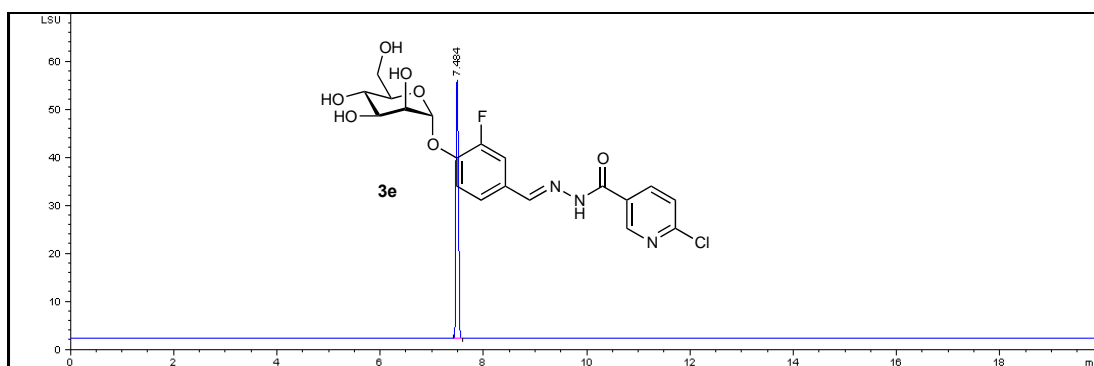
HPLC of Target Compounds

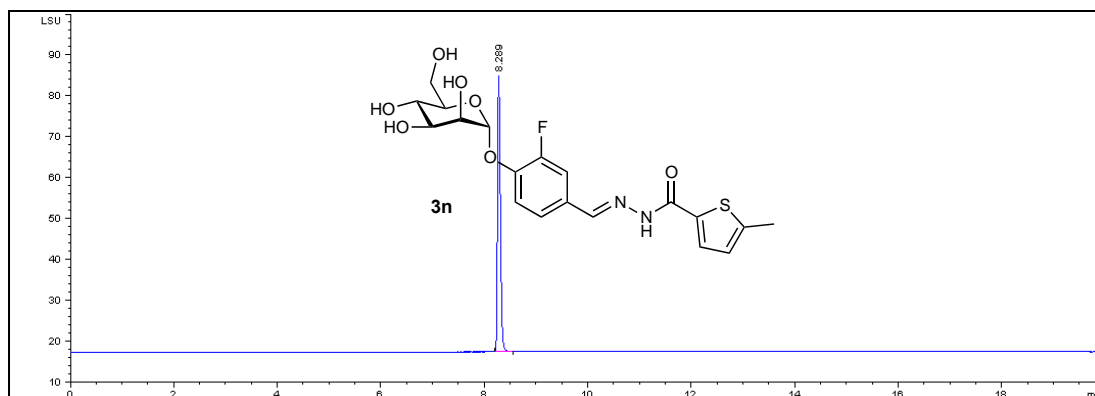
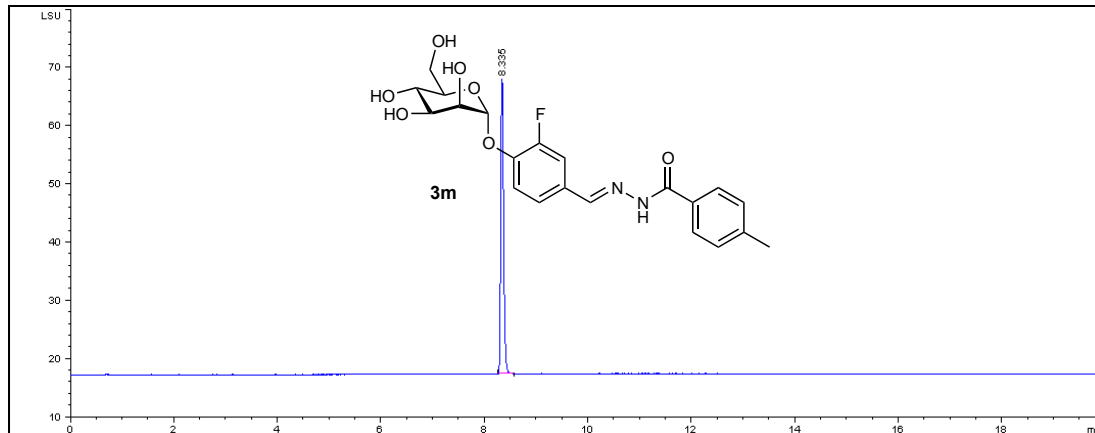
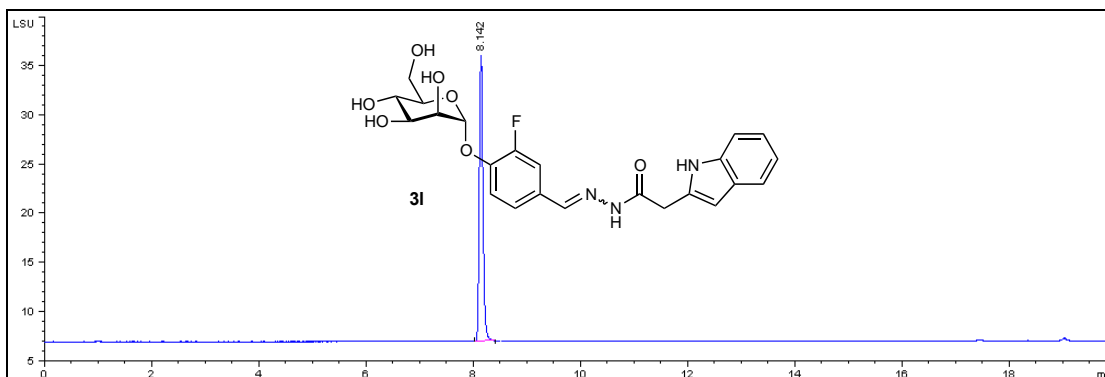
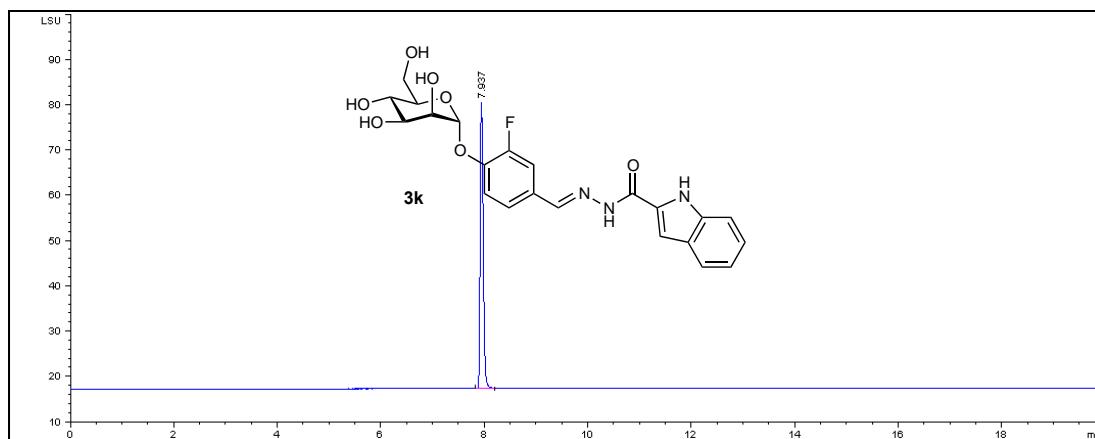
System: Agilent 1100/1200 with UV detector (190-410 nm) and Agilent 380 ELSD detector. Column: Waters Atlantis T3, 3 μ m, 2.1 \times 100 mm (Waters Corporation). A: H₂O + 0.01% TFA; B: MeCN + 0.01% TFA. Detection: Light scattering (Nebulizer control 70%, drift tube temperature 50 °C, gas pressure 50 psi, gain 500). Gradient: 5% B \rightarrow 95% B (20 min); flow rate: 0.5 mL/min. The results of the purity analysis are summarized in Table S8.

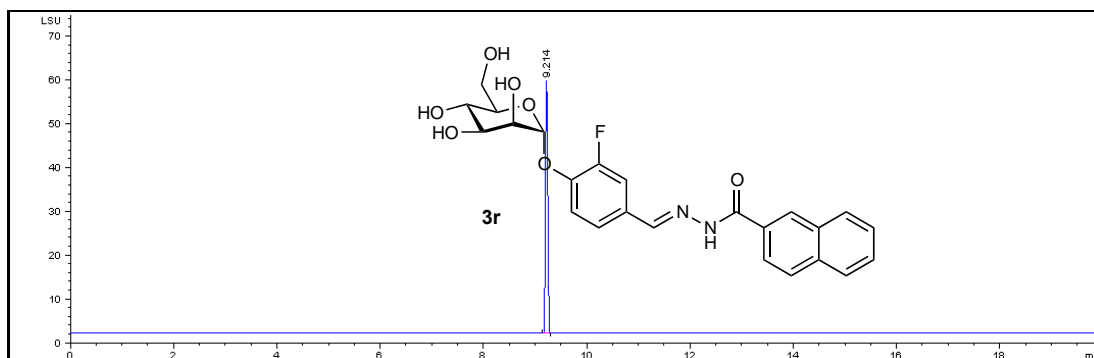
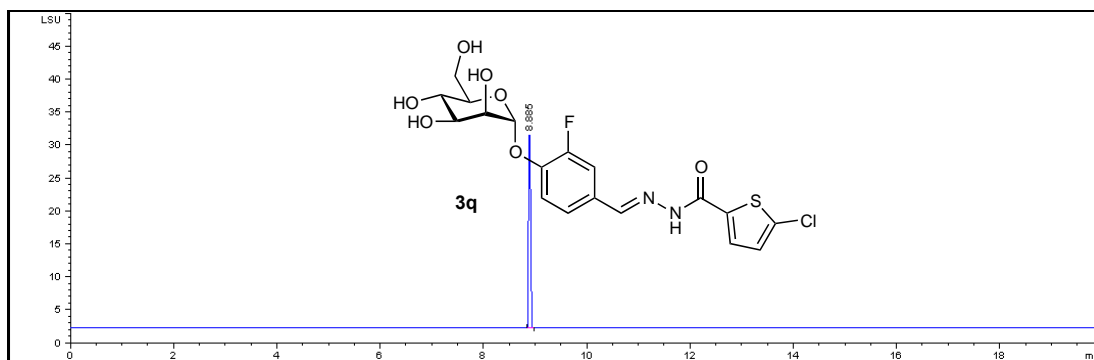
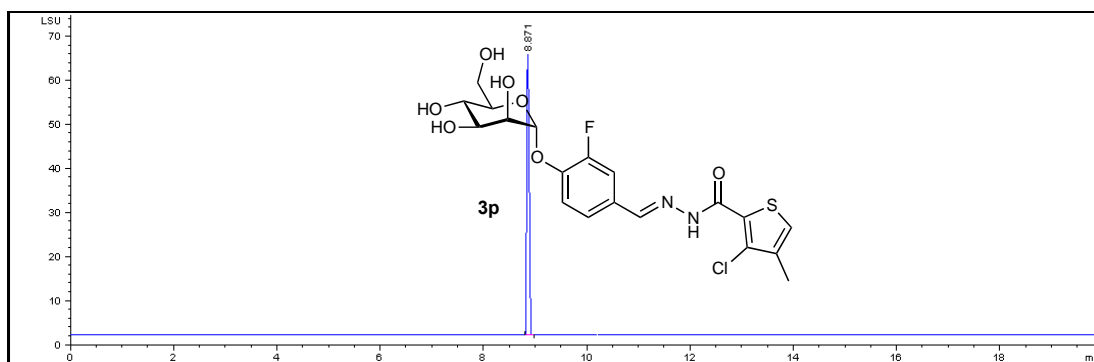
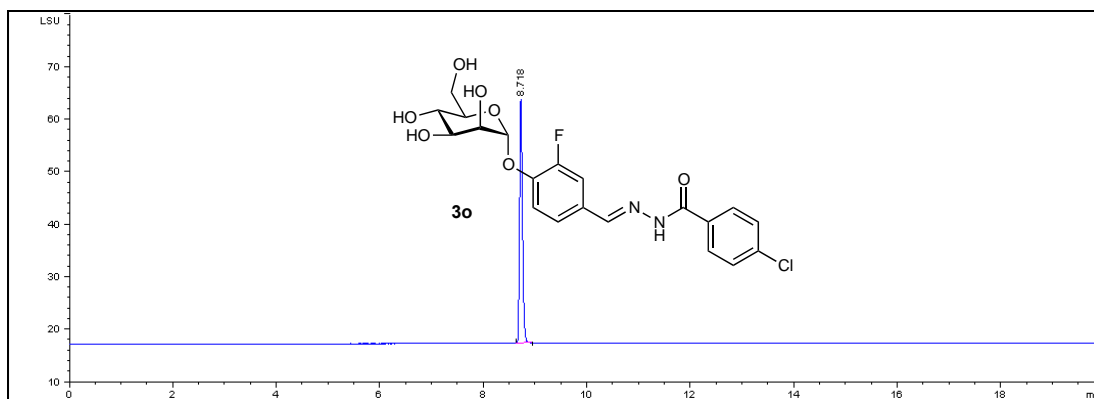
Table S8. HPLC analysis of target compounds.

