

A practical synthesis of (3-(phenanthren-9-yl)-4,5-dihydroisoxazol-5-yl)methyl (tert-butoxycarbonyl)-L-alaninate

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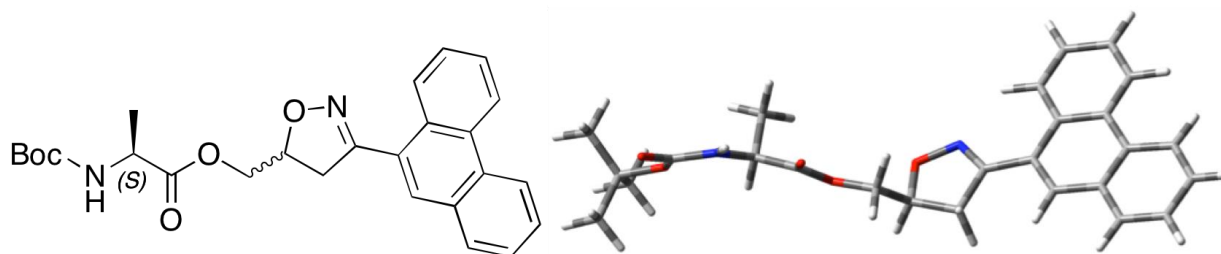
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Abstract

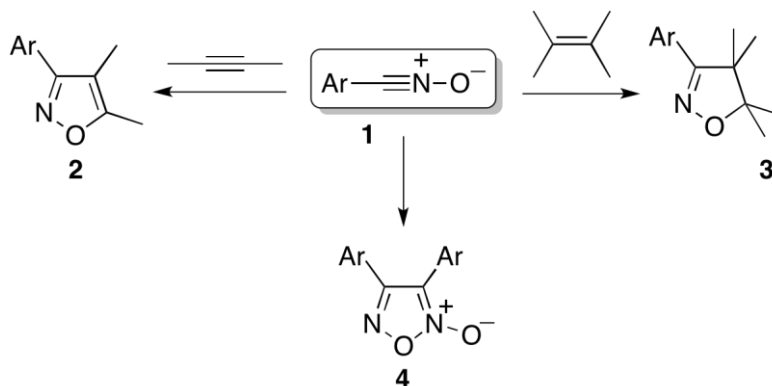
Nitrile oxides are highly reactive, but often unstable, compounds that can be used to substitute for other 1,3-dipoles in [3+2] cycloaddition reactions. Bioorthogonal chemistry reactions that can occur within living systems without interfering with their native biochemical processes, is an area in which stable nitrile oxides can find useful and valuable applications. The rapid and selective synthesis of a phenanthrene-isoxazolyl derivative containing a protected amino acidic residue, through a facile 1,3-dipolar cycloaddition of a stable phenanthrenenitrile oxide to a *N*-Boc-protected (*S*)-alanine allyl ester as the dipolarophile is described. The synthetic strategy and the fluorescence properties of the single cycloadduct are presented along with the corresponding analytical and spectroscopic data.



Keywords: Stable nitrile oxides, phenanthrenenitrile oxide, 1,3-dipolar cycloaddition reactions, L-alaninate, bioorthogonal chemistry, chemical ligation, fluorescent probe

Introduction

Nitrile oxides (**1**) are highly reactive 1,3-dipoles, however, because of their instability, their use in organic syntheses is seriously overlooked other than for the mere preparation of O,N-containing heterocycles such as isoxazoles (**2**) and isoxazolines (**3**) (Scheme 1). At ordinary temperatures, the dimerization rate is extremely high with simple aliphatic and aromatic nitrile oxides, and the corresponding furoxans are normally obtained quantitatively.¹



Scheme 1. Synthesis of isoxazoles (**2**) and isoxazolines (**3**) from nitrile oxide (**1**) and dimerization to furoxan (**4**).

