

Preparation of 2,2-dimethylchroman-4-ones from 5-alkyl-substituted resorcinols: microwave-assisted synthesis and theoretical calculations

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Dedicated to Professor Rosa María Claramunt Vallespi on the occasion of her 65th birthday

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Abstract

The influence of different 5-alkyl-substituted resorcinols on the formation of 2,2-dimethylchroman-4-ones is examined experimentally and theoretically. Structures are fully assigned by means of experimental and theoretical ¹³C and ¹H NMR chemical shifts. Based on experimental and theoretical calculations of Friedel-Crafts acylation, it is possible to explain the formation of 2,2-dimethyl-5-hydroxychroman-4-ones and/or 2,2-dimethyl-7-hydroxychroman-4-ones. Evaluation of their biological activity as cannabinoid receptor ligands is also reported.

Keywords: Chromanone, DFT calculation, Friedel-Crafts reactions, cannabinoids

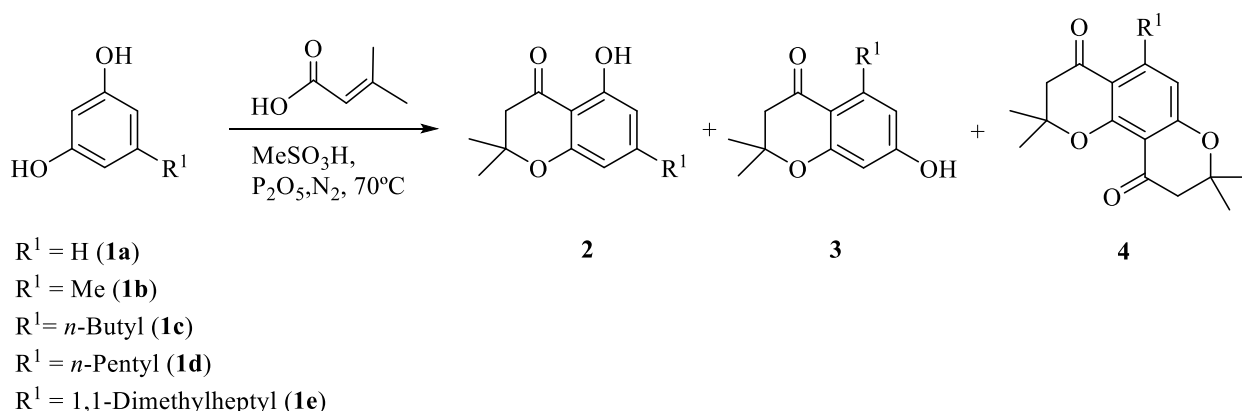
Introduction

Chroman-4-ones are attractive compounds not only because of their therapeutic applications but also as intermediates in preparation of more complex structures. As reported by Saengchantara and Wallace¹ in a review covering chromanols, chromanones and chromones, many chroman-4-ones are isolated from natural sources such as *Dysophylla stellata*, rhizomes of rhubarb, or *Penicillium frequentans*. Examples of these compounds include cannabichromanones identified as components of the *Cannabis sativa* plant.^{2,3} Moreover, 2,2-dimethyl-5-hydroxychroman-4-ones have been described as intermediates in the synthesis of tetrahydrocannabinols⁴ and, as we have reported, in the synthesis of cannabinoid chromenopyrazoles.⁵ A survey of the preparations of differently substituted 2,2-dimethylchroman-4-ones was published by Tímár *et al.*⁶ twelve years ago. Most of them are prepared by reacting the appropriate phenol with 3,3-dimethyl-

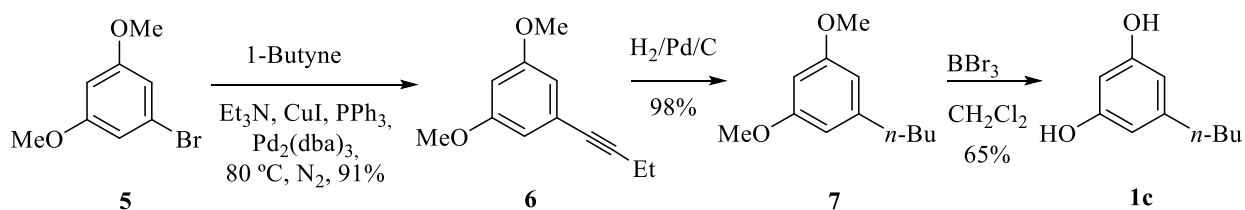
acryloyl chloride or its corresponding acid in presence of Lewis acid. However, it has been mentioned in the literature that the reactions are not always successful, providing poor yields or mixtures of different chromanones, or requiring long reaction times.^{7,8} On consideration of this, a study was undertaken to examine the influence of different 5-alkyl-substituted resorcinols on the formation of 2,2-dimethylchroman-4-ones using both microwave (MW) assisted reactions and conventional heating (C.H.). Structures have been fully assigned by means of experimental and theoretical ¹³C and ¹H NMR chemical shifts. Finally, the first step, Friedel-Crafts acylation reaction has been examined theoretically in order to correlate the experimental evidence using Density Functional Theory (DFT).

Results and Discussion

2,2-Dimethyl-5-hydroxychroman-4-ones (**2**) and 2,2-dimethyl-7-hydroxychroman-4-ones (**3**) were prepared from various resorcinols as indicated in Scheme 1. Resorcinol (**1a**), orcinol (**1b**) and olivetol (**1d**) are commercially available. The starting resorcinol **1c** was prepared starting from 1-bromo-3,5-dimethoxybenzene. Sonogashira coupling⁹ of the starting material gave 5-(1-butynyl)-1,3-dimethoxybenzene in good yield (Scheme 2). Then, 5-butyl-1,3-dimethoxybenzene was obtained in quantitative yield by hydrogenation using Pd/C as catalyst. Demethylation with boron tribromide finally provided the desired resorcinol **1c** in acceptable yield. Resorcinol **1e** bearing a dimethylheptyl substituent was prepared from the corresponding dimethoxy derivative.