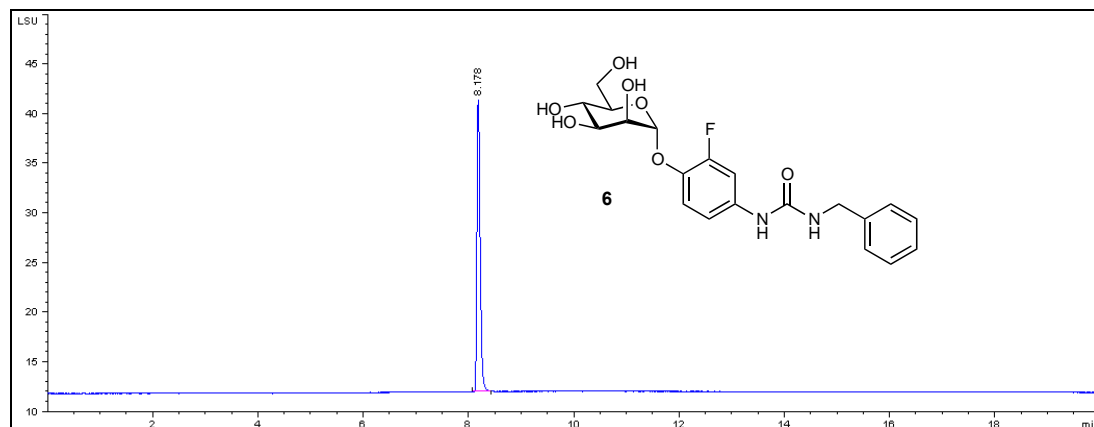
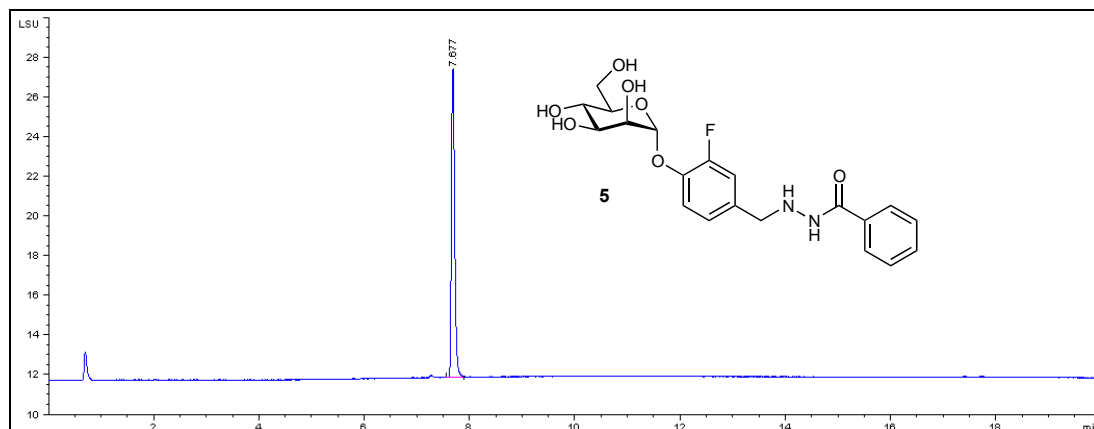
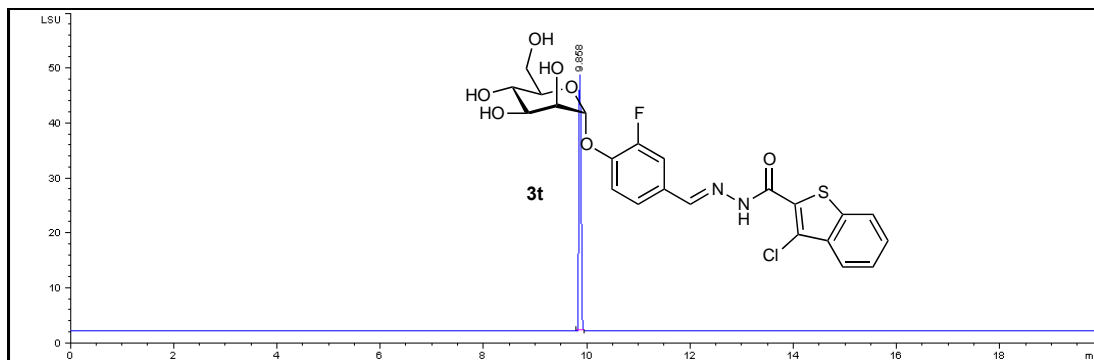
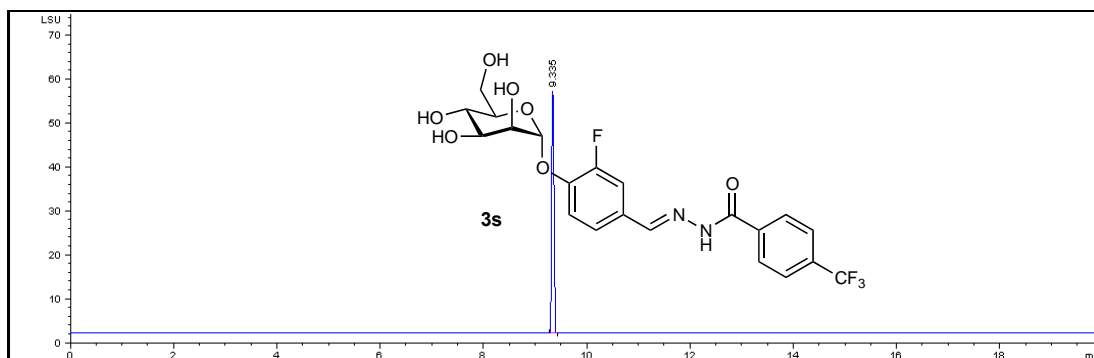
| Compound | Retention time [min] | Purity (%) |
|-----------------|----------------------|------------|
| 3a | 5.990 | >99.50 |
| 3c | 7.316 | >99.50 |
| 3d | 7.247 | >99.50 |
| 3e | 7.484 | >99.50 |
| 3f ¹ | 7.627 | >99.50 |
| 3g | 7.625 | >99.50 |
| 3h | 7.667 | >99.50 |
| 3j | 7.922 | >99.50 |
| 3k | 7.937 | >99.50 |
| 3l | 8.142 | >99.50 |
| 3m | 8.335 | >99.50 |
| 3n | 8.289 | >99.50 |
| 3o | 8.718 | >99.50 |
| 3p | 8.871 | >99.50 |
| 3q | 8.885 | >99.50 |
| 3r | 9.214 | >99.50 |
| 3s | 9.335 | >99.50 |
| 3t | 9.858 | >99.50 |
| 3u ¹ | 10.087 | >99.50 |
| 5 | 7.677 | >99.50 |
| 6 | 8.178 | >99.50 |
| 7 | 8.380 | >99.50 |
| 8 | 8.849 | >99.50 |
| 9 | 7.960 | >99.50 |
| 10 | 7.962 | >99.50 |

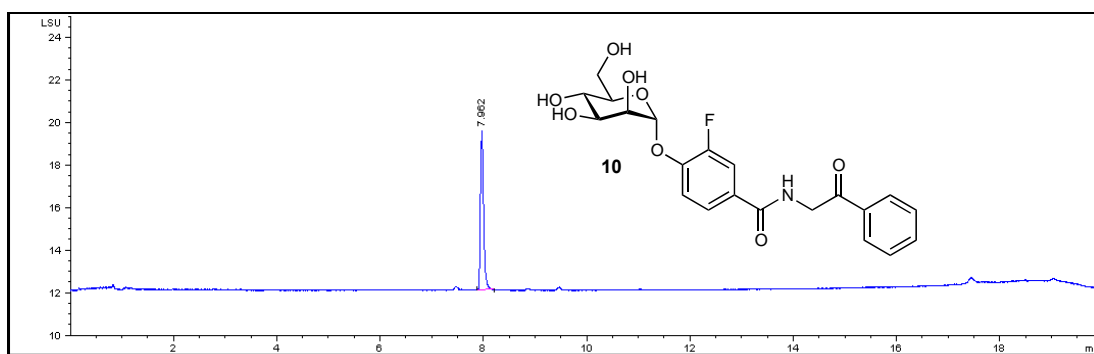
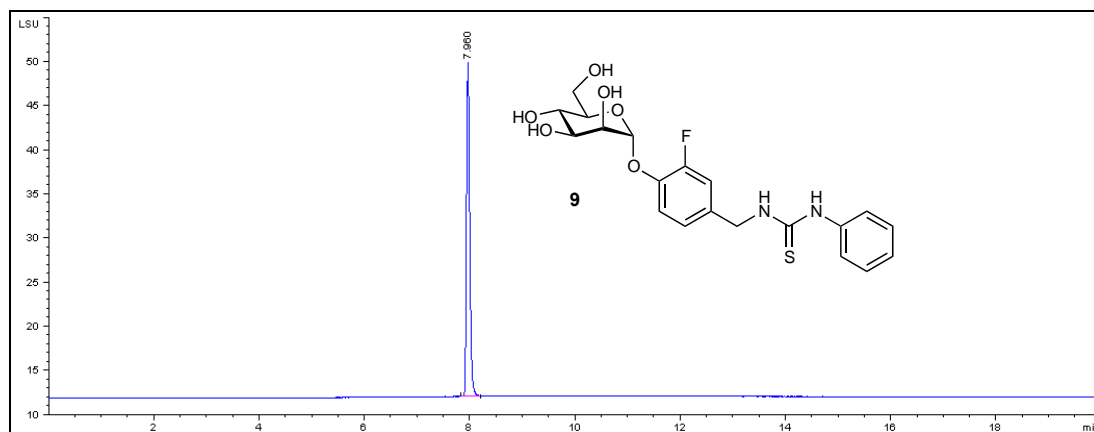
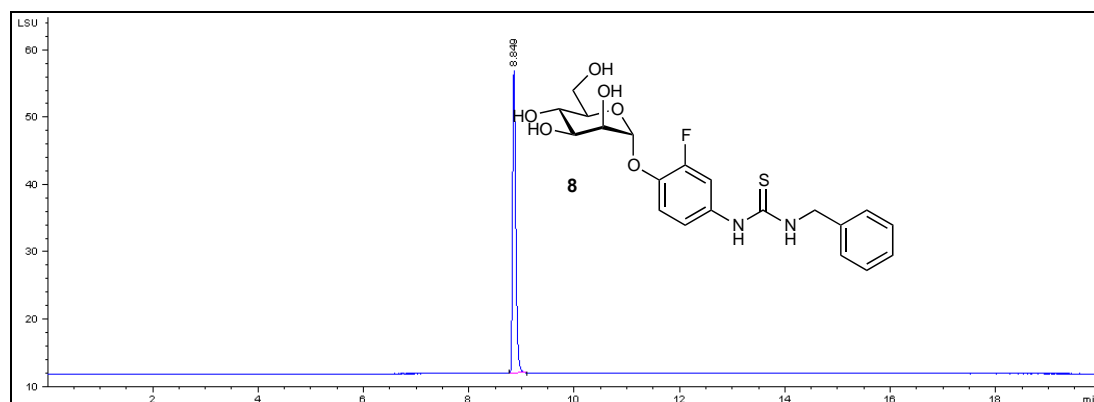
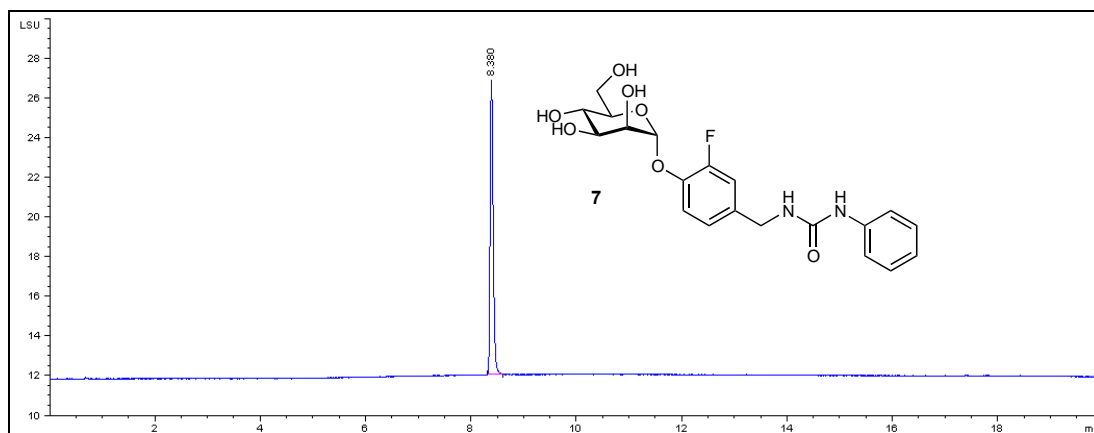
HPLC Traces of Target Compounds