The synthetic simplicity of nitrile oxide precursors, such as oximes, however, starting from the large aldehyde family, has prompted several studies directed towards improving the stability of these 1,3-dipoles.² Luckily, it was found that the dimerization process could be sterically blocked by bulky substituents in the residue attached to the nitrile-oxide moiety.¹⁻³ Recently, the dimerization process was thoroughly investigated through discrete Fourier transform (DFT) calculations by Houk and co-workers, demonstrating that the rate-determining step is the formation of the C-C bond in a diradical stepwise process. The retardation of dimerization in aromatic nitrile oxides arises from interruption of conjugation between the aromatic substituents and the nitrile oxide group.⁴ The reduced conformation freedom in sterically encumbered aliphatic nitrile oxides presumably increases the activation energy towards the C-C bond formation in the same diradical structures.

These findings were somewhat ignored for a long time although they could be of great benefit for several studies where nitrile oxides can nicely replace other 1,3-dipoles in [3+2] cycloaddition reactions. Bioorthogonal chemistry is one of these areas in which stable nitrile oxides can find useful and valuable applications. The expression “bioorthogonal chemistry” refers to any chemical reaction occurring inside of living systems without interfering with native biochemical processes (Figure 1).⁵

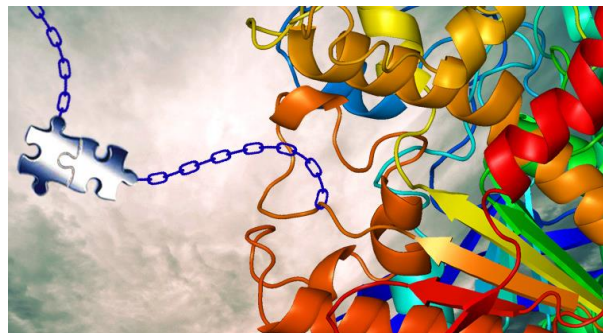


Figure 1. A pictorial concept of bioorthogonal chemistry in living systems.

The chemical nature of bioorthogonal chemistry has enabled the study of glycans, proteins, and lipids in living systems. Chemical reagents must not alter the structure of the substrate dramatically to avoid adversely affecting its bioactivity. Taking into account the chemical data, a reaction is considered to be bioorthogonal if it is selective, biologically and chemically inert, kinetically favorable and biocompatible.⁶ The classical methodologies to perform a reliable chemical ligation are the well-known Staudinger ligation,⁷ the Click Reactions (such as the Huisgen 1,3-dipolar cycloaddition of azides),⁸ as well as the Diels-Alder (DA) reactions of suitably designed diene systems.⁹

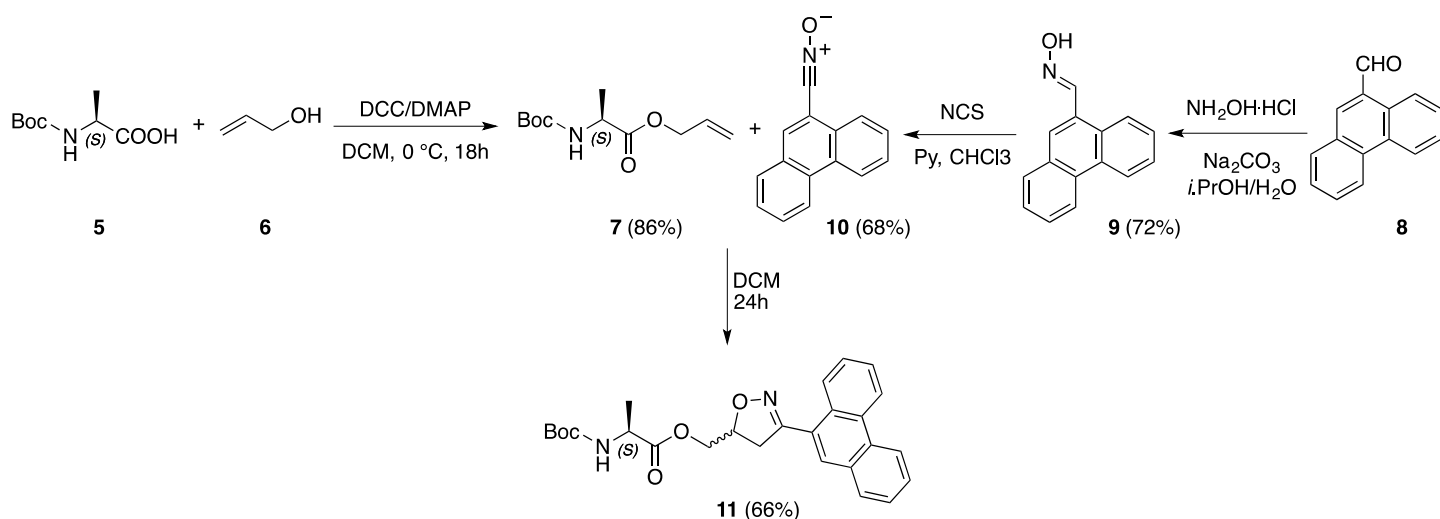
Exploiting the concept of bioorthogonal chemistry, the use of organic molecules with a reduced molecular weight as chemical probes allows a new and interesting chemical design, and represents an outbreking task in modern organic chemistry research.¹⁰ Indeed these molecular probes can interact with specific targets *in vivo* in order to obtain a greater amount of information in complex biological systems. Typically, a general chemical probe contains a fluorescent tag for imaging analysis.¹¹