Scheme 1. Reaction of resorcinols with 3,3-dimethylacrylic acid.



Scheme 2. Preparation of 5-*n*-butyl-1,3-dihydroxybenzene (**1c**).

Upon treatment of resorcinols (**1a-1e**) with 3,3-dimethylacrylic acid in methanesulfonic acid in presence of phosphorus pentoxide, the Friedel-Crafts acylation was expected to take place at either the 2 or 6 position of 1,3-dihydroxybenzene (Scheme 1). To study the effect of the alkyl substituents on the resorcinol, reaction parameters (solvent, molar ratio of reactants, catalyst, and temperature) were at first investigated. The reactions were carried out under the most advantageous reactant ratio: resorcinol / phosphorus pentoxide / 3,3-dimethylacrylic acid [1/1.2-1.5/1.5-2] using methanesulfonic acid as solvent at 70 °C. Under these conditions the reaction was performed under microwave irradiation for 10, 20 or 60 minutes or with conventional heating for 12 hours. The reactions were monitored by HPLC/MS (C-18 reverse phase column, eluent: MeCN/H₂O) that allowed the ratio in which regioisomers **2** and **3** were formed to be determined.

As reported in Table 1, 2,2-dimethyl-7-hydroxychroman-4-one was isolated as the only regioisomer in the case of **3a** and **3b** (R¹ = H and R¹ = Me), whereas the regioisomer 2,2-dimethyl-5-hydroxychroman-4-one was isolated as the unique isomer for **2e** (R¹ = 1,1-dimethylheptyl). The reaction of 3,3-dimethylacrylic acid with 5-butyl-1,3-dihydroxybenzene (R¹ = *n*-butyl) and olivetol (R¹ = *n*-pentyl) resulted in the formation of both isomers **2** (**2c**, **2d**) and **3** (**3c**, **3d**). The ratio in the crude product mixture could be easily determined by HPLC/MS. These ratios are reported in Table 1. Regioisomers **2** and **3** could be isolated by column chromatography. Two-dimensional NMR experiments were of particular value for the identification of the regioisomers **2** and **3**, as shown in Figure 1. Moreover, the chemical shifts of

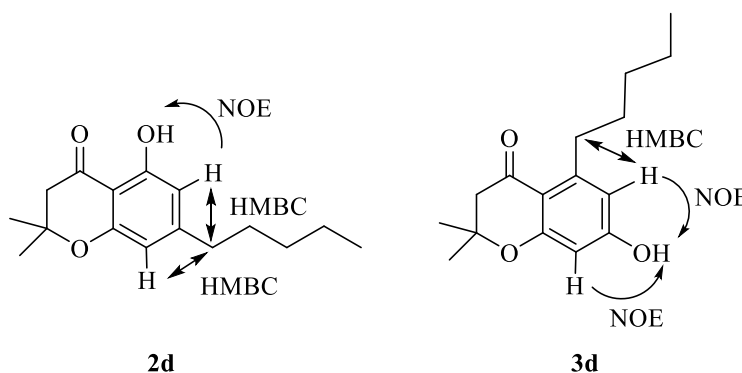
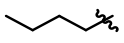
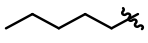
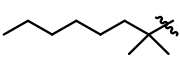


Figure 1. Specific assignments of regioisomers **2** and **3** were based on ¹³C-¹H Heteronuclear Multiple-Bond Correlation (HMBC) and Nuclear Overhauser Enhancement (NOE) spectroscopy.

All the molecules **2** and **3** obtained experimentally have also been calculated using GIAO-DFT methods at B3LYP/6-311++G(d,p) computational level. The ¹³C-NMR theoretical data nicely resemble the experimental ones, showing an excellent linear correlation with the experimental findings (R² = 0.993) and an absolute average error of 3.1 ppm. The ¹H-NMR theoretical results show an absolute average error of 0.18 ppm and a R² value of 0.99.

Table 1. Reaction of 5-alkyl-substituted resorcinols with 3,3-dimethylacrylic acid in methanesulfonic acid under given heating conditions and the ratios of the two isomers and the diacylated product determined by HPLC/MS areas analysis

Entry	R ¹	Heating condition ^a	2	3	4
1	H	10 min MW	-	100% (3a)	-
2		20 min MW	-	100% (3a)	-
3		1h MW	-	87% (3a)	13% (4a)
4		12h C.H.	-	100% (3a)	-
5	Me	10 min MW	-	100% (3b)	-
6		20 min MW	-	100% (3b)	-
7		1h MW	-	94% (3b)	6% (4b)
8		12h C.H.	-	100% (3b)	-
9		10 min MW	22% (2c)	78% (3c)	-
10		20 min MW	20% (2c)	80% (3c)	-
11		1h MW	13% (2c)	70% (3c)	17% (4c)
12		12h C.H.	24% (2c)	76% (3c)	-
13		10 min MW	41% (2d)	59% (3d)	-
14		20 min MW	26% (2d)	50% (3d)	24% (4d)
15		1h MW	17% (2d)	39% (3d)	44% (4d)
16		12h C.H.	25% (2d)	75% (3d)	-
17		10 min MW	100% (2e)	-	-
18		20 min MW	100% (2e)	-	-
19		1h MW	100% (2e)	-	-
20		12h C.H.	100% (2e)	-	-