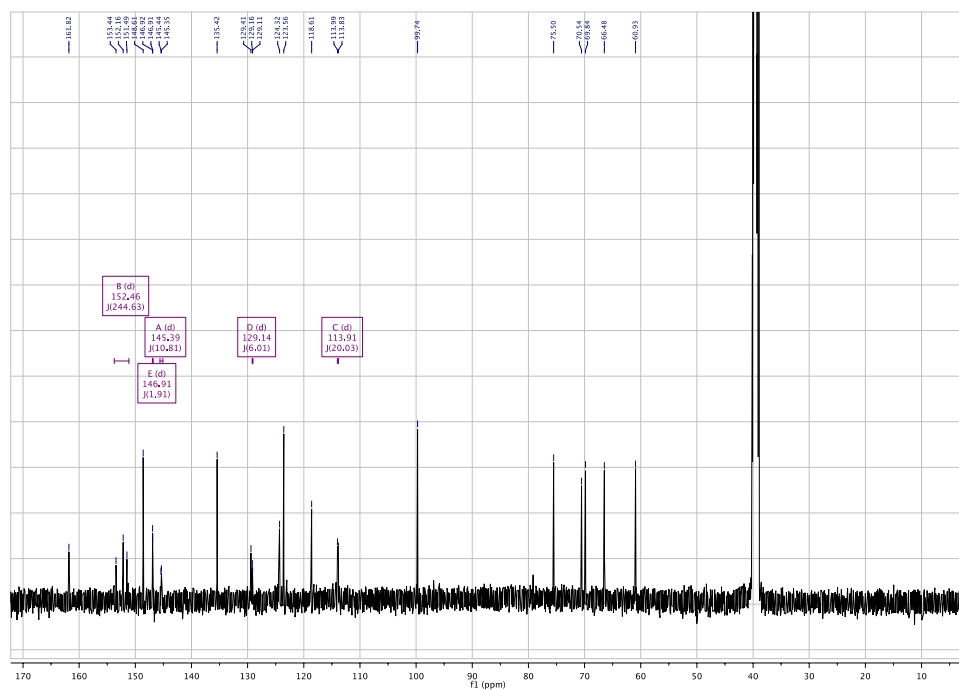
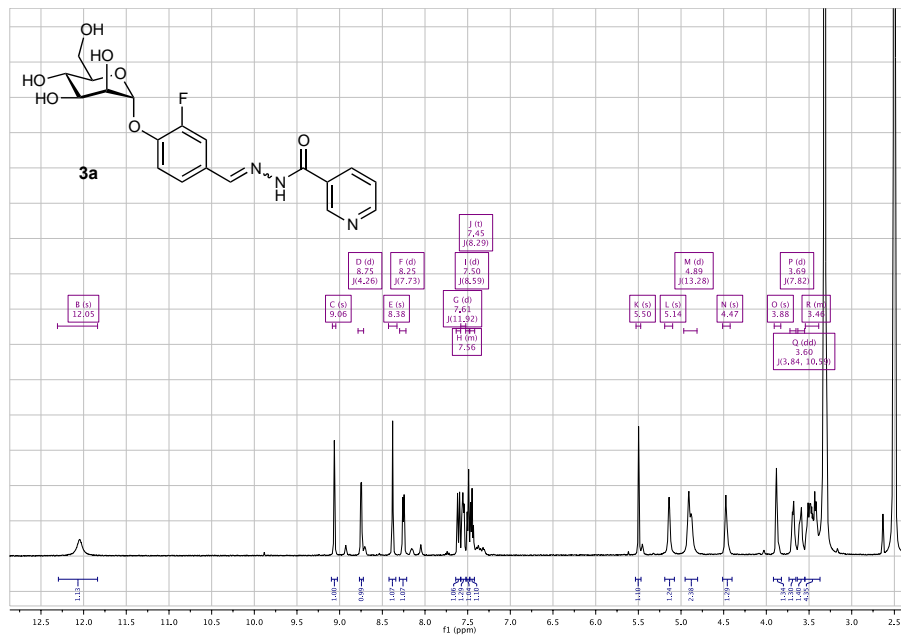


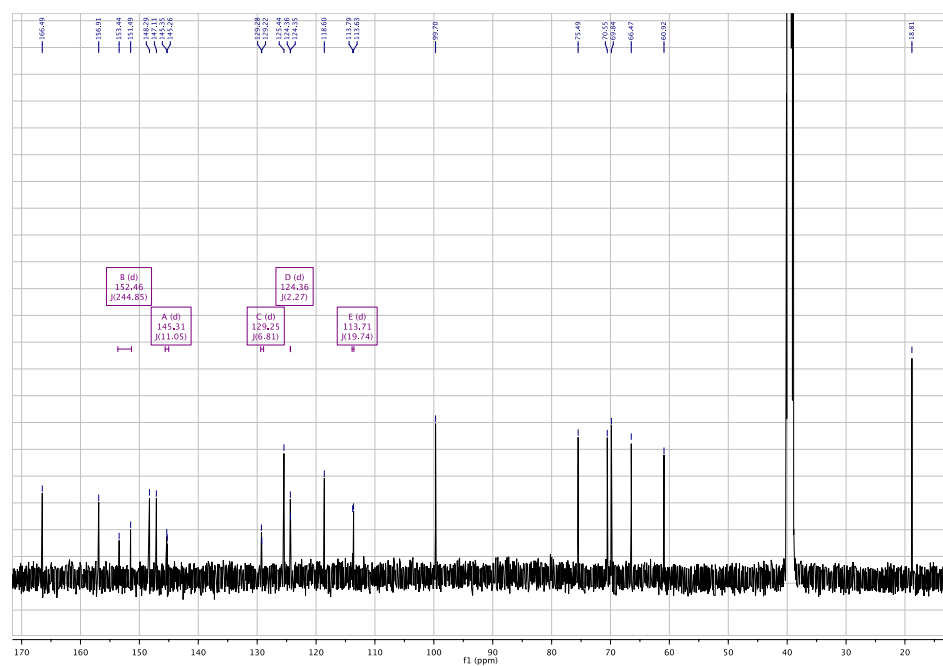


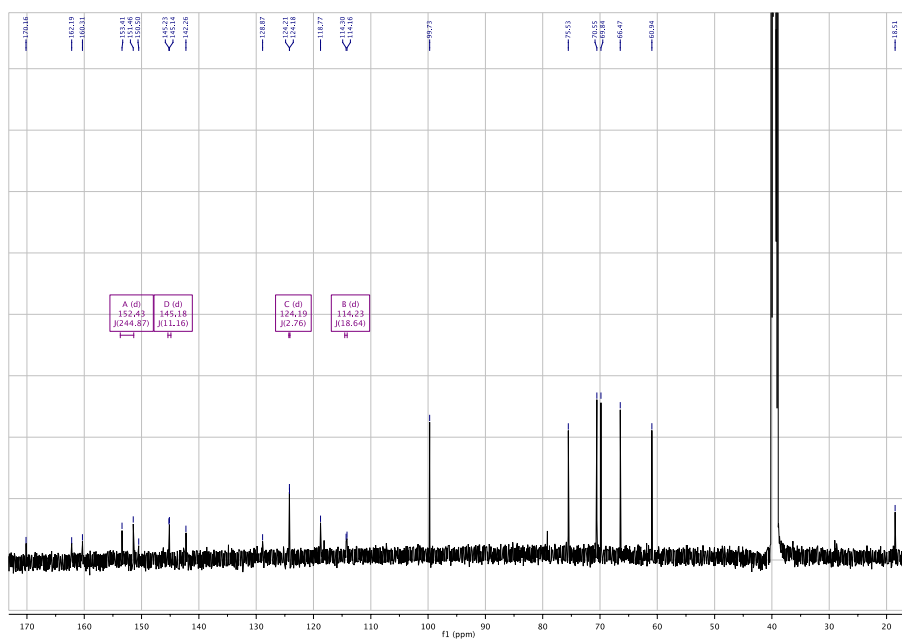
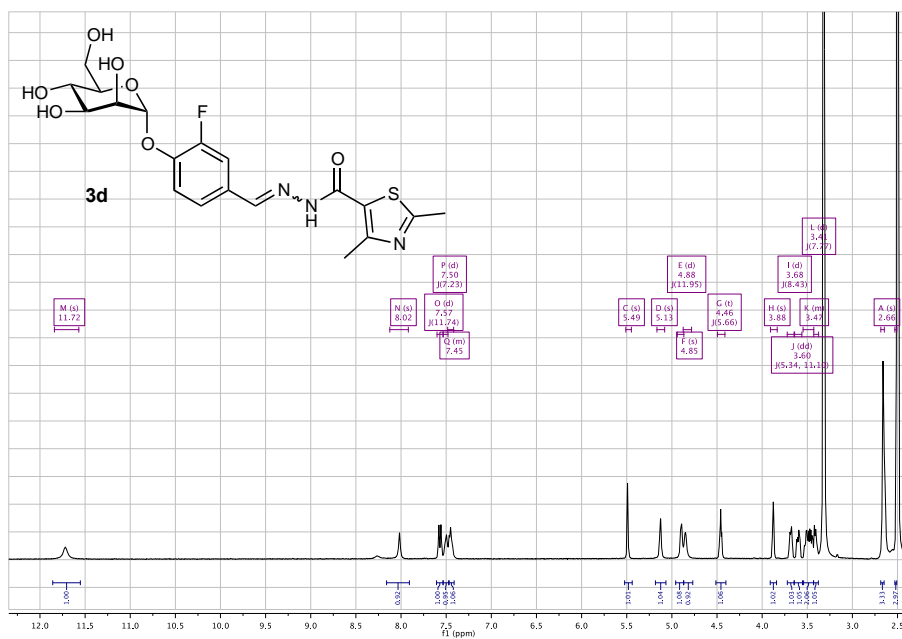


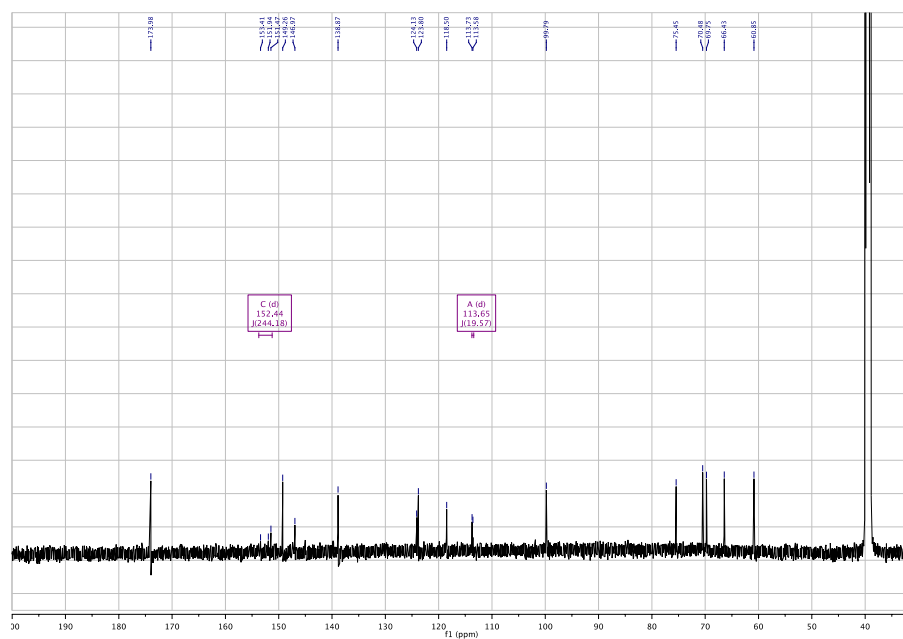
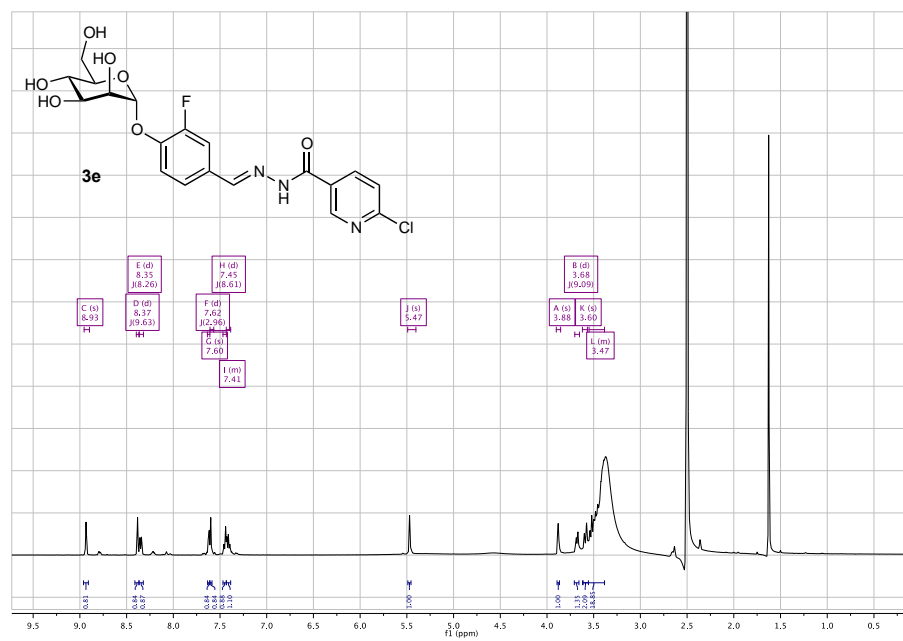


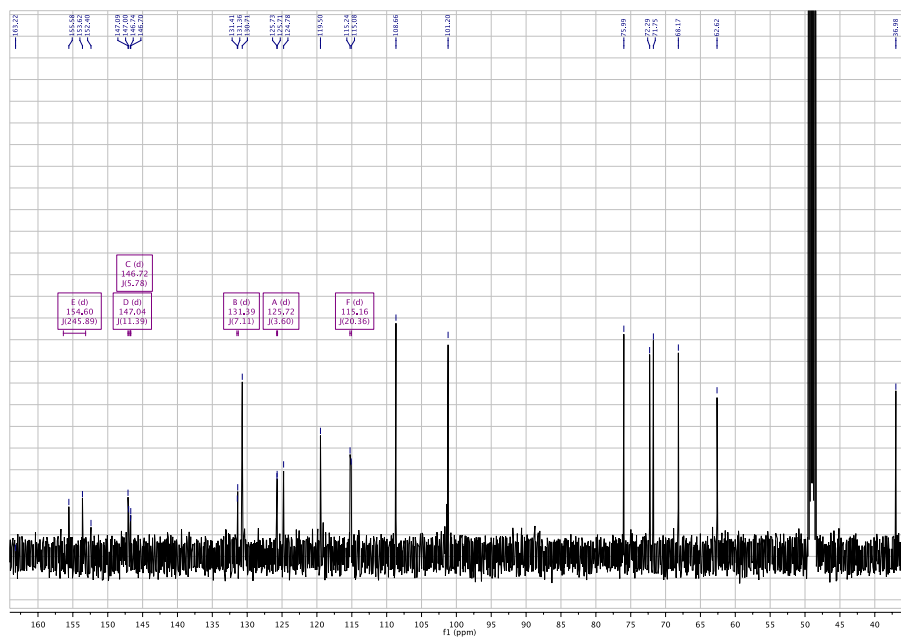
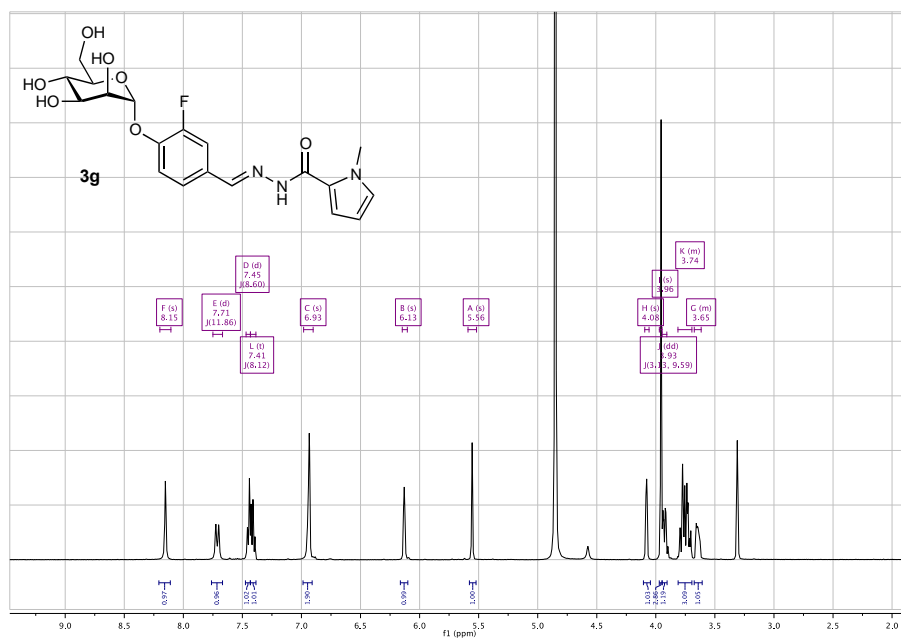