Aiming novel synthetic approaches to Activity-Based Protein Profiling (ABPP),¹² we describe the rapid and selective synthesis of a phenanthrene-isoxazolyl derivative containing a protected amino acidic residue, based upon a fast and reliable synthesis of a stable phenanthrenenitrile oxide.

Results and Discussion

The *N*-Boc-protected amino acids are commercially available and can be easily esterified by inserting allylic moieties. This type of unsaturation is prone to become a dipolarophile in 1,3-dipolar cycloaddition reactions. In particular, it is known that mono-substituted alkenes are the most reactive dipolarophiles and, thus, the most suitable compounds for this type of investigations.¹³

A typical esterification reaction was performed through standard DCC/DMAP coupling between the *N*-Boc-protected (*S*)-Alanine (**5**) and allylic alcohol (**6**), affording the desired ester (*S*)-**7** (Scheme 2). The white-solid ester (**7**) was isolated through column chromatography and obtained in 86 % yield. It was found to be identical to previously synthesized compounds.^{14,15}



Scheme 2. Synthetic pathways towards the (3-(phenanthren-9-yl)-4,5-dihydroisoxazol-5-yl)methyl (*tert*-butoxycarbonyl)-*L*-alaninate (**11**).

The purified allyl-amino acid (**7**) was submitted to 1,3-dipolar cycloaddition with a slight excess (1.2 equiv.) of freshly prepared phenanthrenitrile oxide (**10**) in dichloromethane (DCM) solution.¹ The stable nitrile oxide (**10**) was prepared starting from the phenanthrene 9-carbaldehyde (**8**) through conversion to the corresponding oxime (**9**) by standard hydroxylamine addition in hydro-alcoholic solution in the presence of a carbonate, followed by *in situ* chlorination with NCS and simultaneous dehydrochlorination with pyridine.¹⁶ Both steps gave the desired compounds in very good yields, and the generation of the nitrile oxide was confirmed by the IR spectroscopy where the $\text{C}\equiv\text{N}-\text{O}$ band was found at 2289 cm^{-1} .

The cycloaddition reaction was conducted at room temperature for 24 h. Chromatographic separation of the worked-up reaction mixture afforded the expected single regioisomer **11**, in 66 % yield, as an inseparable mixture of diastereoisomers.