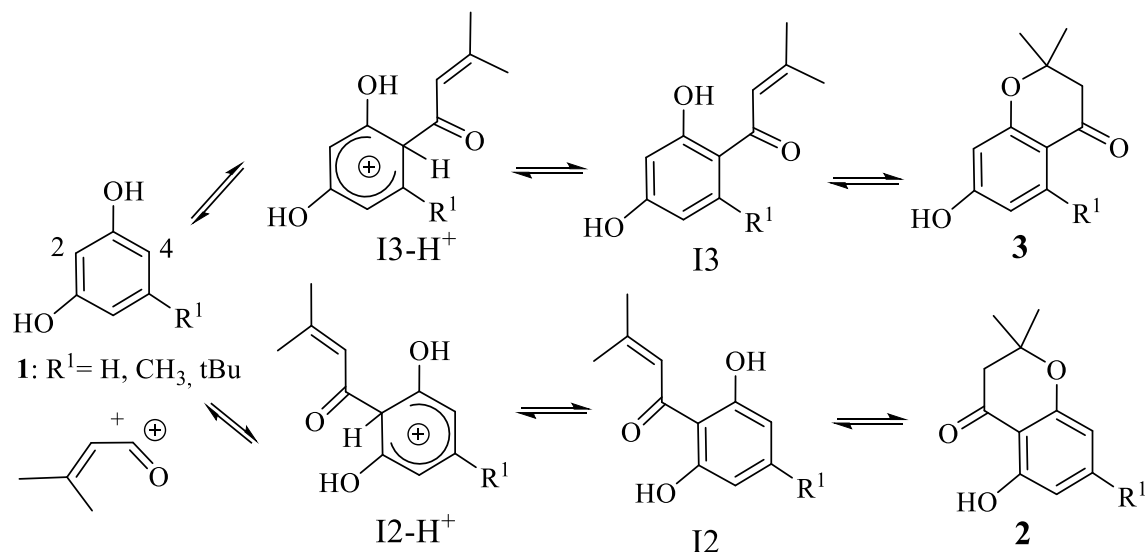
^a MW: microwave irradiation; C.H.: conventional heating

Altering the reaction conditions did not significantly affect the regiochemical outcome. However, by increasing the microwave exposure to 1 hour, diacylated product (**4**) was detected in the reaction mixture except for 1,1-dimethylheptyl derivative that was not observed (Table 1 entry 19). These compounds (**4a**, **4b**, **4c** and **4d**) were identified by HPLC/MS that also allowed the determination of the ratios in the crude reaction. The structures of **4a** and **4d**, which were

isolated by chromatography, were unambiguously determined by MS and NMR experiments. Note that the diacylated product **4** could be formed from both 2,2-dimethyl-5-hydroxychroman-4-one (**2c**, **2d**) and 2,2-dimethyl-7-hydroxychroman-4-one (**3c**, **3d**).

It is interesting to note the difference of reactivity between the microwave-assisted reactions and the reaction carried out by conventional heating. Compounds **3a**, **3b** and **3d** (10 min) formed faster than **4a**, **4b** and **4d** (1h) are the kinetic products of this reaction. Compounds **4a**, **4b** and **4d** obtained in lower amount are thermodynamically more stable than the corresponding compounds **3**. Under conventional heating conditions, the reaction seems to be fully kinetically controlled since there is no formation of **4**.

In an attempt to explain the reactivity of different resorcinols with 3,3-dimethylacrylic acid, theoretical calculations have been performed. Thus, the reactivity of three systems has been considered (Scheme 3); resorcinol, orcinol and 5-*tert*-butyl-1,3-dihydroxybenzene have been selected. Hydrogen and methyl groups in R¹ are the corresponding substituents of resorcinol and orcinol. The *tert*-butyl group in R¹ was chosen to mimic the 1,1-dimethylheptyl moiety.



Scheme 3. Schematic representation of the reaction.

For each of the three starting molecules, two possibilities have been considered: the reaction on the C2 and the one in C4. Two possible transition state (TS) paths have been explored for the Friedel-Crafts acylation depending on the disposition of the attacking carbonyl molecule with respect to the aromatic ring (Figure 2).

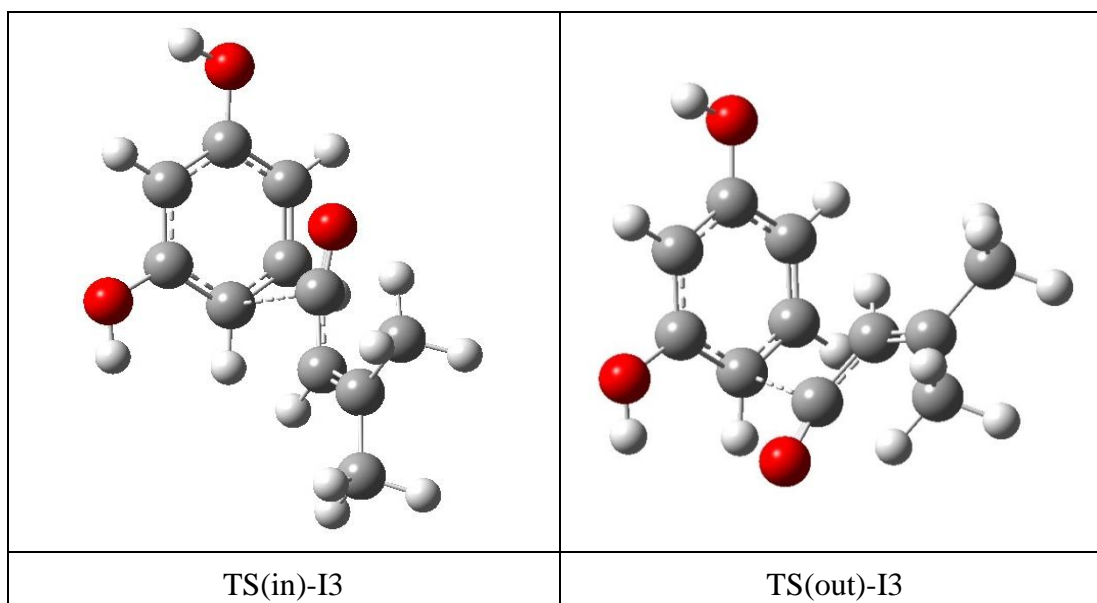


Figure 2. The two TS found for the Friedel-Crafts acylation that yield the formation of I3-H⁺, R¹ = H.