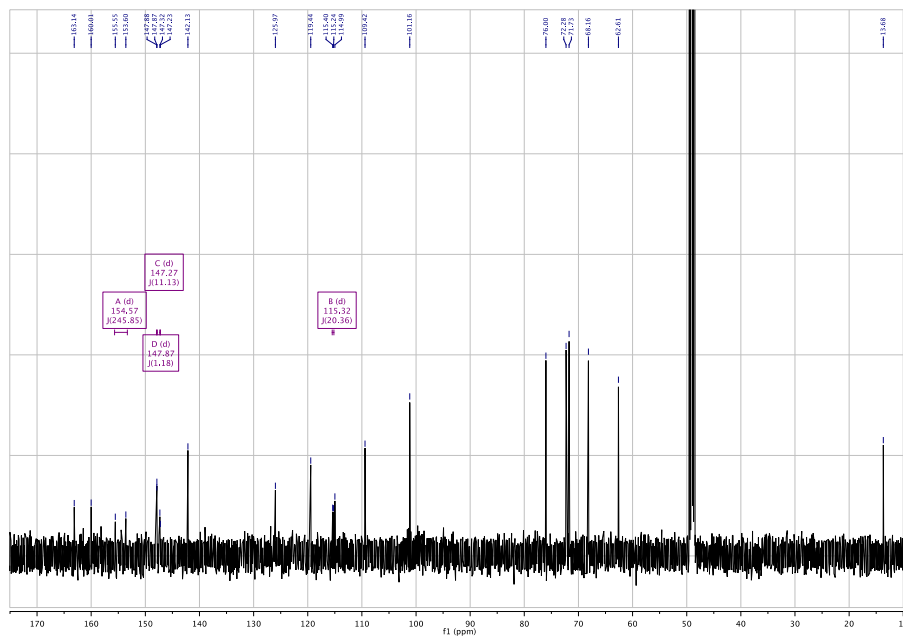
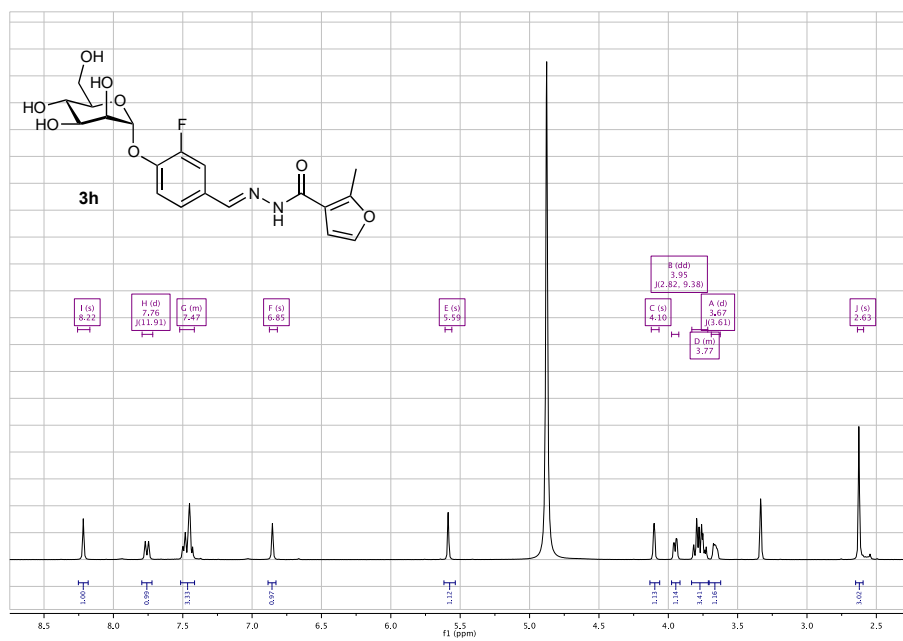
¹H & ¹³C NMR Spectra of Target Compounds

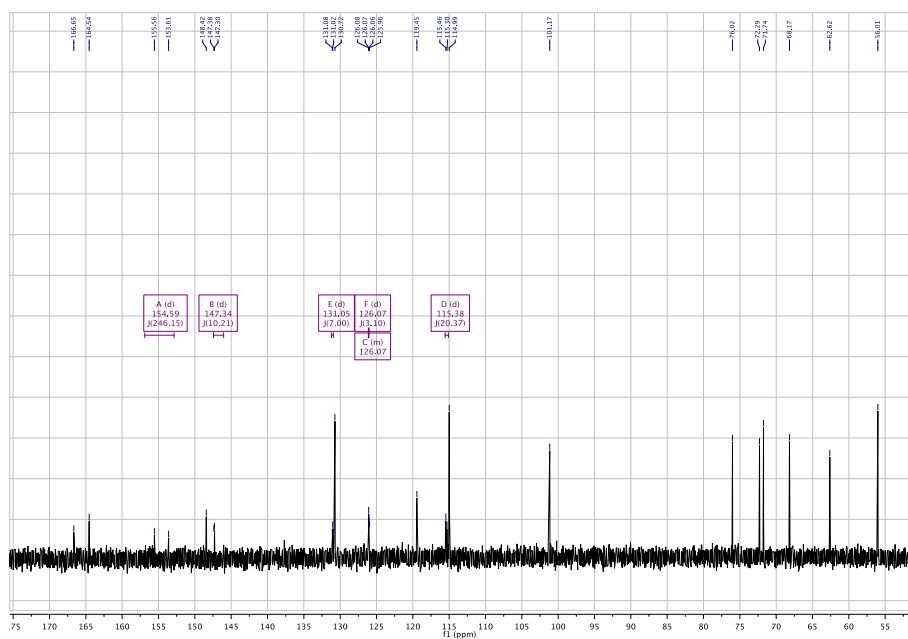
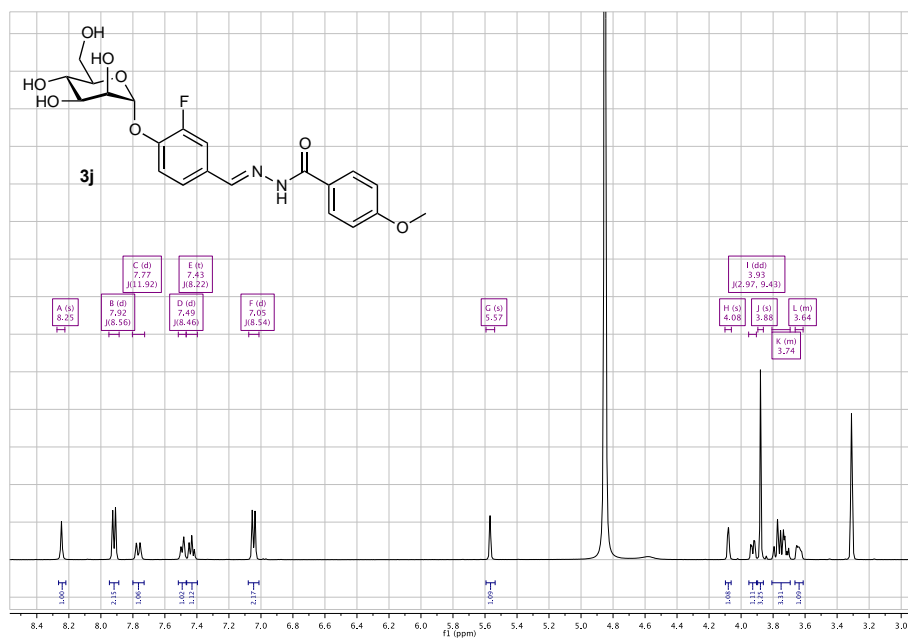


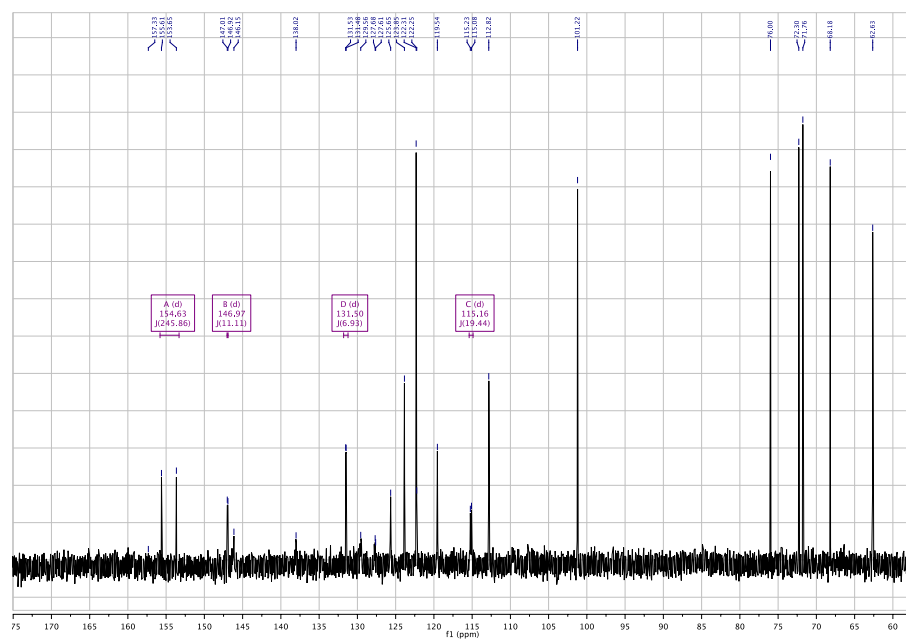
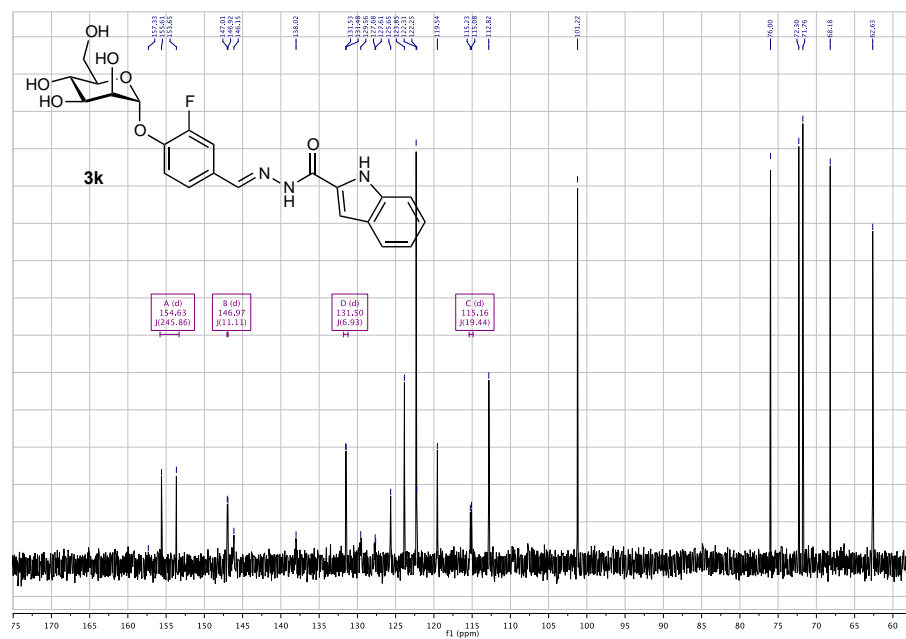