The structure of the cycloadduct **11** was assigned through the corresponding analytical and spectroscopic data. From the IR spectrum, the presence of the NH of the Boc-protected amino acidic residue is shown by a band centered at 3352 cm^{-1} ; the C=O groups are shown by the strong and intense bands at 1747 and 1714 cm^{-1} , respectively. The ¹H-NMR spectrum (*d*₆-DMSO) shows the presence of the phenanthrene ring in the aromatic region with signals ranging from $\delta 7.78$ ppm to 8.99 ppm. The presence of the newly formed isoxazoline ring is confirmed by the presence of a broad signal at $\delta 5.02$ ppm relative to the CH-O of the heterocyclic ring, coupled with the exocyclic methylene group, the signals of which are found at $\delta 3.54$ and 3.86 ppm (AB system). Another AB system at $\delta 4.36$ and 4.42 ppm corresponds to the isoxazoline methylene. The alanine moiety gave the expected signals of the methyl group at $\delta 1.02$ ppm as a doublet coupled with the CH found at $\delta 4.20$ ppm as a typical quartet. The NH of the amino acid was also found at $\delta 7.39$ ppm as a triplet and exchanges with D₂O. Most of the signals are slightly split, denoting the diastereoisomeric mixture. This fact is clearly shown in the ¹³C-NMR spectrum. The mixture of diastereoisomers is observed to be in a ratio of approximately 1:1, and is due to the undefined chirality of the C5 isoxazoline carbon atom.

The steric effects produced by the phenanthrene moiety of the 1,3-dipole can be seen at work in determining the exclusive regioselectivity of the reaction. Upon investigation of the composition of the crude reaction mixture by NMR techniques, no other compounds were identified in addition to the isolated cycloadduct **11**, confirming the specific regiochemical outcome.

The fluorescence properties given off by the phenanthrene residue to the cycloadduct **11** were determined by recording a fluorescence spectrum in methanol solvent. The emission maximum is centered at 470 nm (Figure 2) and is displayed in the cycloadduct only.

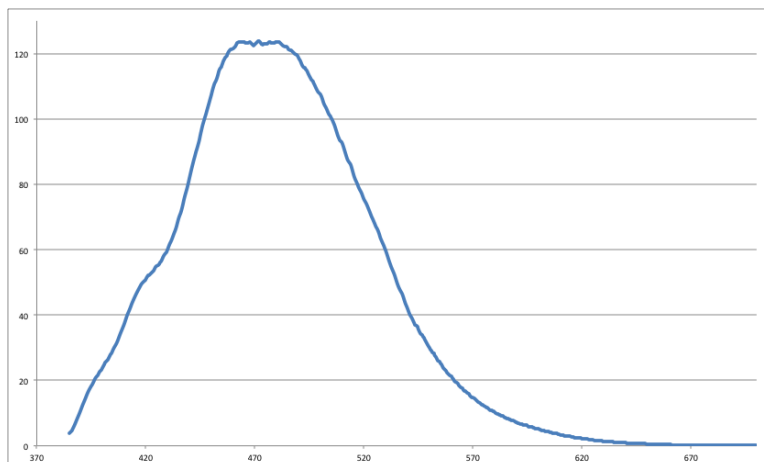


Figure 2. Fluorescence spectrum of the cycloadduct **11** in methanol as solvent.

Conclusions

In conclusion, the present protocol represents a valuable, reliable, robust and innovative methodology for the synthesis of fluorescence chemical probes for ligation chemistry. The synthesis of the fluorescent cycloadduct of type **11** can be easily performed by the simple and straightforward preparation of allylic-substituted amino acids, followed by 1,3-dipolar cycloaddition reactions with stable nitrile oxides. Having in hand a variety of aldehydes, the corresponding oximes offer a wide range of nitrile-oxide precursors that, if displaying fluorescence properties,¹⁷ represent an interesting and promising application for the synthesis of chemical probes for performing ligation studies in living systems without interfering with natural biochemical processes.¹⁸

Experimental Section

General. All melting points (m.p.) are uncorrected. Elemental analyses were done on an elemental analyzer available at the department. ¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded on a Bruker AVANCE 300 spectrometer (solvents specified). Chemical shifts (δ) are expressed in ppm from internal tetramethylsilane and coupling constants (*J*) are in Hertz (Hz). IR spectra (nujol mulls) were recorded on a Perkin Elmer RX-1 spectrophotometer available at the Department; absorptions (ν) are in cm^{-1} . The fluorescence spectrum was recorded in methanol solution on a Perkin Elmer LS-55. Column chromatography was run on Kieselgel 60, 70-230 mesh (Merck); eluent cyclohexane/ethyl acetate 9:1 to 1:1).