Small values of the barriers are found with respect to the isolated reactants (Table 2). The differences between the most favorable TS that yields I3-H⁺ and I2-H⁺ are small being in all the cases the one that yields I3-H⁺ the smaller. However, the relative energies of the charged species obtained in this first step of the reaction are different depending on the R¹ group. Thus, for R¹ = H and Me, the most stable molecule correspond to the I3-H⁺ while for R¹ = *t*-Bu it is I2-H⁺.

The relative energies of the neutral intermediate molecules I3 and I2, as well as the bicyclic structures **3** and **2** obtained after a subsequent intramolecular Michael addition are gathered in Table 3. Only in the case of X = H is I3 more stable than I2. In the rest of the cases, I2 is more stable than I3 and the same happened between **2** and **3**. In addition, the energetic differences increase with the size of the R¹ group.

Table 2. Relative energy (kJ/mol) of the TS and Friedel-Crafts acylation with respect to the isolated reactants. In parenthesis, the population based on the energies of I3-H⁺ and I2-H⁺ is indicated. Values at B3LYP/6-311++G(d,p) computational level in PCM, ε = 80

	TS(in)-I3	TS(out)-I3	TS(in)-I2	TS(out)-I2	I3-H ⁺	I2-H ⁺
H	0.4	2.3	6.1	4.2	-49.9 (99.9%)	-32.9 (0.1%)
Me	-3.9	-0.1	0.6	-0.9	-53.8 (92 %)	-47.8 (8%)
<i>t</i> -Bu	-6.2	-4.7	-3.7	-5.5	-54.2 (16%)	-58.3 (84%)

Table 3. Relative energies (kJ/mol) of the I3, I2, **3** and **2** molecules

R ¹	I3	I2	3	2
H	0.0	18.8	0.0	-21.0
Me	0.0	-9.1	0.0	-32.2
<i>t</i> -Bu	0.0	-53.9	0.0	-66.3

Based on the experimental and the computational results shown here, the Friedel-Crafts acylation that yields the formation of I3-H⁺ and I2-H⁺ seems to be the one responsible of the percentage of final products obtained. Thus, the calculated relative energies of the I3-H⁺ and I2-H⁺ intermediates are able to explain the experimental results. In other terms, the influence of different 5-alkyl-substituted resorcinols on the formation of 2,2-dimethylchroman-4-ones does not depend on the energetic barrier of the first step involved in Friedel-Crafts acylation. But it is rather the difference of relative energies of the charged species at the transition state that depends on the nature of the alkyl substituent. Based on these theoretical considerations, it has been possible to explain the formation of 2,2-dimethyl-5-hydroxychroman-4-ones and/or 2,2-dimethyl-7-hydroxychroman-4-ones obtained experimentally.

Affinity towards cannabinoid receptors

The affinity of chromanones **2d-e**, **3a-d**, **4a** and **4d** at CB₁ and CB₂ cannabinoid receptors were evaluated using radiolabeled binding assays.

Table 4. Binding affinity of chromanones and the reference cannabinoid WIN55,212-2 for CB₁ and CB₂ cannabinoid receptors

Compd	CB ₁ K _i (μM) ^a	CB ₂ K _i (μM) ^a
2d	> 40	3.44 ± 0.27
2e	1.35 ± 0.38	2.54 ± 0.06
3a	> 40	> 40
3b	> 40	> 40
3c	> 40	> 40
3d	> 40	1.28 ± 0.08
4a	> 40	4.41 ± 0.18
4d	> 40	3.37 ± 0.19
WIN55,212-2 ^b	0.045 ± 0.009	0.037 ± 0.002

^a Values obtained from competition curves using [³H]CP55,940 as radioligand for CB₁ and CB₂ cannabinoid receptors and are expressed as mean SEM values of at least three experiments. ^b

WIN55,212-2: (R)-(+)-[2,3-dihydro-5-methyl-3-(4-morpholinylmethyl)pyrrolo[1,2,3-de]-1,4-benzoxazin-6-yl]-1-naphthalenylmethanone.

The affinity constant K_i was determined measuring the ability of the compound to displace [³H]-CP55,940 (2-[(1R,2R,5R)-5-hydroxy-2-(3-hydroxypropyl)cyclohexyl]-5-(2-methyloctan-2-yl)phenol) from human CB₁ and CB₂ receptors transfected into HEK293 EBNA cells.⁵ Standard cannabinoid ligand WIN55,212-2 was also assessed for comparison with the new compounds. The tested compounds displayed much lower affinity than the reference cannabinoid ligand. Although these affinities are low, these data suggest some relation structure-activity. In the 2,2-dimethyl-7-hydroxychroman-4-one series (**3**), the lipophilic methylenic chain is necessary for CB₂ receptor binding (**3d**). However, this is not the case for the tricyclic scaffold (**4**) for which the moderate CB₂ affinity was independent of the lipophilic nature of the alkyl chain (**4a** and **4d**). 2,2-Dimethyl-5-hydroxychroman-4-one **2d** showed weak but selective affinity for the CB₂ cannabinoid receptors whereas the dimethylheptyl derivative **2e** binds weakly to both receptors.

Conclusions

The influence of different 5-alkyl-substituted resorcinols on the formation of 2,2-dimethylchroman-4-ones does not depend on the energetic barrier of the first step involved in Friedel-Crafts acylation. But it is rather the difference of relative energies of the charged species at the transition state that depends on the nature of the alkyl substituent. Based on these theoretical considerations, it has been possible to explain the formation of 2,2-dimethyl-5-hydroxychroman-4-ones and/or 2,2-dimethyl-7-hydroxychroman-4-ones obtained experimentally. From an experimental point of view, the use of microwave radiation considerably reduced reaction times but it did not increase reaction yields. At longer microwave irradiation, diacetylated products started to form in the reaction mixture. With respect to their biological activity, some of the chromanones described here showed moderate but selective affinity for the CB₂ cannabinoid receptor.