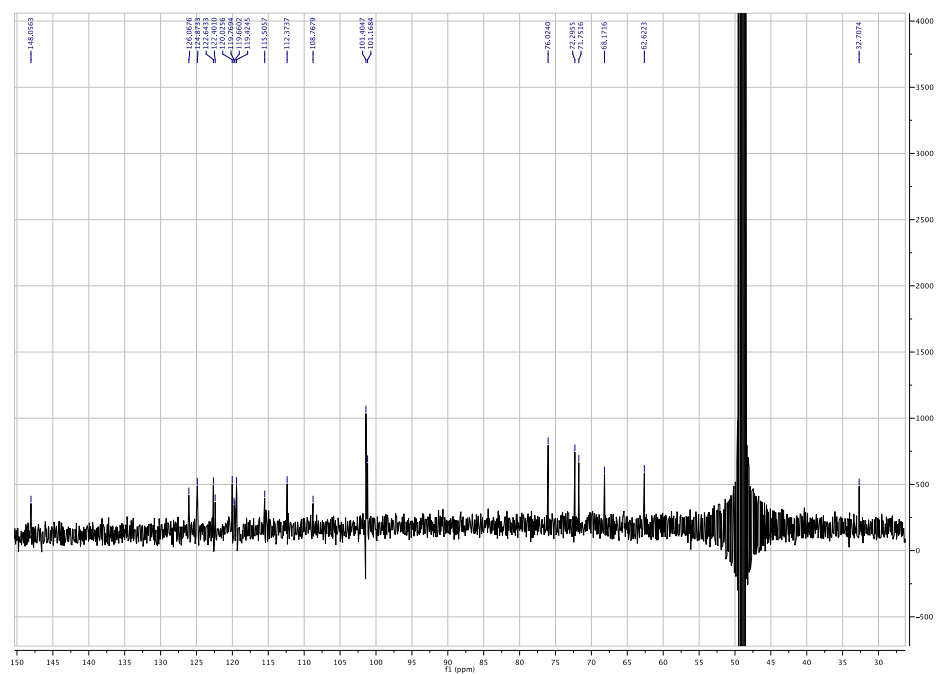
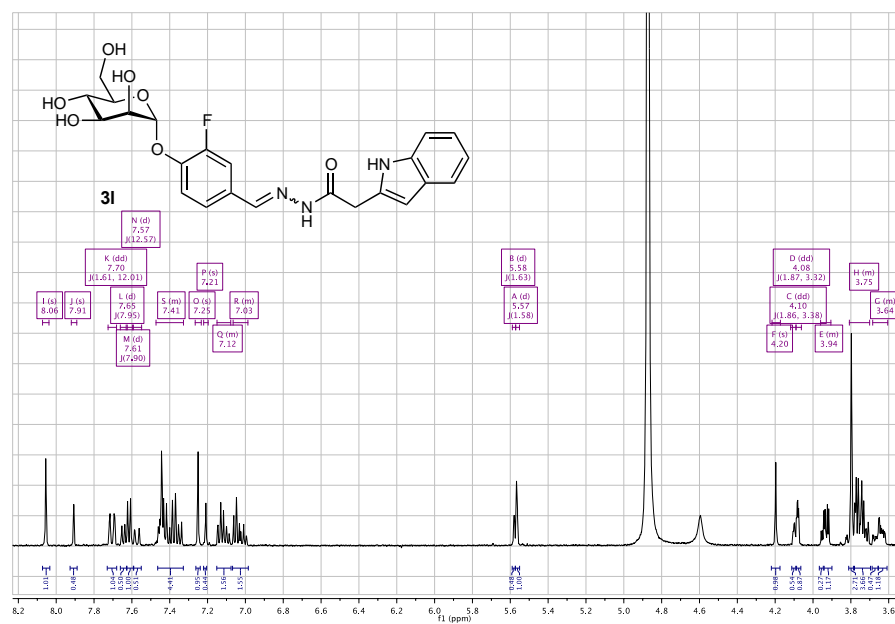


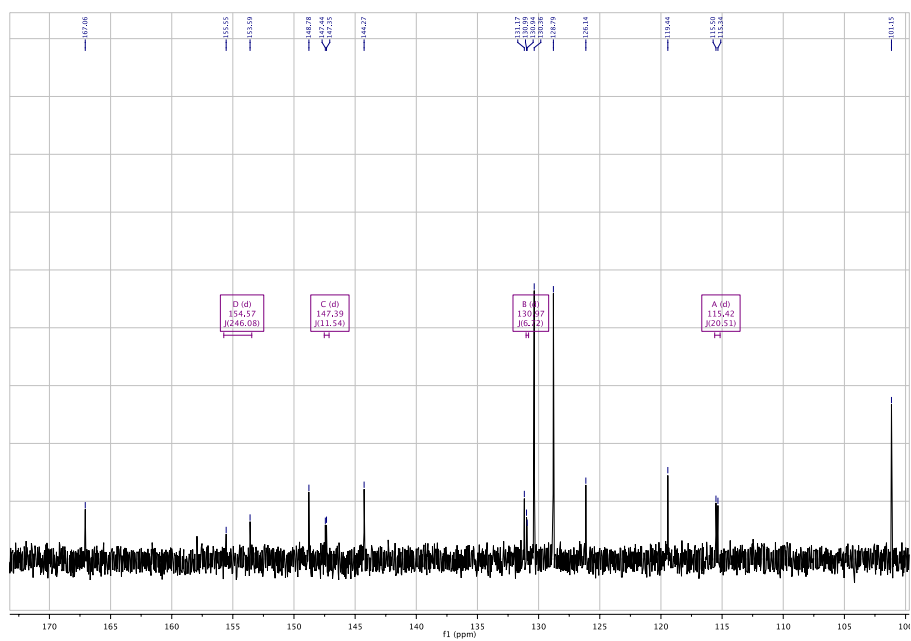
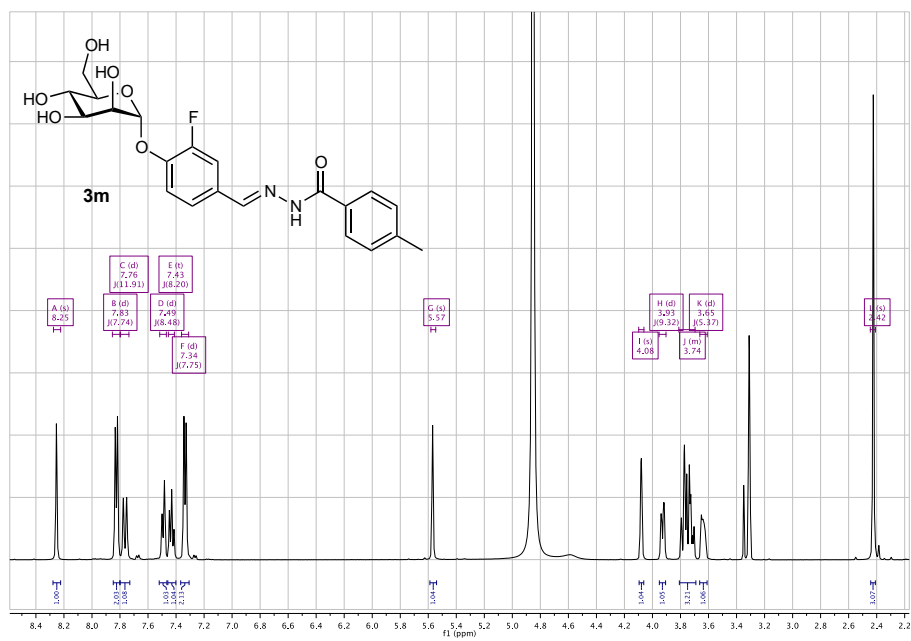


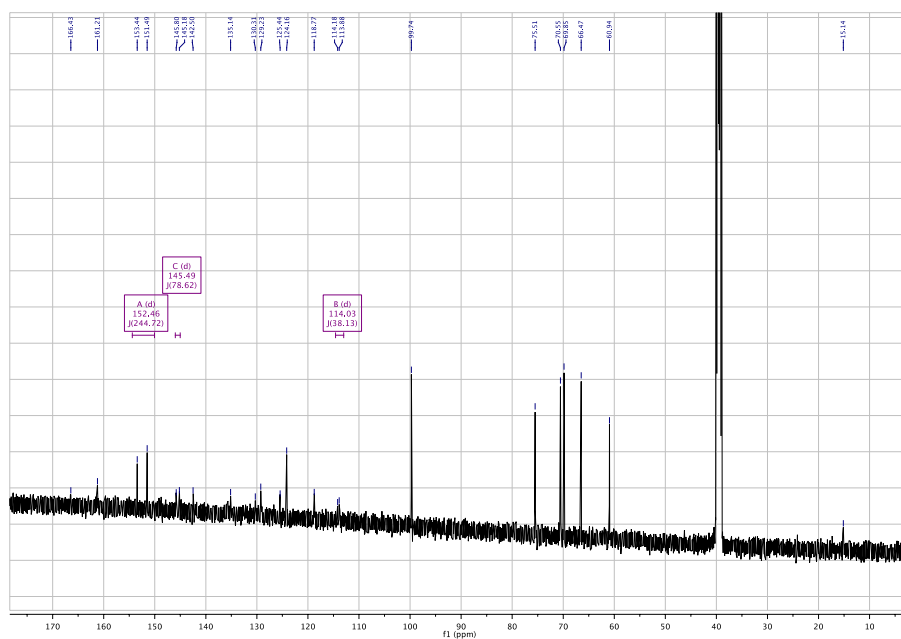
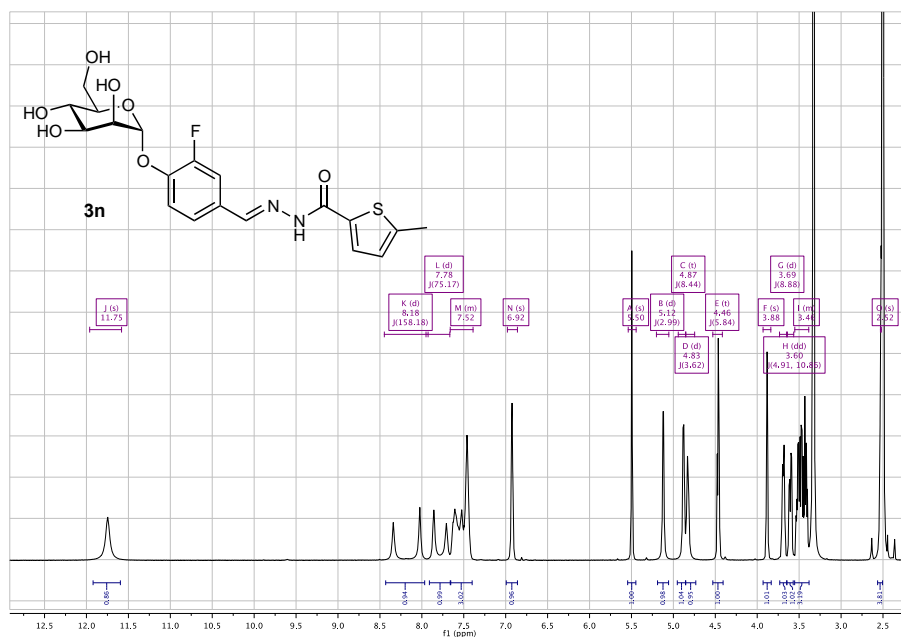


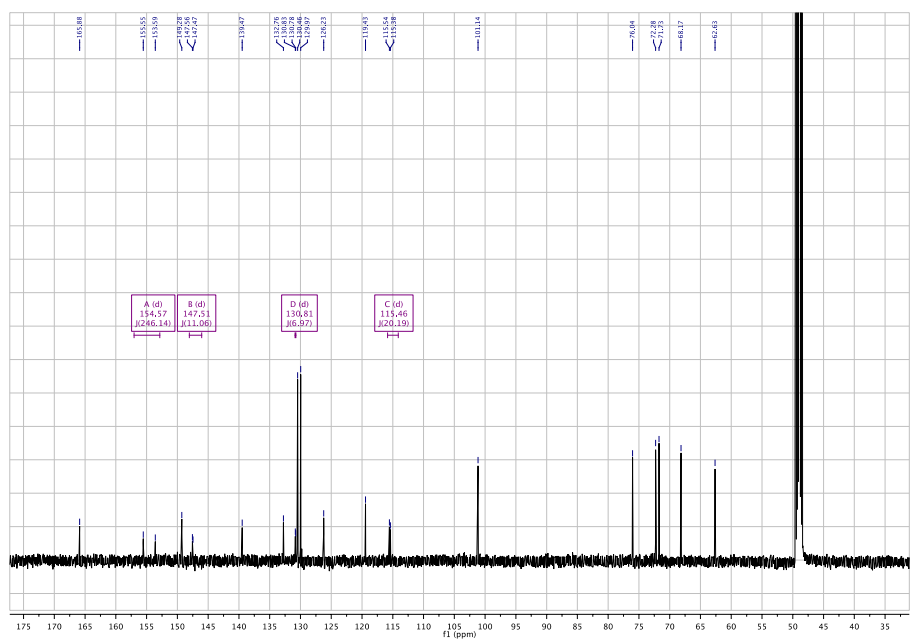
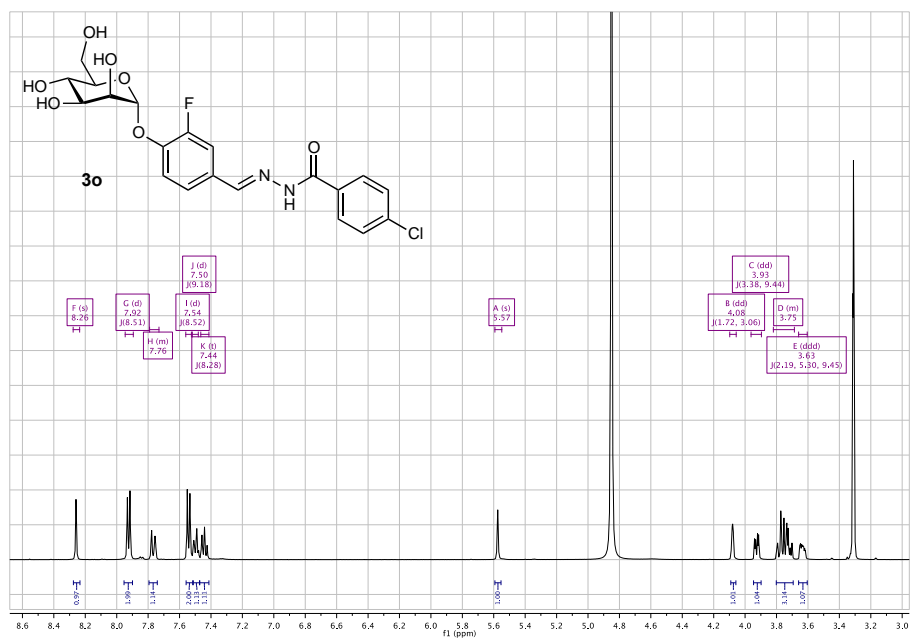