Synthesis of the phenanthrenenitrile oxide (**10**)

Phenanthrene oxime (**9**) (0.82 g, 3.7 mmol) was dissolved in 70 mL chloroform and 0.4 mL of freshly distilled pyridine was added. The mixture was then cooled down to 0 °C and an excess of N-chlorosuccinimide (NCS)

(0.57 g, 4.3 mmol) was added portion-wise under stirring over half an hour. The reaction reached completion in a couple of hours. The organic phase was washed with water and dried over anhydrous Na₂SO₄. Evaporation of the solvent left an oily residue that was taken up with diisopropyl ether to crystallize the product. Collection by filtration afforded the nitrile oxide (**10**) in 68 % yield, mp 176-180 °C; IR $\nu_{\text{C}\equiv\text{N-O}}$ 2289 cm⁻¹. ¹H NMR (DMSO-*d*₆) δ 7.80 (m, 5H, arom.), 8.12 (m, 2H, arom.), 8.92 (m, 2H, arom.). ¹³C NMR (DMSO-*d*₆) δ 123.3, 123.9, 124.1, 125.8, 127.8, 128.2, 128.7, 129.4, 129.6, 129.8, 130.1, 130.4, 145.4, 149.2, 179.7. *Anal.* Calcd. for C₁₅H₉NO: C, 82.18; H, 4.14; N, 6.39. Found: C, 82.20; H, 4.13; N, 6.38.

Synthesis of (3-(phenanthren-9-yl)-4,5-dihydroisoxazol-5-yl)methyl (tert-butoxycarbonyl)-L-alaninate (**11**).

To a solution of 204 mg (0.93 mmol) of phenantrene nitrile oxide **10** in 50 mL of DCM, 300 mg (1.33 mmol) of *N*-Boc protected alanine allyl ester **7** were added portion-wise under stirring at room temperature and the reaction was conducted overnight. After evaporation of the solvent at reduced pressure, the residue was submitted to column chromatography to isolate the final product **11** as a yellow solid in 66 % yield, mp 65-70 °C from diisopropyl ether/petrol ether; IR ν_{NH} 3352 cm⁻¹; $\nu_{\text{C=O}}$ 1746, 1694 cm⁻¹. ¹H NMR (DMSO-*d*₆) δ 1.04 (d, 3H, *J* 7 Hz, CH₃), 3.54, 3.86 (m, 1H + 1H, CH₂-O), 4.20 (q, 1H, *J* 7 Hz, CH-CH₃), 4.36, 4.42 (dt, 1H + 1H, *J* 13, 9 Hz, CH-CH₂), 5.02 (bs, 1H, CH-O), 7.39 (m, 1H, NH), 7.78 (m, 4H, arom.), 8.08 (1H, arom.), 8.16 (s, 1H, arom.), 8.99 (m, 3H, arom.). ¹³C NMR* (DMSO-*d*₆) δ 16.8 [16.7], 28.1, 49.0, 65.1 [65.0], 77.1 [77.0], 78.1, 122.9, 123.4, 124.8, 127.2, 127.3, 127.4, 128.4, 129.3, 130.0, 130.2, 130.3, 130.4, 155.2, 157.2. *Anal.* Calcd. for C₂₆H₂₈N₂O₅: C, 69.63; H, 6.29; N, 6.25. Found: C, 69.60; H, 6.30; N, 6.26.

*Where attributable, the signals of the second diastereoisomer are shown in brackets.

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Supplementary Material

¹H and ¹³C spectra are presented in the supplementary material file associated to this manuscript.

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