Experimental Section

Chemistry

General. All commercially available reagents and solvents were used as supplied without further purification. Microwave-assisted synthesis was carried out with a Biotage Initiator Microwave (0-400W at 2.45 GHz). Column chromatography was performed using silica gel 60 (230-400 mesh). The purity of final compounds was determined by HPLC-MS and elemental analyses. HPLC-MS analysis was performed on a Waters 2695 HPLC system equipped with a Photodiode Array 2996 coupled to Micromass ZQ 2000 mass spectrometer (ESI-MS), using a reverse-phase

column SunFireTM (C-18, 4.6 x 50 mm, 3.5 μ m) and 5 min gradient A) MeCN/0.08% formic acid, B) H₂O/1% formic acid visualizing at $\lambda = 254$ nm. Elemental analyses were performed using a LECO CHNS-932 apparatus. ¹H and ¹³C NMR spectra were recorded on a Bruker 300 (300 and 75 MHz respectively) at 25°C with CDCl₃ as solvent and TMS as internal standard. Chemical shifts were expressed in ppm (δ) and coupling constants were reported in Hz. Melting points were determined in open capillary tubes with a MP70 Mettler Toledo apparatus. 5-(1,1-Dimethylheptyl)-1,3-dihydroxybenzene (**1e**) was prepared as described previously by some of us.⁵

5-(1-*n*-Butynyl)-1,3-dimethoxybenzene (6). To a solution of the commercially available 1-bromo-3,5-dimethoxybenzene (**5**) (1.0 g, 4.6 mmol) in Et₃N (12 mL) was added CuI (0.05 g, 0.27 mmol), PPh₃ (0.36 g, 1.3 mmol), and Pd₂(dba)₃ (6.12 g, 0.13 mmol). Butyne was bubbled for 6 minutes under N₂ atmosphere. The reaction was stirred at 80 °C overnight. The mixture was cooled to room temperature. The reaction mixture was filtered and the filtrate was evaporated *in vacuo*. The crude residue was further purified by column chromatography (CH₂Cl₂/hexane, 1:3) to give the pure title compound as a red oil (0.80 g, 91%); ¹H NMR (CDCl₃): δ 6.67-6.55 (m, 2H, 4-H and 6-H), 6.39 (m, 1H, 2-H), 3.83 (s, 6H, OCH₃), 2.40 (q, *J* 7.6 Hz, 2H, 3'-H), 1.25 (t, *J* 7.8 Hz, 3H, 4'-H). ¹³C NMR (CDCl₃): $\delta = 160.4$ (1-C and 3-C), 125.3 (5-C), 109.8 (4-C or 6-C), 109.3 (4-C or 6-C), 101.1 (2-C), 91.3 (2'-C), 79.8 (1'-C), 55.3 (OCH₃), 13.8 (3'-C), 13.0 (4'-C); HPLC-MS: [A, 15% - 100%], *t*_R = 5.72 min, (100%); MS (ES⁺, *m/z*) 191 [M+H]⁺; Anal. calcd. for C₁₂H₁₄O₂ (190.24): C, 75.76; H, 7.42. Found: C, 75.89; H, 7.23.

5-*n*-Butyl-1,3-dimethoxybenzene (7). A solution of 5-(1-*n*-butynyl)-1,3-dimethoxybenzene (**6**) (0.74 g, 3.8 mmol) in CH₂Cl₂ (15 mL) was degassed and flushed with N₂ three times. Then 2 mg 10% Pd/C was added and the solution was degassed again with nitrogen. The reaction mixture was hydrogenated at 45 psi at room temperature for 4 h. The mixture was filtered over Celite and the filtrate was concentrated to afford essentially pure product as yellow oil²³ (0.75 g, 98%); ¹H NMR (CDCl₃): $\delta = 6.50$ -6.42 (m, 2H, 4-H and 6-H), 6.31 (m, 1H, 2-H), 3.72 (s, 6H, OCH₃), 2.63 (t, *J* 7.1 Hz, 2H, 1'-H), 1.54 (m, 2H, 2'-H), 1.32 (m, 2H, 3'-H), 1.16 (t, *J* 7.9 Hz, 3H, 4'-H). ¹³C NMR (CDCl₃): $\delta = 160.6$ (1-C and 3-C), 145.4 (5-C), 107.3 (4-C and 6-C), 97.5 (2-C), 56.1 (OCH₃), 35.9 (1'-C), 33.4 (2'-C), 22.3 (3'-C), 13.9 (4'-C); HPLC-MS: [A, 15% - 100%], *t*_R = 5.97 min, (96%); MS (ES⁺, *m/z*) 195 [M+H]⁺

5-*n*-Butyl-1,3-dihydroxybenzene (1c). To a solution of 5-*n*-butyl-1,3-dimethoxybenzene (**7**) (0.63 g, 3.7 mmol) in dry CH₂Cl₂ was added dropwise boron tribromide (1M in CH₂Cl₂, 4.4 mL, 27 mmol) at 0 °C under N₂ atmosphere. The reaction mixture was allowed to warm to room temperature and stirred for 20 h. Then MeOH was carefully added at 0 °C until neutralization. The solvent was removed *in vacuo* and the crude mixture was purified by chromatography on silica gel (EtOAc/hexane, 1:2) to give the title compound as a brown solid (0.41 g, 65%), mp: 80-81 °C (lit.¹⁰ mp 81.5-82.5 °C); ¹H NMR (CDCl₃): $\delta = 6.26$ (m, 2H, 4-H and 6-H), 6.20 (d, *J* = 1.8 Hz, 1H, 2-H), 5.83 (s, 2H, OH), 2.50 (t, *J* 7.5 Hz, 2H, 1'-H), 1.57-1.47 (m, 2H, 2'-H), 1.37-1.26 (m, 2H, 3'-H), 0.90 (t, 3H, 4'-H). ¹³C NMR (CDCl₃): $\delta = 146.5$ (5-C), 157.5 (1-C and 3-C),

109.0 (4-C and 6-C), 100.1 (2-C), 36.5 (1'-C), 34.1 (2'-C), 23.2 (3'-C), 14.5 (4'-C); HPLC-MS: [A, 15% - 100%], t_{R} = 4.13 min, (97%); MS (ES⁺, m/z) 167 [M+H]⁺; Anal. calcd. for C₁₅H₂₄O₂ (166.22): C, 72.26; H, 8.49. Found: C, 72.44; H, 8.27.