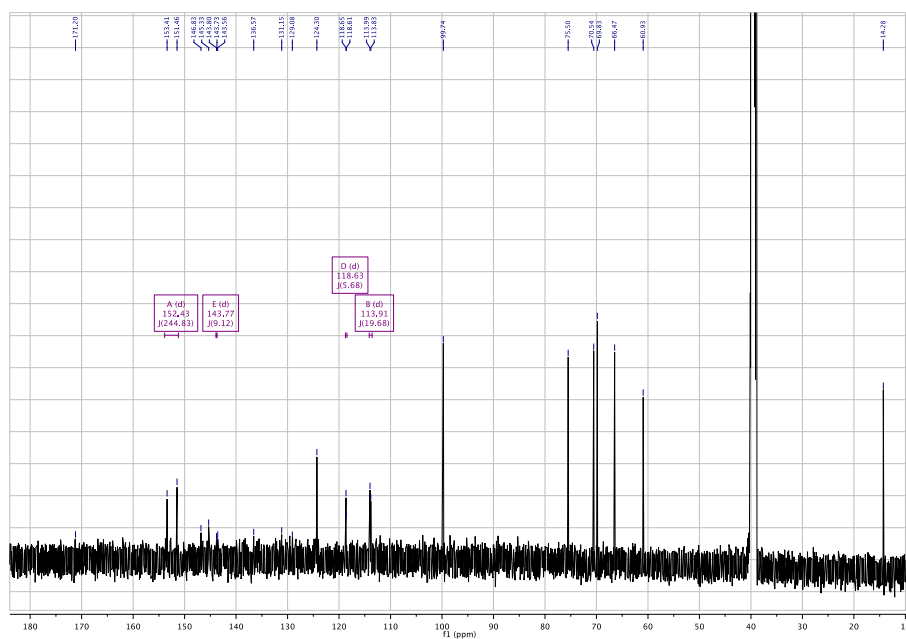
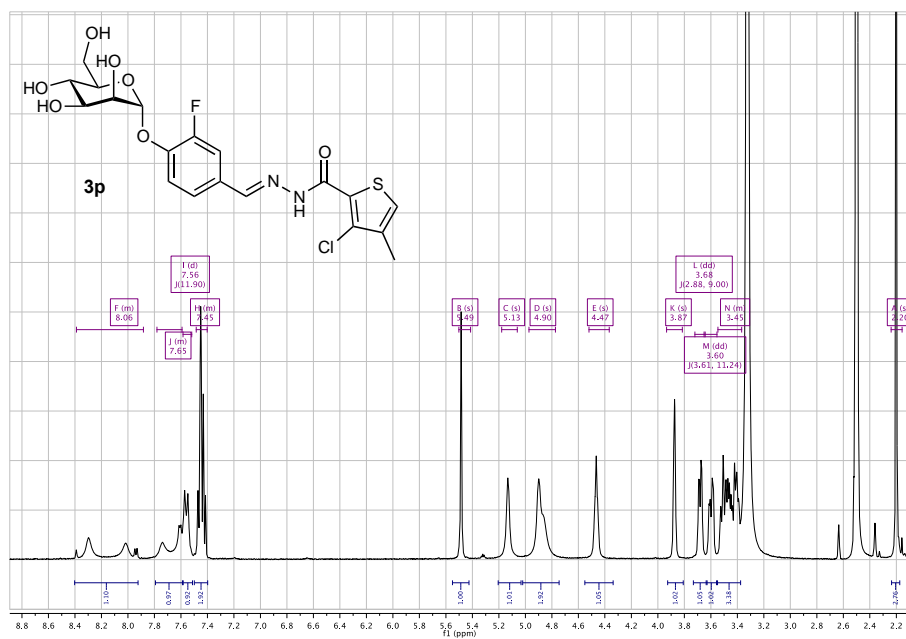


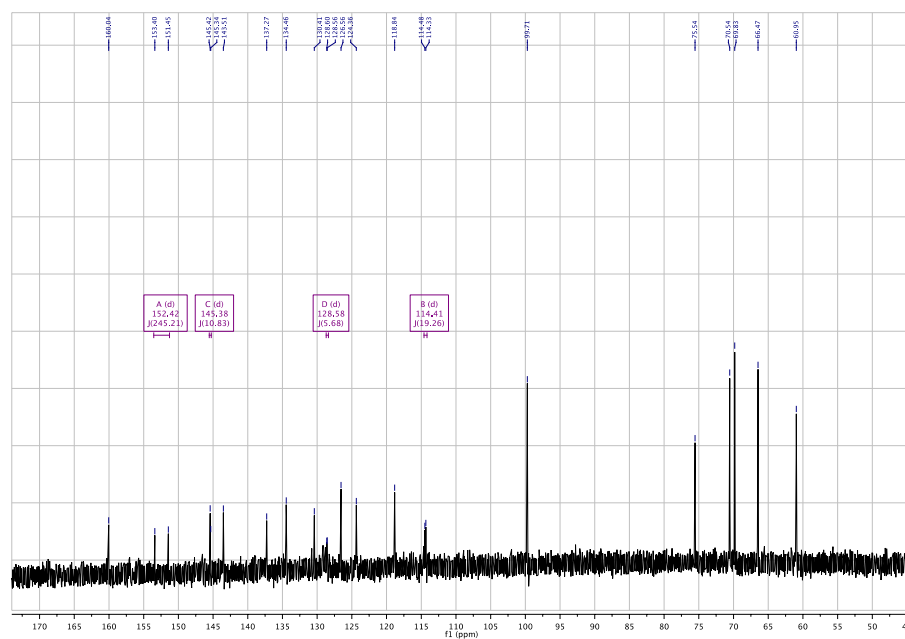
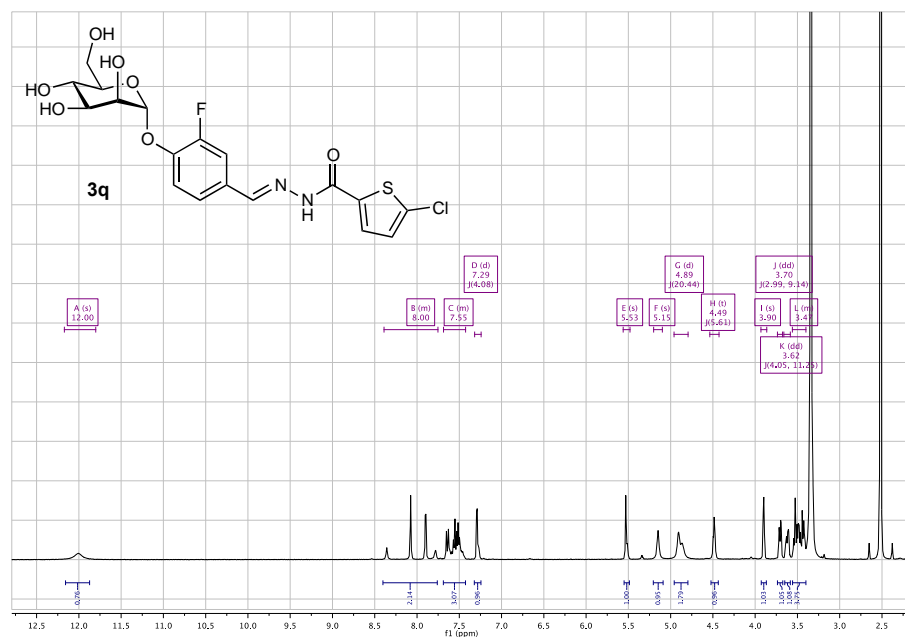


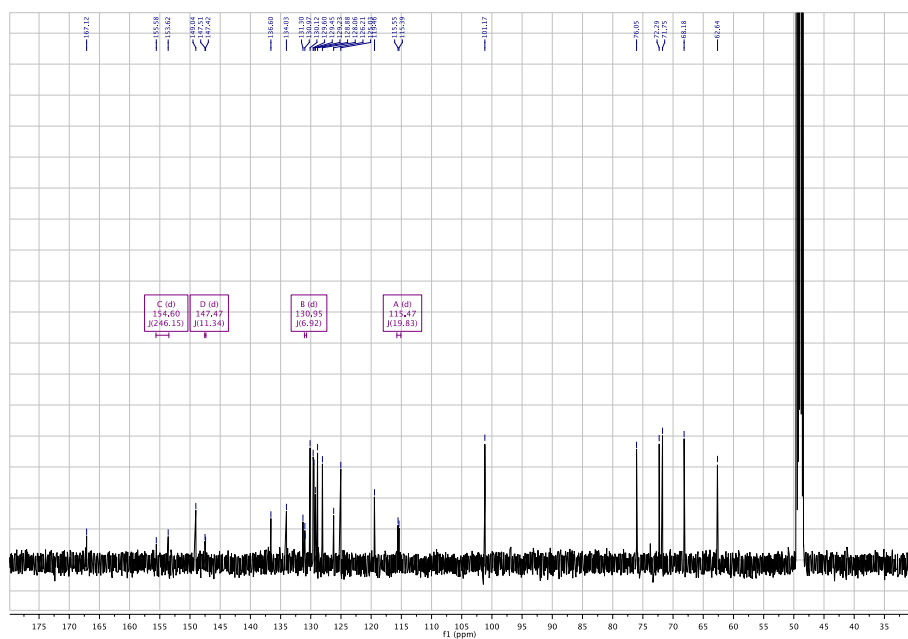
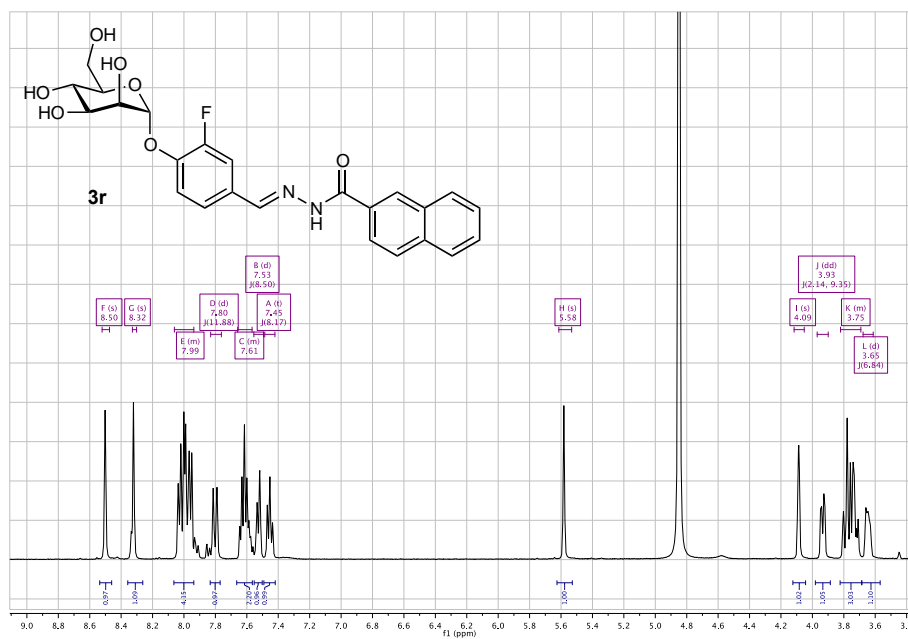


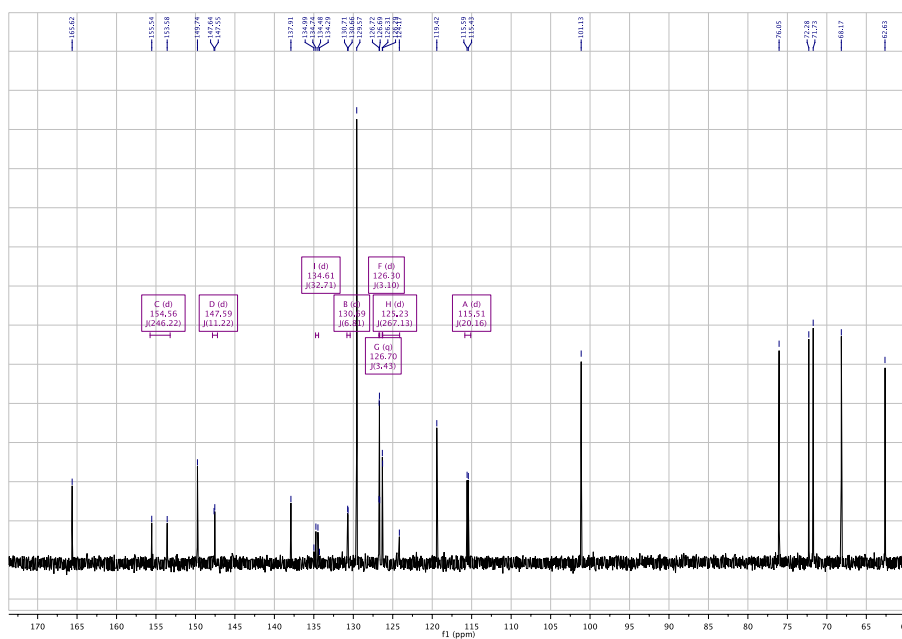
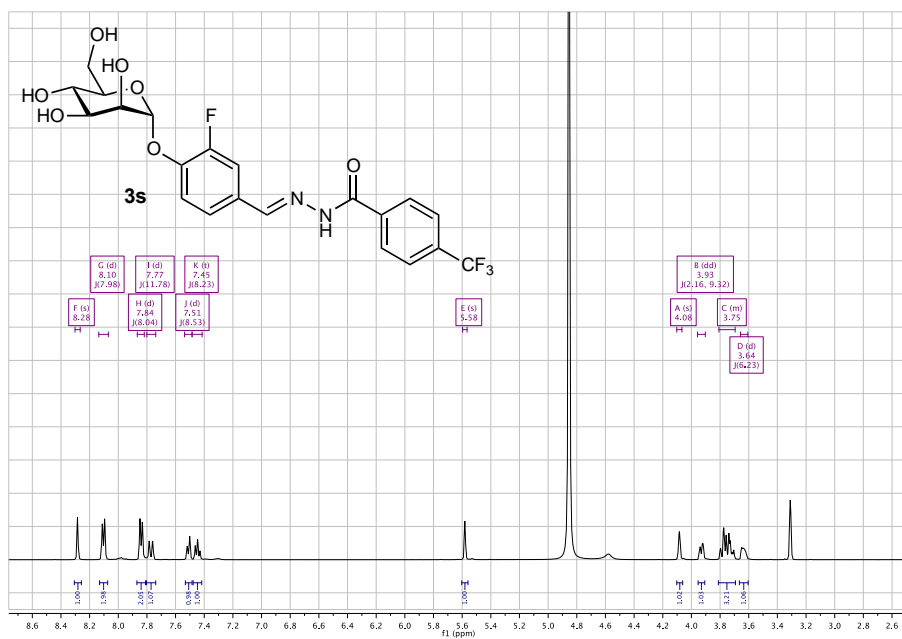