General procedures for the synthesis of 5- and 7-hydroxy-2,2-dimethyl-4-chromanones.

A solution of the appropriate resorcinol **1** in methanesulfonic acid and 3,3-dimethylacrylic acid were added to a suspension of P₂O₅ in methanesulfonic acid under N₂ atmosphere. The reaction mixture was heated following three different procedures: a) irradiation by microwave at 70 °C in a sealed reactor for 10 minutes; b) irradiation by microwave at 70 °C in a sealed reactor for 20 minutes; c) irradiation by microwave at 70 °C in a sealed reactor for 1 hour; d) conventional heating at 70 °C for about 12 hours. The reaction mixture was poured onto water/ice and then extracted with CH₂Cl₂. The organic layer was dried over anhydrous MgSO₄. Solvent was removed under reduced pressure. The crude compounds were purified and isolated by chromatography on silica gel (hexane/EtOAc).

7-Hydroxy-2,2-dimethylchroman-4-one (3a) and **2,2,8,8-tetramethyl-2,3,8,9-tetrahydro-4H,10H-pyrano[2,3-f]chromene-4,10-dione (4a)**. Chromanone **3a** and compound **4a** were prepared from resorcinol (**1a**) (0.4 g, 3.60 mmol) using P₂O₅ (0.76 g, 5.4 mmol), methanesulfonic acid (5 mL, 77.08 mmol), and 3,3-dimethylacrylic acid (0.72 g, 7.26 mmol). Following procedures a, b, and d, **3a** was isolated [b: yield = 98%; d: 89%]. Following procedure c **3a** [c: yield = 64%] and **4a** [c: yield = 9%] were isolated.

7-Hydroxy-2,2-dimethylchroman-4-one (3a). White solid, mp 171-173 °C (lit.¹¹ mp 172 °C). ¹H NMR (CDCl₃): δ = 7.95 (d, J 7.1 Hz, 1H, 5-H), 6.54 (dd, J 1.5 Hz, 1H, 8-H), 6.36 (dd, J 7.1 Hz, J 1.5 Hz, 1H, 6-H), 2.67 (s, 2H, 3-H), 1.46 (s, 6H, OC(CH₃)₂). ¹³C NMR (CDCl₃): δ = 191.6 (4-C), 163.3 (7-C), 162.1 (8a-C), 128.8 (5-C), 114.1 (4a-C), 109.8 (6-C), 103.6 (8-C), 79.5 (2-C), 48.5 (3-C), 26.6 (OC(CH₃)₂); HPLC-MS: [A, 40% - 100%], t_{R} = 1.50 min, (93%); MS (ES⁺, m/z) 193 [M+H]⁺; Anal. calcd. for C₁₁H₁₂O₃ (192.21): C 68.74, H 6.29, found: C 67.96, H 6.52.

2,2,8,8-Tetramethyl-2,3,8,9-tetrahydro-4H,10H-pyrano[2,3-f]chromene-4,10-dione (4a). Yellow solid, mp 160-162 °C (lit.¹² mp 160-161 °C); ¹H NMR (CDCl₃): δ = 7.85 (d, J = 7.6 Hz, 1H, 5-H), 6.87 (d, J 7.6 Hz, 1H, 6-H), 2.66 (s, 2H, 9-H), 2.71 (s, 2H, 3-H), 1.52 (s, 6H, 11-H), 1.48 (s, 6H, 12-H). ¹³C NMR (CDCl₃): δ = 196.6 (10-C), 190.3 (4-C), 161.4 (6a-C), 157.1 (10b-C), 141.2 (10a-C), 134.8 (5-C), 111.1 (4a-C), 108.9 (6-C), 87.8 (8-C), 85.7 (2-C), 48.9 (9-C), 47.6 (3-C), 25.1 (11-C), 26.0 (12-C); HPLC-MS: [A, 40% - 100%], t_{R} = 3.71 min, (92%); MS (ES⁺, m/z) 275 [M+H]⁺; Anal. calcd. for C₁₆H₁₈O₄ (274.31): C 70.06, H 6.61, found: C 70.41, H 6.58.

7-Hydroxy-2,2,5-trimethylchroman-4-one (3b). Chromanone **3b** was prepared from orcinol (**1b**) (0.10 g, 0.81 mmol) using P₂O₅ (0.17 g, 1.21 mmol), methanesulfonic acid (3 mL, 46.25 mmol), and 3,3-dimethylacrylic acid (0.12 g, 1.20 mmol) following procedures a, b [yield = 70%], c [yield = 53%], and d [yield = 72%]. Using procedure c **4b**¹² was detected by HPLC/MS but not isolated. **3b**: Dark-orange oil (lit.¹³ mp = 190 °C). ¹H NMR (CDCl₃): δ = 6.73 (d, J 1.7 Hz, 1H, 6-H), 6.39 (d, J 1.7 Hz, 1H, 8-H), 2.87 (s, 2H, 3-H), 2.35 (s, 3H, 1'-H), 1.48 (s, 6H, OC(CH₃)₂). ¹³C NMR (CDCl₃): δ = 191.6 (4-C), 167.4 (7-C), 162.6 (8a-C), 151.0 (5-C), 116.1

(4a-C), 109.9 (6-C), 99.6 (8-C), 82.3 (2), 49.8 (3-C), 27.1 (OC(CH₃)₂); 21.5 (1'-C); HPLC-MS: [A, 10% - 100%], t_R = 4.10 min, (90%); MS (ES⁺, *m/z*) 207 [M+H]⁺; Anal. calcd. for C₁₂H₁₄O₃ (206.24): C 69.88, H 6.84, found: C 70.11, H 6.76.

7-*n*-Butyl-5-hydroxy-2,2-dimethylchroman-4-one (2c) and 5-*n*-butyl-7-hydroxy-2,2-dimethylchroman-4-one (3c). Chromanones **2c** and **3c** were prepared from 5-*n*-butyl-1,3-dihydroxybenzene (**1c**) (0.06 g, 0.36 mmol) using P₂O₅ (0.06 g, 0.43 mmol), methanesulfonic acid (2 mL, 30.83 mmol), and 3,3-dimethylacrylic acid (0.054 g, 0.54 mmol) by following procedures a, b [**3c**: yield = 20%; **2c**: yield = 12%], c [**3c**: yield = 54%; **2c**: yield = 4%] and d. Using procedure c **4c** was detected by HPLC/MS but not isolated.

7-*n*-Butyl-5-hydroxy-2,2-dimethylchroman-4-one (2c). Yellow oil²⁴; ¹H NMR (CDCl₃): δ = 11.53 (s, 1H, OH), 6.28 (d, *J* 1.5 Hz, 1H, 6-H), 6.19 (d, *J* 1.5 Hz, 1H, 8-H), 2.65 (s, 2H, 3-H), 2.45 (t, *J* 7.5 Hz, 2H, 1'-H), 1.55 (m, 2H, 2'-H), 1.40 (s, 6H, OC(CH₃)₂), 1.25 (m, 2H, 3'-H), 0.85 (t, *J* 7.2 Hz, 3H, 4'-H). ¹³C NMR (CDCl₃): δ = 198.0 (4-C), 162.5 (8a-C), 160.4 (5-C), 155.5 (7-C), 109.9 (6-C), 108.4 (8-C), 105.7 (4a-C), 79.0 (2-C), 49.0 (3-C), 36.9 (1'-C), 33.0 (2'-C), 22.5 (3'-C), 27.0 (OC(CH₃)₂), 14.0 (4'-C); HPLC-MS: [A, 80% - 95%], t_R = 2.17 min, (99%); MS (ES⁺, *m/z*) 249 [M+H]⁺

5-*n*-Butyl-7-hydroxy-2,2-dimethylchroman-4-one (3c). Yellow solid²⁴, mp 130-131 °C; ¹H NMR (CDCl₃): δ = 6.27 (d, *J* 2.5 Hz, 1H, 6-H), 6.21 (d, *J* 2.5 Hz, 1H, 8-H), 5.39 (s, 1H, OH), 2.90 (t, *J* 7.5 Hz, 2H, 1'-H), 2.62 (s, 2H, 3-H), 1.55-1.45 (m, 4H, 2', 3'-H), 1.40 (s, 6H, OC(CH₃)₂), 0.95 (t, *J* 7.2 Hz, 3H, 4'-H). ¹³C NMR (CDCl₃): δ = 192.0 (4-C), 163.5 (8a-C), 162.5 (7-C), 150.0 (5-C), 112.7 (4a-C), 112.0 (6-C), 102.4 (8-C), 78.8 (2-C), 50.5 (3-C), 35.8 (1'-C), 34.0 (2'-C), 26.9 (OC(CH₃)₂), 23.5 (3'-C), 14.1 (4'-C); HPLC-MS: [A, 80% - 95%], t_R = 0.94 min, (95%); MS (ES⁺, *m/z*) 249 [M+H]⁺; Anal. calcd. for C₁₅H₂₀O₃ (248.34): C 72.55, H 8.12, found: C 72.61, H 8.22.

5-Hydroxy-2,2-dimethyl-7-*n*-pentylchroman-4-one (2d), 7-hydroxy-2,2-dimethyl-5-*n*-pentylchroman-4-one (3d) and 2,2,8,8-tetramethyl-5-*n*-pentyl-2,3,8,9-tetrahydro-4H,10H-pyrano[2,3-*f*]chromene-4,10-dione (4d). Chromanones **2d**, **3d** and compound **4d** were prepared from olivetol (**1d**) (0.42 g, 2.37 mmol) using P₂O₅ (0.40 g, 2.80 mmol), methanesulfonic acid (5 mL, 77.08 mmol), and 3,3-dimethylacrylic acid (0.35 g, 3.50 mmol). Following procedures a and d **2d** and **3d** were isolated [**3d**: yield = 50% (d); **2d**: yield = 11% (d)]. Following procedures b and c **2d**, **3d** and **4d** were isolated [**3d**: yield = 37% (b), 50% (c); **2d**: yield = 15% (b), 5% (c); **4d**: yield = 7% (b), 10% (c)].

5-Hydroxy-2,2-dimethyl-7-*n*-pentylchroman-4-one (2d). Orange oil (lit.⁴ mp 23-27 °C); ¹H NMR (CDCl₃): δ = 6.32 (d, *J* 2.4 Hz, 1H, 8-H), 6.26 (d, *J* 2.4 Hz, 1H, 6-H), 2.98 (t, *J* 7.1 Hz, 2H, 1'-H), 2.66 (s, 2H, 3-H), 1.48-1.53 (m, 2H, 2'-H), 1.44-1.38 (m, 2H, 3'-H), 1.41 (s, 6H, OC(CH₃)₂), 1.30-1.40 (m, 2H, 4'-H), 0.87 (t, *J* 6.9 Hz, 3H, 5'-H). ¹³C NMR (CDCl₃): δ = 191.2 (4-C), 161.9 (5-C), 159.7 (8a-C), 147.6 (7-C), 110.2 (8-C), 109.9 (6-C), 100.1 (4a-C), 76.4 (2-C), 48.3 (3-C), 33.3 (1'-C), 30.1 (3'-C), 28.1 (2'-C), 24.8 (OC(CH₃)₂), 20.7 (4'-C), 12.2 (5'-C); HPLC-MS: [A, 80% - 100%], t_R = 2.78 min, (98%); MS (ES⁺, *m/z*) 263 [M+H]⁺; Anal. calcd. for C₁₆H₂₂O₃ (262.34): C 73.25, H 8.45, found: C 72.98, H 8.71 %.