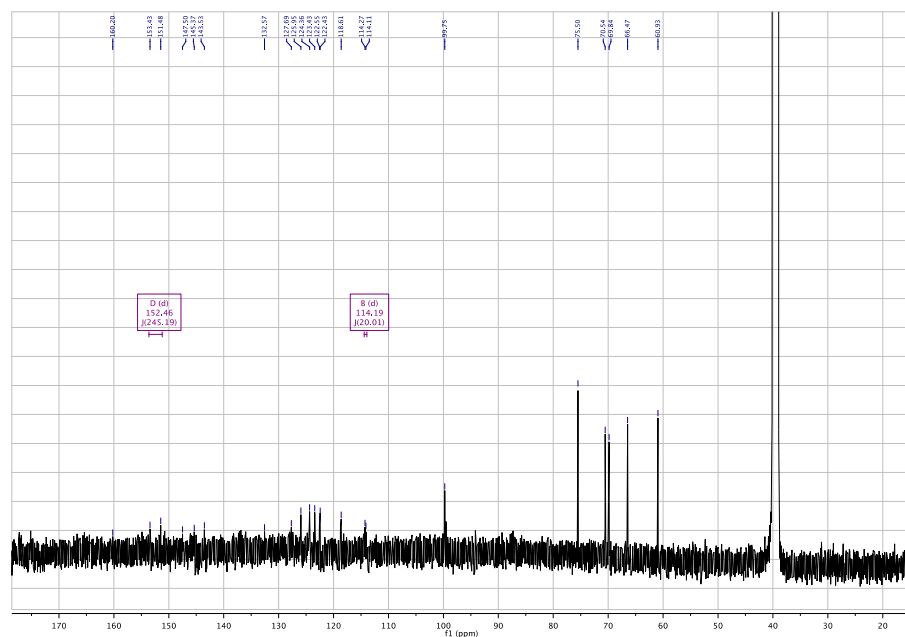
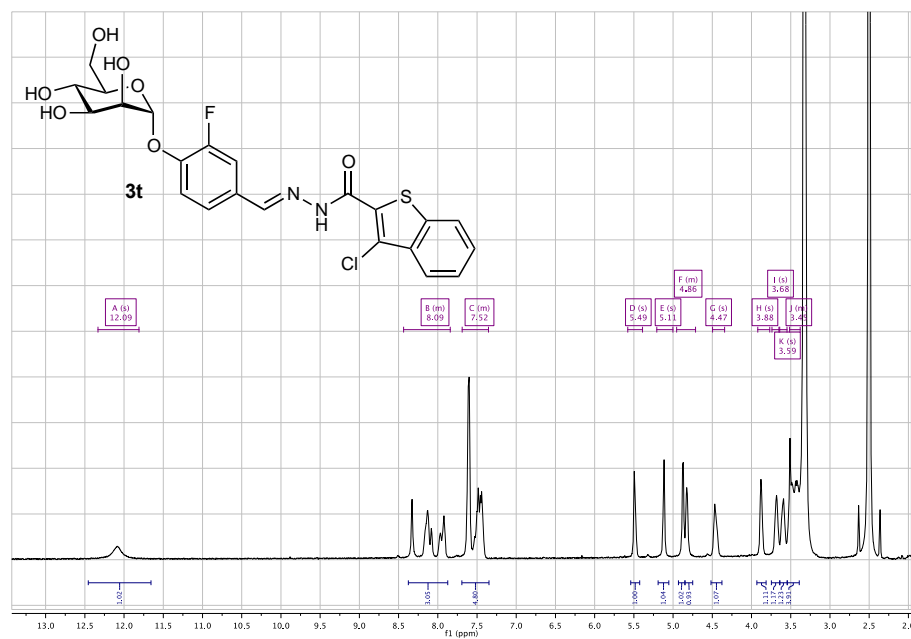


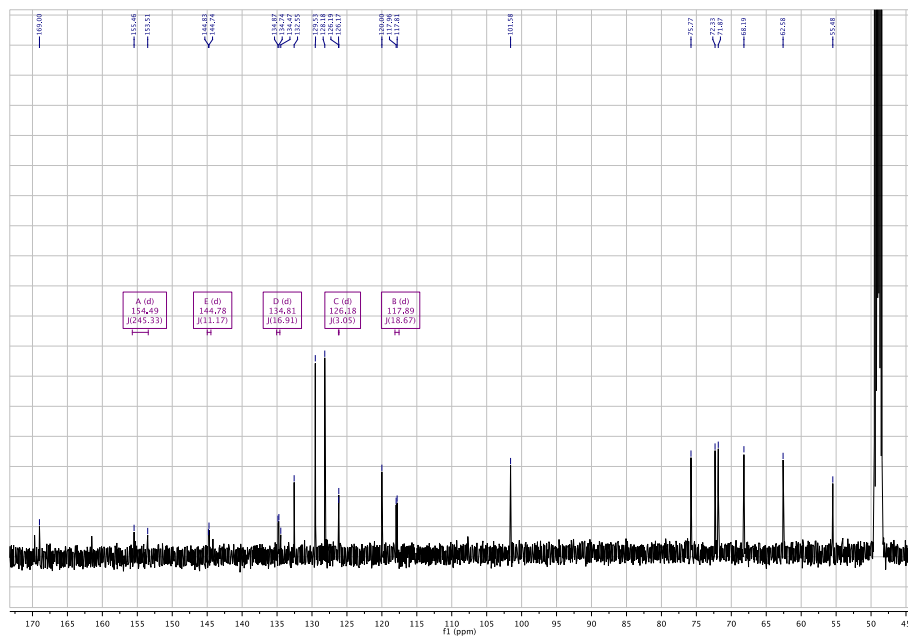
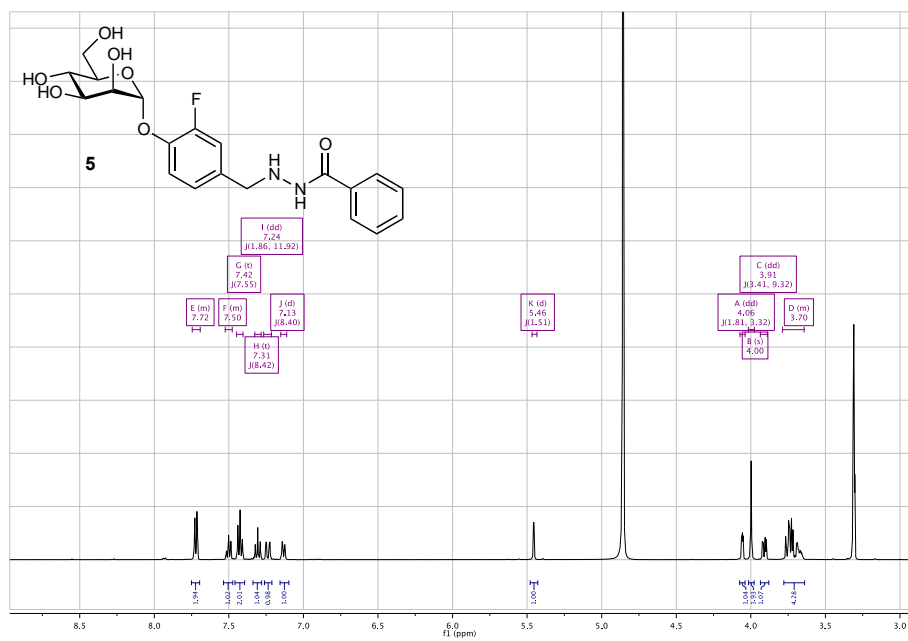


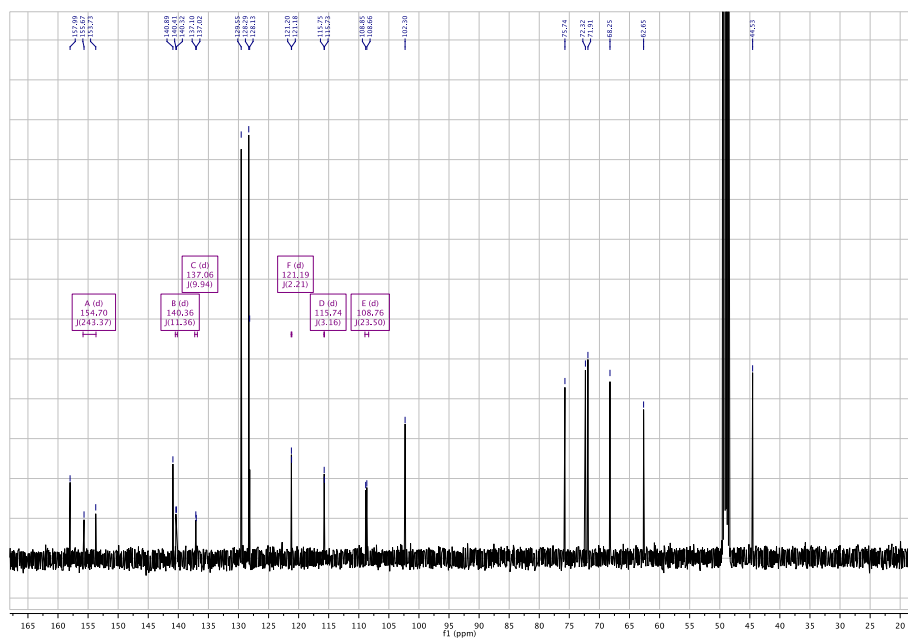
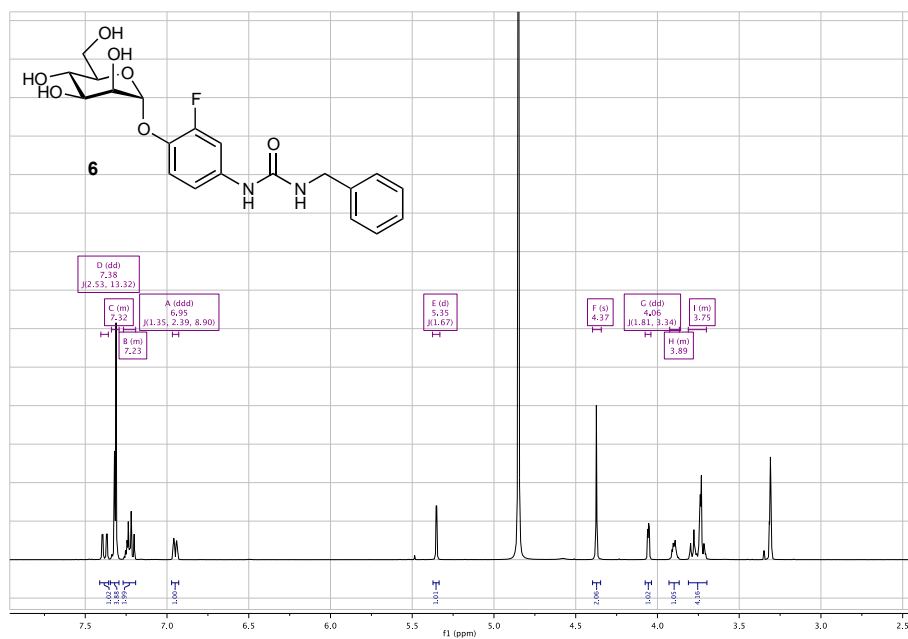


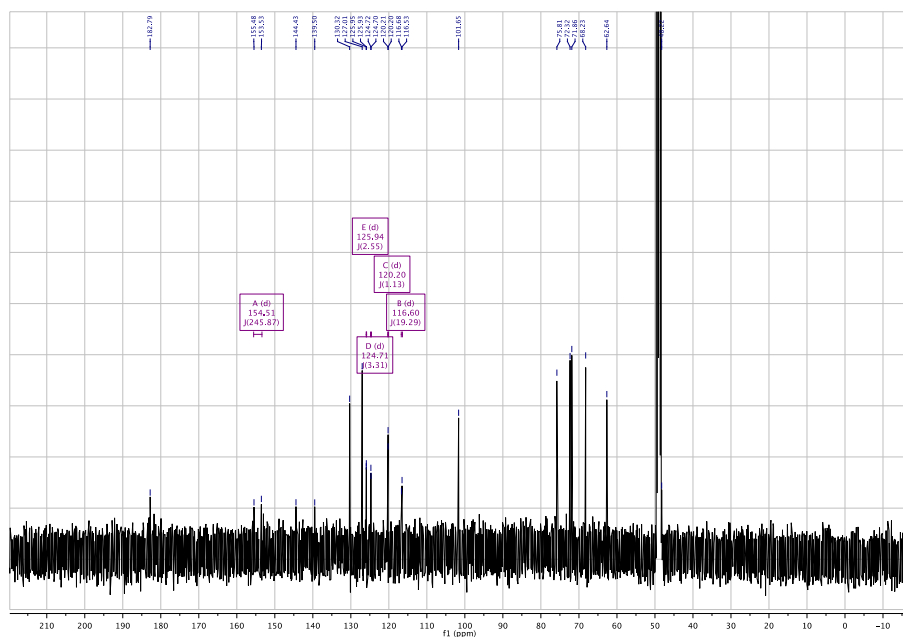
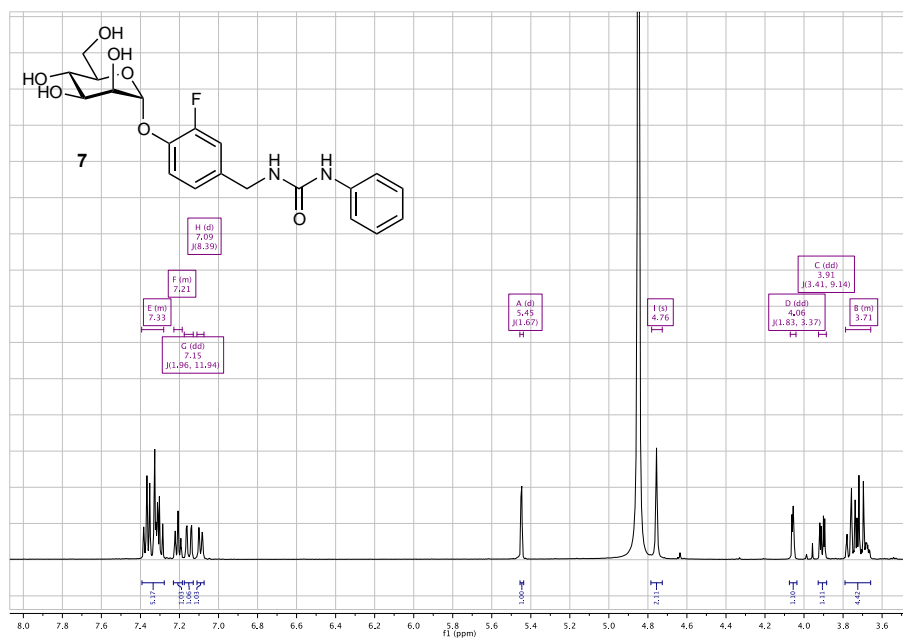