7-Hydroxy-2,2-dimethyl-5-*n*-pentylchroman-4-one (3d). Pale yellow solid, mp 92-94 °C (mp lit.¹⁴ = 91-93 °C); ¹H NMR (CDCl₃): δ = 8.25 (s, 1H, OH), 6.24 (d, *J* 2.1 Hz, 1H, 6-H), 6.16 (d, *J* 2.1 Hz 1H, 8-H), 2.95 (s, 2H, 3-H), 2.60 (t, *J* 7.1 Hz, 2H, 1'-H), 1.68 (m, 2H, 2'-H), 1.41 (s, 6H, OC(CH₃)₂), 1.33-1.26 (m, 2H, 3', 4'-H), 0.80 (t, *J* 7.0 Hz, 3H, 5'-H). ¹³C NMR (CDCl₃): δ = 192.9 (4-C), 163.9 (8a-C), 162.0 (7-C), 149.9 (5-C), 112.7 (4a-C), 111.9 (6-C), 102.3 (8-C), 78.7 (2-C), 50.6 (3-C), 35.6 (2'-C), 32.4 (1'-C), 30.9 (3'-C), 29.1 (4'-C), 26.9 (OC(CH₃)₂), 14.5 (5'-C); HPLC-MS: [A, 80% - 100%], t_R = 1.23 min, (96%); MS (ES⁺, *m/z*) 263 [M+H]⁺; Anal. calcd. for C₁₆H₂₂O₃ (262.34): C 73.25, H 8.45, found: C 72.83, H 8.69 %.

2,2,8,8-Tetramethyl-5-*n*-pentyl-2,3,8,9-tetrahydro-4*H*,10*H*-pyrano[2,3-*f*]chromene-4,10-dione (4d). Yellow gummy solid; ¹H NMR (CDCl₃): δ = 6.30 (s, 1H, 6-H), 2.91 (t, *J* 7.1 Hz, 2H, 1'-H), 2.63 (s, 2H, 9-H), 2.61 (s, 2H, 3-H), 1.64 (m, 2H, 2'-H), 1.50 (s, 6H, 11-H), 1.45 (s, 6H, 12-H), 1.31-1.20 (m, 4H, 3', 4'-H), 0.88 (t, *J* 8.0 Hz, 3H, 5'-H). ¹³C NMR (CDCl₃): δ = 192.1 (4-C), 189.6 (10-C), 165.1 (6a-C), 163.3 (10b-C), 150.9 (5-C), 135.7 (10a-C), 117.0 (4a-C), 109.1 (6-C), 85.2 (8-C), 79.1 (2-C), 49.1 (9-C), 42.4 (3-C), 31.9 (1'-C), 30.9 (2'-C), 29.8 (3'-C), 24.3 (4'-C), 26.7 (11-C), 25.8 (12-C), 14.6 (5'-C); HPLC-MS: [A, 60% - 100%], t_R = 4.33 min, (94%); MS (ES⁺, *m/z*) 345 [M+H]⁺; Anal. calcd. for C₂₁H₂₈O₄ (344.44): C 73.23, H 8.19, found: C 72.86, H 8.35 %.

7-(1,1-Dimethylheptyl)-5-hydroxy-2,2-dimethylchroman-4-one (2e). Chromanone **2e** was prepared from 5-(1,1-dimethylheptyl)-1,3-dihydroxybenzene (**1e**) (0.10 g, 0.43 mmol) using P₂O₅ (0.08 g, 0.63 mmol), methanesulfonic acid (3 mL, 46.25 mmol), and 3,3-dimethylacrylic acid (0.06 g, 0.63 mmol) by following procedures a (yield = 60 %), b (yield = 52 %), c (yield = 44%) and d (yield = 81%). Pale-yellow oil⁵; ¹H NMR (CDCl₃): δ = 6.45 (d, *J* = 1.6 Hz, 1H, 8-H), 6.37 (d, *J* = 1.6 Hz, 1H, 6-H), 2.71 (s, 2H, 3-H), 1.52-1.59 (m, 2H, 2'-H), 1.46 (s, 6H, OC(CH₃)₂), 1.22 (s, 6H, C(CH₃)₂), 1.19 (bs, 6H, 3'-H, 4'-H, 5'-H), 1.05 (bs, 2H, 6'-H), 0.87 (m, 3H, 7'-H). ¹³C NMR (CDCl₃): δ = 197.9 (4-C), 163.0 (5-C), 161.7 (8a-C), 156.0 (7-C), 107.0 (8-C), 106.2 (6-C), 105.7 (4-C), 79.1 (2-C), 48.5 (3-C), 41.0 (2'-C), 39.2 (1'-C), 32.1, 27.1 and 23.0 (3'-C, 4'-C and 5'-C), 30.3 (C(CH₃)₂), 28.8 (OC(CH₃)₂), 25.0 (6'-C), 14.5 (7'-C); HPLC-MS: [A, 80% - 100%], t_R = 4.9 min, (97%); MS (ES⁺, *m/z*) 318 [M+H]⁺; Anal. calcd. for C₂₀H₃₀O₃ (318.45): C, 75.43; H, 9.50. Found: C, 75.52; H 9.64 %.

Computational method

The geometry of the systems has been optimized with the B3LYP/6-31G(d) computational level^{15,16} within the Gaussian-09 program.¹⁷ In order to include the effect of the solvent, methanesulfonic acid, the Polarizable Continuum Method (PCM) model¹⁸ has been considered with a dielectric constant of 80 which is an intermediate value of the ones described in the literature for this solvent.¹⁹ Frequency calculations have been carried out to confirm that the structures obtained correspond to energy minima or true transition states (TS).

The NMR theoretical chemical shielding of the synthesized molecules has been calculated using the GIAO method²⁰ at the B3LYP/6-311++G(d,p) computational level using the geometries optimized at the same level. The effect of the solvent used in the experimental NMR

measurements (CDCI₃) has been simulated using the PCM model of this solvent. The calculated absolute chemical shieldings have been transformed into chemical shifts using empirical relationships previously described.^{21,22}

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