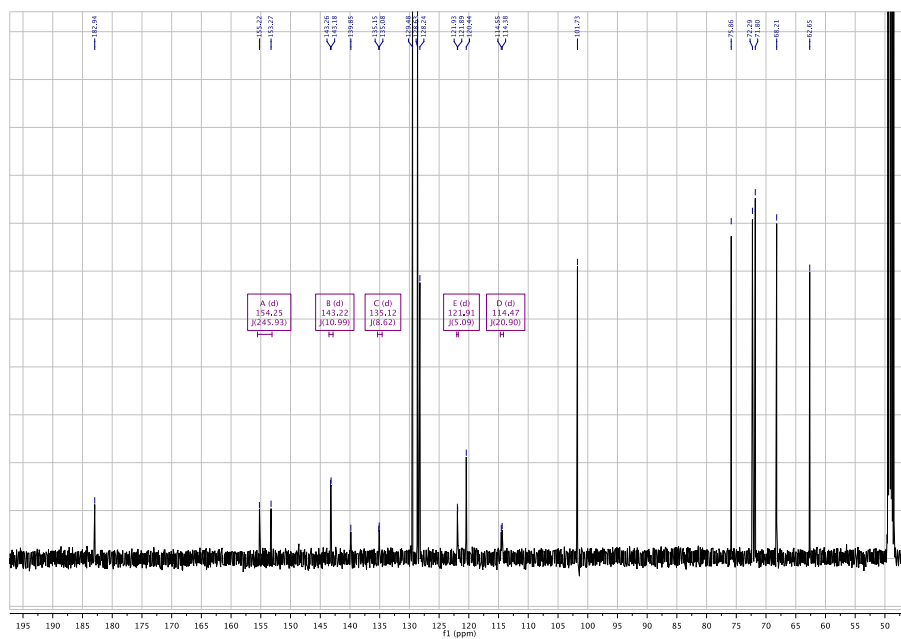
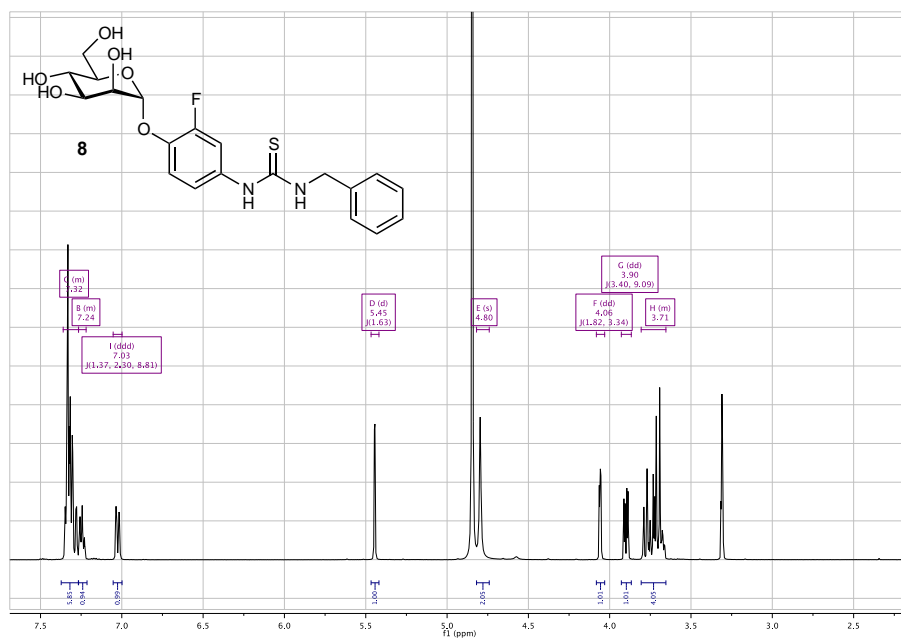


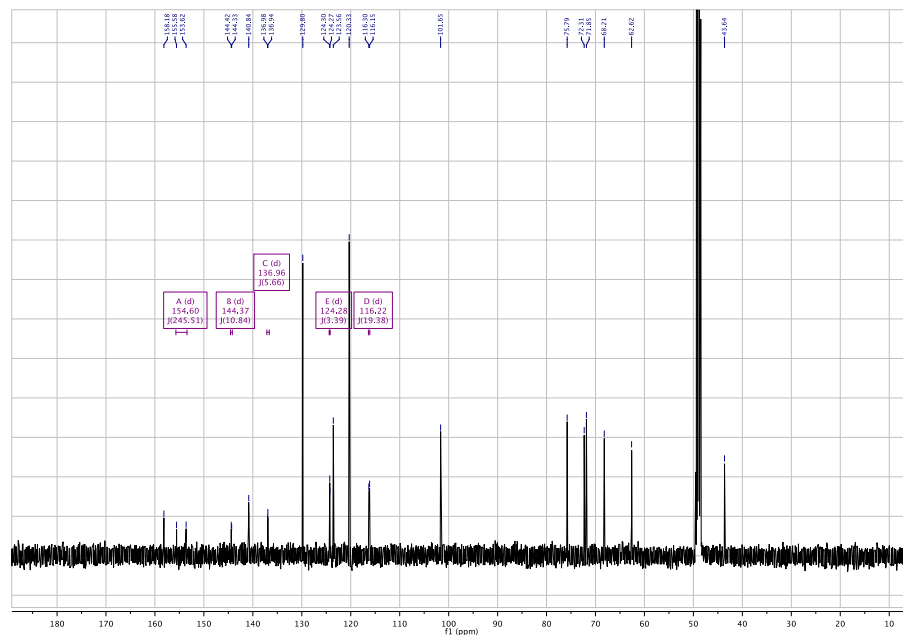
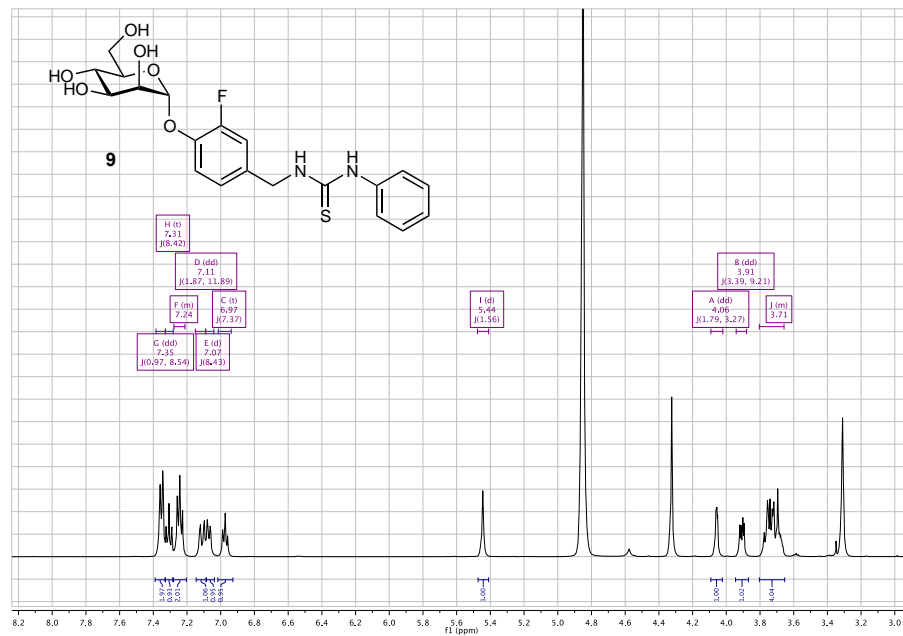


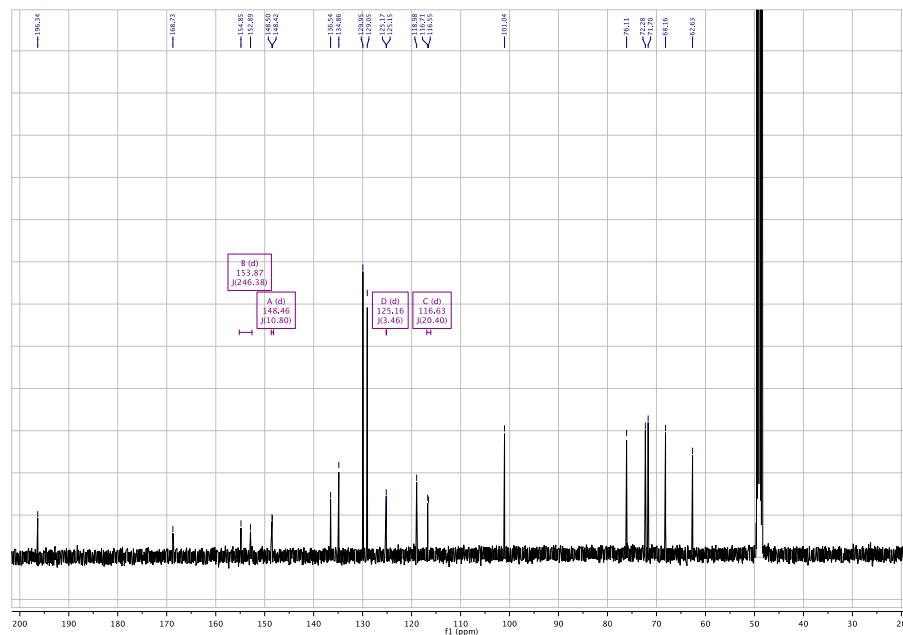
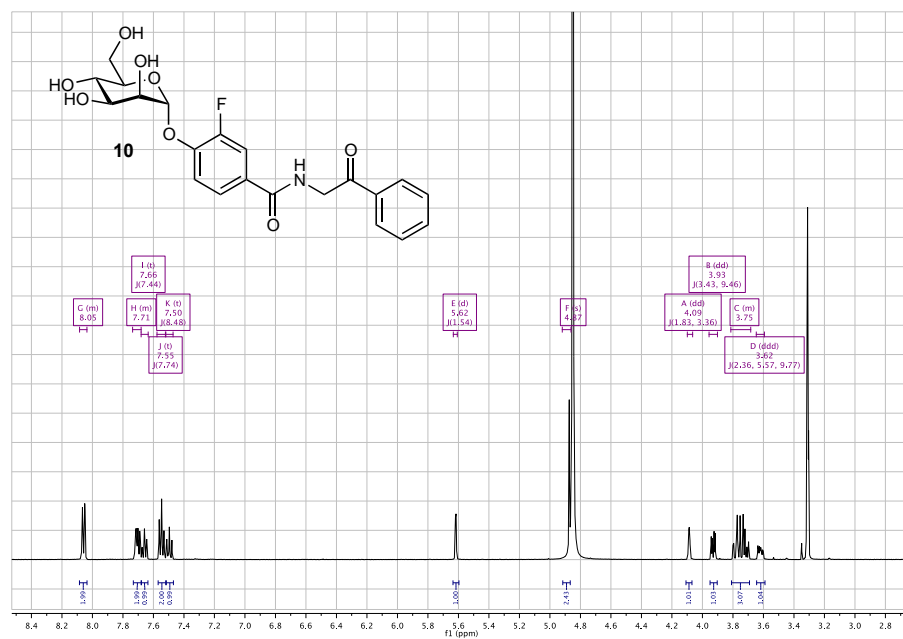












References

1. Frei, P.; Pang, L.; Silbermann, M.; Eriş, D.; Mühlethaler, T.; Schwarzt, O.; Ernst, B., Target-directed Dynamic Combinatorial Chemistry: A Study on Potentials and Pitfalls as Exemplified on a Bacterial Target. *Chem. Eur. J.* **2017**, *23*, 11570-11577.
2. Kleeb, S.; Pang, L.; Mayer, K.; Eris, D.; Sigl, A.; Preston, R. C.; Zihlmann, P.; Sharpe, T.; Jakob, R. P.; Abgottspon, D.; Hutter, A. S.; Scharenberg, M.; Jiang, X.; Navarra, G.; Rabbani, S.; Smiesko, M.; Lüdin, N.; Bezençon, J.; Schwarzt, O.; Maier, T.; Ernst, B., FimH Antagonists: Bioisosteres To Improve the in Vitro and in Vivo PK/PD Profile. *J. Med. Chem.* **2015**, *58*, 2221-2239.
3. Mayer, K.; Eris, D.; Schwarzt, O.; Sager, C. P.; Rabbani, S.; Kleeb, S.; Ernst, B., Urinary Tract Infection — Which Conformation of the Bacterial Lectin FimH Is Therapeutically Relevant? *J. Med. Chem.* **2017**, *60*, 5646-5662.
4. Wang, Z.-X., An exact mathematical expression for describing competitive binding of two different ligands to a protein molecule. *FEBS Lett.* **1995**, *360*, 111-114.